

Post-Finasteride Syndrome as an Epigenetic Post-Androgen Deprivation Syndrome: A potential pathological link between Drug-Induced Androgen Receptor Overexpression and Polyglutamine Toxicity

propeciahelp.com

Post-Finasteride Syndrome info & discussion forum

PDF generated April 11, 2020

Table Of Contents

Abstract	3
Post-Finasteride Syndrome, what makes it novel and propeciahelp.com	4
A summary of published research into PFS	11
Finasteride Drug origin, pharmacology, AR structure and androgen action	22
PFS: Manifestation of a Post-Androgen Deprivation Syndrome following exposure to substances with antiandrogenic effects	41
AR CAG Repeats and Spinal and Bulbar Muscular Atrophy	58
AR deregulation as a key pathological driver of PFS?	81
The role of the AR in areas relevant to the sexual dysfunction in PFS	107
The role of the AR in areas relevant to the physiological symptoms of PFS	117
The role of the AR in areas relevant to the neurological and psychological symptoms of PFS	147
Androgen mediated pleiotropy?	165
Current situation is dangerous	168
Research going forward	182
Conclusion	185

Abstract

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/post-androgen-deprivation-syndrome-abstract/>

Post-Finasteride Syndrome as an Epigenetic Post-Androgen Deprivation Syndrome: A potential pathological link between Drug-Induced Androgen Receptor Overexpression and Polyglutamine Toxicity

You can download a PDF Export [here](#).

Post-Finasteride Syndrome (PFS) is a rare and devastating disease encompassing persistent physiological, sexual, and neurological health problems following exposure to a 5alpha reductase inhibitor. The condition comprises a broad and variable clinical spectrum and is responsible for relationship breakdown, disability preventing work, isolation and suicide. Herein, the administrators of the patient support website propeciahelp.com summarise the current published research into PFS, add to the understanding of the condition, and present a mechanistic hypothesis to support further scientific investigation. We argue that PFS cannot be understood with exclusive consideration as to Finasteride and is of unappreciated significance to health and disease. More appropriately considered a Post-Androgen Deprivation Syndrome, patients are increasingly seeking support following exposure to diverse substances capable of anti-androgenic endocrine disruption including 5alpha reductase inhibitors, isotretinoin, serotonergic antidepressants, saw palmetto extract and concentrated phenolic compounds marketed as health supplements. A symptomatic and potentially mechanistic overlap between PFS and the polyglutamine disease Spinal and Bulbar Muscular Atrophy is discussed. Transgenic models illustrate that polyQ toxicity can be recapitulated through overexpression of the wild-type AR. Persistent AR overexpression has been established in symptomatic tissue of PFS patients and is a mechanistic consequence of androgen deprivation. We suggest that site-specific epigenetic changes induced by androgen deprivation may result in a pathological AR deregulation. The role of the androgen receptor as a ubiquitous and critical regulator in the physiological and neurological domains relevant to PFS symptomatology is reviewed. We urge clinical education to end psychosomatic misdiagnosis, aid patient management and ensure a genuinely informed consent before prescription of these substances to young men. We urge molecular-level investigation of PFS patients to achieve pathomechanistic understanding, discover safe therapeutic options and ultimately disease-modifying treatment. Discovery of predisposing genetic and epigenetic factors will aid in assessing the suitability of young patients for therapies with antiandrogenic modality, while promising significant translational insight to a range of disease states.

Post-Finasteride Syndrome, what makes it novel and propeciahelp.com

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/post-finasteride-syndrome-and-propeciahelp/>

Post-Finasteride Syndrome

Post-Finasteride Syndrome is a life-altering disease occurring rarely following therapeutic use of a 5-alpha reductase inhibitor such as Finasteride. PFS encompasses serious physical, neurological, and sexual symptoms of variable severity and distribution. Duration of use is not positively correlated with the severity or persistence of the symptoms of PFS, and although the condition can develop rapidly after many years of asymptomatic use, severely affected phenotypes can follow as little as one dose (Garreton et al., 2016; Than et al., 2018). The condition is currently without known predictive factors, disease-modifying treatment or effective therapeutic relief, and thus represents a serious and increasingly urgent unrecognised public health risk as online marketing for finasteride increases.

The diverse symptoms of PFS and their potential severity are not adequately appreciated by clinicians nor in medical literature (Traish, 2018). PFS presents heterogeneously, with variably severe symptoms from a broad constellation, in isolation or combination. Despite the significant interindividual differences in presentation, there are key commonalities in the disease behaviour. The health of the most severely affected patients is so profoundly impacted that they cannot continue their lives in a meaningful capacity. PFS is frequently causative of relationship breakdown, disability preventing work, isolation and suicide. Although of controversial practical application, Maslow's hierarchy of needs is a pervasive categorisation of motivating human needs (Kenrick et al., 2010). PFS, by this measure, can prove ruinous to the attainment of basic physiological needs in sleep and sex, safety needs in emotional security, financial security and health, and the interpersonal needs of friendships, intimacy and family.

Symptoms of PFS

Sexual dysfunction, including:

- Erectile Dysfunction
- Loss of libido
- Ejaculatory and orgasm disorders
- Clear, watery ejaculate
- Genital anaesthesia
- Post-orgasm exacerbation of symptoms

Symptoms of androgen-responsive tissue, including:

- Atrophy of penile tissue and penile deformation
- Venous leak, penile calcification, penile fibrosis
- Penile, testicular, perineal and prostate pain
- Testicular atrophy
- Muscle atrophy
- Muscular dysfunction, fasciculations, tremors
- Dry eyes
- Osteopenia, osteoporosis
- Tooth decay and tooth loss
- Skin pigmentation changes including darkened penile skin
- Thinned skin
- Dry skin
- Acceleration or deceleration of male pattern hair loss

Other physiological changes, including:

- Lessened beard growth and altered pigmentation
- Altered body temperature
- Gynecomastia
- Changes in fat distribution; Increased gynoid and android fat
- Alteration in allergic reactions

Neurological and Cognitive dysfunction, including:

- Depression, Anhedonia

- Memory failure (short term and long term)
- Cognitive impairment
- Anxiety and panic attacks
- Insomnia

Autonomic and sensory nervous system and somatic symptoms, including:

- Sinus arrhythmia, bradycardia, tachycardia
- Sleep apnoea (obstructive, central)
- Visual impairment and problems including visual snow
- Head pressure, vertigo and dizziness
- hearing difficulty and tinnitus
- Digestive impairment including dysmotility, pale stools, diarrhoea and constipation
- Numbness, tingling or stinging/burning sensations, often in distal extremities

Endocrine and metabolic alterations can include:

- Alteration in serum hormonal parameters including T, E, LH
- Deregulation between LH and T
- Low vitamin D3
- Increased triglycerides
- Increased creatine kinase
- Metabolic dysfunction, Insulin resistance, glucose intolerance
- Hyperbilirubinemia
- Decreased 3 α -diol-G
- Lowered CSF neurosteroids

What makes PFS novel?

- Persistent and frequently permanent.
- Interindividually variable improvement or deterioration over course of disease progression.
- Disease ordinarily progresses in absence of the drug: Majority of cases involve rapid progressive onset or intensification of health problems that patients colloquially refer to as a "crash" after

cessation of drug.

- Heterogenous presentation: Differing symptoms and severities across patients with variable site-specific involvement.
- Severity is not positively correlated to length of exposure; severe multisystemically affected phenotypes occur after less than 1mg total.
- Atypical spatiotemporal involvement: More common in younger men taking lower dose for hair loss than older men using for BPH. Associated with greater disability in young men (FAERS data).
- Consequential endocrine fragility: Despite a frequent and curious symptomatic relief, severely affected patients liable to permanent phenotypical worsening following exposure to substances with antiandrogenic properties.

Characteristics inadequately explored in medical literature add significantly to the understanding of the condition and its peculiarity. While PFS is frequently mischaracterised as persistent side effects, in a majority of cases PFS involves the onset or intensification of health problems after cessation of the drug. This counter-intuitive phenomenon, which is often sudden and debilitating, is colloquially referred to as the "crash" by patients. Often, this follows a partial or sometimes even complete resolution of any side effects experienced on the drug. This is a remarkable and intrinsic novel characteristic so frequent that media coverage of the condition expresses awareness of the phenomenon (Morgans, 2018). The majority of PFS patients are younger men who have taken Finasteride 1mg, or a division of the 1mg or 5mg tablets, for treatment of AGA. This is represented in Adverse Drug Reaction (ADR) reports to the FDA FAERS scheme. FAERS data shows a markedly higher number of adverse event reports from this group, coherent with a higher incidence of associated disability (Baas et al., 2018). This is notably atypical in that ADRs are usually more common and severe in aged populations (Lavan & Gallagher, 2015). As finasteride is widely prescribed and PFS is proportionally very rare, there is likely to be a predisposition in consumers who develop PFS. The apparent prevalence in younger individuals, as well as reports of rapid development of the condition upon later rechallenge in previously asymptomatic or mildly symptomatic users further suggests the involvement of spatiotemporal factors, perhaps at the level of gene expression.

PFS is without a consistent biomarker, however patients with prior hormonal bloodwork will often report significant alterations to their serum hormonal profile following onset of the condition, including raised or reduced testosterone. Low luteinising hormone values is commonly reported. Additionally, low vitamin D and signs of metabolic alteration including increased triglycerides and elevated bilirubin can be frequently apparent. Basic urological evaluation may be subclinical or unrevealing, but this is not always the case and clinical evaluation of PFS patients describing a severe or total sexual dysfunction and penile changes who are clinically examined regularly receive relevant diagnoses including penile venous leak, microcalcifications, fibrosis and markers of neuropathy. These outcomes, are not dose dependent, often developing and progressing rapidly after cessation in severely affected patients who took only a single dose. Professor Daniel Stewart, a previously healthy man with no pre-existing mental or sexual dysfunction, developed PFS severely following little over one week of 1mg Finasteride. After cessation due to side effects he experienced the crash, developing sexual dysfunction, genital pain and atrophy,

severe muscle atrophy, weight loss, extreme fatigue, cognitive dysfunction, anxiety and insomnia. Daniel committed suicide at age 37 after suffering for eight months, writing to his family that "Finasteride has destroyed my mind and body". He had received a diagnosis of penile venous leakage (PFS Foundation, 2017)?.

Appreciation of PFS has often entailed a narrow clinical focus, and the reality is alarming drug-induced health problems that extend far beyond erectile dysfunction and depression. Given the diverse array of symptoms and lack of interdependence, it is in our view highly likely many consumers will have developed health problems they have failed to associate with the causative product due to the potential for insidious onset and counter-intuitive presentation after withdrawal. As of 2020, many symptoms are recorded in some capacity in medical literature, however the breadth is only apparent upon comprehensive review. Clinical characterisation of PFS in literature review is often incomplete and can be highly selective in line with the specialisation or hypotheses of the authors. The clear establishment of the multidomain symptom profile is therefore of paramount importance in line with increasing commentaries on the condition.

propeciahelp.com

Originating in 2003, propeciahelp is the largest and longest running website for patients suffering from persistent sexual, neurological and physiological side effects arising following use of the drug Finasteride (branded Propecia). Propeciahelp.com aims to provide a place of discussion for those affected by PFS. Propeciahelp's discussion forum is a vast record of PFS patient experiences and has been considered a source of clinical information (Diviccaro et al., 2020)?. Although the quality of discussion is variable, patient posts which provide clear accounts of individual symptoms, the manner of onset and disease progression are of clinical value. Submissions have been the subject of published attempts at recording and categorising the multi-system symptom profile (Walf et al., 2018)?.

In the absence of adequate clinical education regarding PFS, propeciahelp remains a key support to many patients. Along with the families of PFS patients who were driven to suicide by their symptoms, the administrative staff of propeciahelp assisted in the formation of the [PFS foundation](#) which has funded scientific research into the condition through charitable donations. To deliver a structured clinical characterisation of the condition, propeciahelp launched a comprehensive [Post-Drug Syndrome Survey in March 2019](#) and recently passed 200 submissions from post-Finasteride patients experiencing persistent symptoms for a minimum of three months following cessation. We will seek to publish detailed results in the future.

As well as having designed and gathered the largest standardised self-reported dataset concerning PFS, over a decade operating the largest patient support website provides us with a unique insight into the clinical situation. The administrators of propeciahelp have herein summarised current research on PFS. We additionally present a contextual mechanistic hypothesis as basis for future investigation. The vital role of the AR in physiological domains relevant to the symptomatology of PFS is reviewed. This document is intended to aid those with scientific interest in understanding the condition and the practical expertise to uncover the molecular mechanisms underlying this remarkable disease.

Page Bibliography

1. Baas, W. R., Butcher, M. J., Lwin, A., Holland, B., Herberts, M., Clemons, J., Delfino, K., Althof, S., Kohler, T. S., & McVary, K. T. (2018). A Review of the FAERS Data on 5-Alpha Reductase Inhibitors: Implications for Postfinasteride Syndrome. *Urology*, 143–149. <https://doi.org/10.1016/j.urology.2018.06.022>
2. Diviccaro, S., Melcangi, R. C., & Giatti, S. (2020). Post-finasteride syndrome: An emerging clinical problem. *Neurobiology of Stress*, 100209. <https://doi.org/10.1016/j.ynstr.2019.100209>
3. Garreton, A. S., Valzacchi, G. R., & Layus, O. (2016). Post-Finasteride Syndrome: About 2 Cases and Review of the Literature. *Andrology-Open Access*. <https://doi.org/10.4172/2472-1212.1000170>
4. Kenrick, D. T., Griskevicius, V., Neuberg, S. L., & Schaller, M. (2010). Renovating the Pyramid of Needs. *Perspectives on Psychological Science*, 292–314. <https://doi.org/10.1177/1745691610369469>
5. Lavan, A. H., & Gallagher, P. (2015). Predicting risk of adverse drug reactions in older adults. *Therapeutic Advances in Drug Safety*, 11–22. <https://doi.org/10.1177/2042098615615472>
6. Morgans, J. (2018, April 24). *I Need to Quit Hair Loss Drugs Before They Kill Me*. Vice. https://www.vice.com/en_uk/article/43bm3m/i-need-to-quit-hair-loss-drugs-before-they-kill-me
7. PFS Foundation. (2017). *PFS Foundation citizen's petition. FDA-2017-P-5787, Request FDA require the immediate removal of Propecia (and generic formulations of finasteride, 1 mg, for androgenic alopecia) from the market*. Regulations.Gov. <https://www.regulations.gov/docket?D=FDA-2017-P-5787>

8. Than, J. K., Rodriguez, K., & Khera, M. (2018). Post-finasteride Syndrome: A Review of Current Literature. *Current Sexual Health Reports*, 152–157. <https://doi.org/10.1007/s11930-018-0163-4>

 9. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>

 10. Walf, A. A., Kaurejo, S., & Frye, C. A. (2018). Research Brief: Self-Reports of a Constellation of Persistent Antiandrogenic, Estrogenic, Physical, and Psychological Effects of Finasteride Usage Among Men. *American Journal of Men's Health*, 900–906. <https://doi.org/10.1177/1557988317750989>
-

A summary of published research into PFS

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/a-summary-of-published-research-into-pfs/>

In 2011, Traish et al. determined the possibility of a causal relationship between 5 α -reductase inhibitor use and persistent sexual dysfunction in a subset of consumers. They reported the case of a young man who suffered significant sexual dysfunction, libido loss and a depressive symptomatology that continued 11 years after only weeks of exposure to finasteride for AGA (Traish et al., 2011). Irwig and Kolukula first characterised persistent sexual dysfunction in 71 otherwise healthy men who had used finasteride for AGA. 94% experienced low libido, 92% had erectile dysfunction, 92% developed decreased arousal, and 69% developed orgasm dysfunction. Compared with before use, mean sexual episodes per month dropped and sexual dysfunction score per Arizona Sexual Experience Scale increased ($P < 0.0001$) (Irwig & Kolukula, 2011). In a follow up report, 54 of these men were reassessed at a mean of 14 months following their initial interview dates. The mean age of the patients was 31 years and the mean age at assumption of finasteride was 26 years, with a mean duration of 23 months of use. Participants had no baseline sexual dysfunction or psychiatric conditions before use of finasteride. Persistent sexual side effects continued to be present in 96%. 89% of subjects continued to meet the definition of sexual dysfunction according to ASEX. Mean (\pm SD) total scores were 7.2 ± 2.0 before finasteride, 22.2 ± 2.6 after finasteride at the time of the interview, and 20.8 ± 3.6 at reassessment. Severity of sexual dysfunction did not correlate to duration of finasteride use or duration of persistent effects. Irwig noted a broad range of commonly reported persistent effects beyond those formally assessed by the ASEX including decreased volume of ejaculate, loss of penile size and/or testicular size, testicular pain, prostatitis, penile curvature, reduced penile sensation, a reduction in spontaneous erections. Additionally, subjects reported difficulty sleeping, mental impairment, and depressive symptoms (Irwig, 2012b).

In a group of 61 PFS subjects who completed the BDI-II, rates of depressive symptoms were markedly higher (75%) than the control group (10%). While 3% of controls reported suicidal thoughts, this was significantly more frequent in PFS patients. 39% of PFS patients exhibited suicidality with 5% having chosen the statement "I would like to kill myself". Mean (\pm SD) scores from the 21-item BDI-II were $23.67 (\pm 12.56)$ in PFS patients and $5.93 (\pm 4.48)$ in the control group ($P < .0001$) (Irwig, 2012a). Irwig additionally reported a decrease in alcohol consumption in a cohort of 63 PFS patients compared to before use of finasteride. Mean alcoholic beverages consumed per week declined from 5.2 ± 0.7 before finasteride to 2.0 ± 0.3 after finasteride ($p < 0.0001$), consistent with observations regarding finasteride's attenuation of alcohol consumption in animal experiments (Irwig, 2013). Reporting androgen levels and semen parameters in 24 PFS patients, 13% were found to have low total testosterone and 13% had low serum DHT, however mean levels were close to other studies and could not explain the persistent symptoms. 16% (3 of 19) had severe oligospermia, whereas this finding would be expected in 3% of fertile-age men (Irwig, 2014). Considering the medical records of 6 men who committed suicide following use and cessation of finasteride, Irwig noted common symptoms of persistent sexual dysfunction and insomnia across all cases (Irwig, 2020).

Drasa et al. enrolled 35 patients with persistent sexual and nonsexual symptoms following use and cessation of finasteride for male pattern hair loss. Patients had an average age of 30 and mean use of finasteride was 24 months. Assessment was a mean of 12 months after discontinuation. ASEX completion with respect to before and after use of finasteride showed a mean increase of 14.75. 59% of patients experienced severe symptoms per the AMS. 68% of participants reported a worsening after cessation of the drug, and a trend of symptomatic worsening over the course of their condition was reported by 63%. Duration of use and symptom severity were not statistically associated (Drasa, 2014)?.

Ganzer et al. reported a characterisation of the persistent physical, psychological, and cognitive symptoms experienced by PFS patients who had used the drug for at least 3 months and experienced health problems after cessation of finasteride. The authors constructed a web-based questionnaire including ad-hoc questions based around a symptom profile generated from literature review and 100 private patients' reports to the author's practice. Demographic characteristics, data regarding use and cessation of the drug, and the onset of symptoms were also ascertained. Additionally, men were questioned as to the medical support they had sought and their satisfaction with their clinical assessment and treatments. 100 patients who had sought medical assistance were invited by email and additional patients were recruited from propeciahelp.com. No participant had a pre-existing sexual dysfunction or psychiatric condition. 93% reported having used the 1mg finasteride preparation. 84% of patients reported that they were asymptomatic during use of the medication and symptom onset began after cessation, rapidly so in 68% of patients. Respondents reported experiencing physical symptoms of fatigue (69%), muscle atrophy and weakness (56%), fasciculations (47%), decreased oil and sebum (41%), dry and thinned skin (68%), metabolic changes and increased fat deposition (54%). 14% of respondents reported a finding of raised fasting glucose and triglycerides. Sexual dysfunction included diminished libido (93%), loss of spontaneous and morning erections (89%), complete impotence (40%), reduced semen volume and ejaculatory force (82%), orgasm dysfunction (40%) and sexual anhedonia (70%). Penile atrophy (79%) scrotal atrophy (51%) and sensory changes were reported. 20% reported Peyronie's disease. Cognitive complaints were highly prevalent, including severe memory impairment (56%), mental cloudiness or brain fog (75%), impaired problem solving (69%) and attentional deficits (74%). Chronic insomnia was reported by 58% of men. Nearly three quarters of respondents reported increased anxiety, low mood, and anhedonia. Of concern, 63% of respondents had suicidal ideation and felt they could not keep living on with their extreme side effects. In terms of medical support, 50% had initially consulted a urologist while 62% saw their primary care provider. Physicians generally attributed physical symptoms to being of a psychological nature and recommended psychiatric consultation (69%). 93% of men were frustrated by clinical ignorance, inadequate recognition of the validity of their symptoms, and were dissatisfied with the medical care that they received. The authors conclude the aggregate multi-domain symptom profile could constitute a definable syndrome (Ganzer et al., 2014)?.

Chiriaco et al. conducted a similar retrospective evaluation. 79 men who had used finasteride for AGA and experienced persistent symptoms for a minimum of six months were asked to answer 100 ad-hoc questions, both a pre and post-finasteride ASEX questionnaire, and the Aging Male Symptom Scale

(AMS) questionnaire. Mean age of participants was 33. Finasteride had been taken for an average of approximately 2 years. All subjects were still symptomatic at assessment. 89.9% of participants noticed some symptoms during finasteride use, and the trend of symptoms after discontinuation was worsening in 62% of patients, with a trend of improvement reported in 13.9%. Sexual symptoms included loss of penile sensitivity (87.3%), decreased ejaculatory force (82.3%), decreased penile temperature (78.5%), reduced ejaculate volume (73.4%), reduction in penile dimension (65.8%), perineal tightness (45.6%). Other symptoms included anhedonia (75.9%), concentration problems (72.2%), loss of muscle tone and mass (51.9%), and increased body weight (48.1%). Post-finasteride ASEX score ranged from 13–30 (21.0 ± 2.67), with 78.5% having ASEX ≥ 19 points indicating sexual dysfunction. This included 44.3% of patients indicating severe difficulty or incapability of getting/keeping an erection. Pre-finasteride ASEX score was far lower ($p < 0.001$) ranging 5–15 (7.7 ± 2.52), indicating no overt sexual dysfunction. Of 78 patients with available data, all had some signs of androgen deficiency per the AMS, with 60.3% with an AMS score of ≥ 50 points indicating severe deficiency. The authors note the reports by their PFS patients suggest androgen deficiency across different tissues where 5 α reductase is expressed at an average of four years after finasteride discontinuation, indicating that permanent changes occurred in the human body (Chiriaco et al., 2016).

Walf et al. sought to characterise persistent symptoms following finasteride treatment and its discontinuation by assessment of subjective patient reports on propeciahelp.com. 244 cases were isolated from discussions in a discrete time period. Walf et al. placed symptoms into four broad categories: Antiandrogenic effects, estrogenic effects, central effects and nonspecific adverse effects. Antiandrogenic adverse effects were described to be genital dysfunction, testicular dysfunction and infertility, accessory sexual or genitourinary organ dysfunction, psychosexual function, and hormonal function. Estrogenic AEs included breast cancer, breast neoplasm or breast mass, gynecomastia, breast pain, and increased serum estrogen. Central effects involved depression, anxiety, confusion and “brain fog”, insomnia and attentional difficulties. The nonspecific/severe AEs were defined as muscle twitching, lower back pain, weight gain, fatigue, numbness in the anal region, muscle spasms, excessive sweating, bleeding gums, tinnitus, hot flashes, irregular stool, scoliosis, and discoloration of the urine. While these presented heterogeneously, some individuals experienced adverse events across all categories (Walf et al., 2018).

In a retrospective control matched study, Di Loreto et al. evaluated expression of the androgen receptor and nerve density in multiple cell lines of prepuce tissue in PFS patients aged 29–43 years who had experienced persistent sexual symptoms for over 6 months, with the notable finding of persistent androgen receptor overexpression (Di Loreto et al., 2014). Patients had used finasteride for an average of 32 months and had stopped using an average of 56 months to the point of study, at which point all patients remained symptomatic. PFS patients self-reported symptoms including loss of penile sensation, erectile dysfunction, pain in the penis, scrotum or testes, penile tissue changes, reduced penile dimensions, and reduced volume of ejaculate. PFS cases experienced sexual dysfunction at point of interview per Arizona Sexual Experience Scale (22.5 ± 2.78). PFS patients were additionally asked to complete the ASEX survey considering themselves before use of finasteride and these pre-finasteride scores indicated no pre-existing sexual dysfunction (7.6 ± 1.92). Histological evaluation of nerve density revealed similarity with controls. Immunohistochemistry revealed a significantly higher percentage of

nuclear AR-positive epithelial cells in all cases (mean±SD, 80.6±8.63%) than in controls (mean±SD, 65.0±19.1%), $P = 0.043$. Stromal cells in all cases showed a significantly greater expression of AR in the nuclei compared to controls (mean±SD, 40.0±15.1% in cases versus 23.4±8.68% in controls), $P = 0.023$. Percentage of AR positive vessel smooth muscle cells did not differ significantly between the 2 groups. Averagely, AR positive cells in the 3 tissues was higher in cases than in controls. Di Loreto et al. speculate that the ostensibly permanent effects could be due to mechanisms of ageing prematurely induced by artificially reduced androgen levels with finasteride. They conclude a better understanding of the molecular events may inform possible therapies for these severe effects in young men of fertile age (Di Loreto et al., 2014)?. Although unreported in the manuscript, La Marra, co-author of the study, further elaborated on the data in a thesis centring on the investigation. He reported the percentages of AR positive cells are always higher in the cases than in the controls. He further reported positive correlations between the increase of AR levels in the epithelial and stromal cells and the decrease in ability/frequency to perform sexually per the AMS, the increase of AR in the vessels cells and the intensification of ASEX sexual dysfunction and physical exhaustion, and the increase of AR in the epithelial cells and the worsening of muscular weakness and feeling "burnt out" per AMS. La Marra noted that exogenous androgens do little to improve - and sometimes worsen - PFS symptoms, concluding that investigations should centre on epigenetic alterations relevant to the changed sensitivity of the AR (La Marra, 2010)?. Di Loreto's investigation was the first to report significant objective differences at the molecular level and it has subsequently been suggested that local AR levels could play a pathological role in PFS (Than et al., 2018; Traish, 2018)?.

Demonstrating multisystem involvement in absence of what the authors regarded to be the "typical" neurological and sexual complaints, Gupta et al. reported a 33 year old man with PFS who suffered itching, burning micturition, abdominal discomfort, skin rash, and seborrhoea after a first use of 0.5mg dutasteride for a month. These symptoms subsided with the adoption of exercise but had reoccurred and persisted after attempting AGA therapy with finasteride 1mg four years later. Keratotic follicular papules and pustules were apparent on his shoulders and back. Semen analysis revealed pus cells and moderate growth of *Enterococcus faecalis* following culture. Therapeutic attempts over the following years at three centres were not successful (Sharma et al., 2016)?. Motofei also reported an uncommon presentation in a 52 year old who presented with generalised vitiligo 2 months after cessation alongside symptoms including bilateral gynecomastia, sexual dysfunction and depression that were not present upon pre-treatment evaluation (Motofei et al., 2017)?.

Cecchin et al. reported a significantly higher occurrence of "extreme length" AR polymorphisms (CAG-rs4045402 and GGN-rs3138869) in PFS patients following finasteride use for AGA as compared to controls without AGA, suggestive of a potential genetic role in the development of AGA and PFS (Cecchin et al., 2014)?. Cauci et al. expanded on this in a subsequent study exploring the relationship of AR polyglutamine stretch-encoding (CAG) and polyglycine stretch-encoding (GGN) polymorphisms with the individual symptoms of PFS in 66 patients experiencing symptoms for a median of three years after cessation. Patients were asked to describe their trend of symptoms after discontinuation (improved, unchanged or worsening). 57.6% of PFS patients responded that their trend after was worsening. Androgen receptor polymorphisms were correlated to the frequency of several PFS symptoms. Patients

completed a bespoke symptom questionnaire in addition to the ASEX and the AMS. While total scores of the ASEX and AMS did not differ with length of (CAG)_n and (GGN)_n repeats, significant differences were found within individual PFS symptoms. Patients with shorter CAG repeat lengths (9-19) used finasteride for a shorter time than those with medium (20-24) or long (>25) repeat lengths, and 83.3% of this short CAG_n group reported severe libido loss, scoring 5 on item 17 of the AMS. Increased body weight (>2kg) following use of finasteride was most associated with those with long CAG repeats. Interestingly, skin dryness showed a parabolic curvilinear profile, with short and long CAG_n groups having higher frequencies (50% and 63.6% respectively) than the medium CAG_n group (18.9%). Muscle spasms were found to be more frequent amongst long CAG_n carriers (72.7%). Patients with long (>23) GGN repeats did not report experiencing scrotal pain compared with 34.1% of those with medium (23) GGN repeats and 32.7% of those with medium to short length (<23) repeats. Penile pain was likewise more often seen in those with short or medium rather than long GGN repeats (34.6% vs 7.1%). Long GGN repeats were also associated with a better phenotype regarding fatigue, loss of vitality, depression and the feeling of passing one's peak than those with medium repeats. Loss of perineal fullness was reported by 100% of men with short GGN_n repeat lengths, 70.5% of men with medium GGN repeats and 57.1% of those with long repeats. The results of Cauci et al. suggest genetic involvement in the symptom profile of PFS, and the authors conclude the need for much more research into the pathophysiology, particularly with a precision medicine approach (Cauci et al., 2017)?.

In a clinical assessment of 24 PFS patients, Basaria et al. found no significant sequence variations in AR, SRD5A1 or SRD5A2. Depression scores were significantly higher in PFS patients via BDI, Hamilton Depression inventory and PHQ-9. PHQ-9 scoring was not significantly related to either the duration of finasteride use or the time since discontinuation of the drug. Some characteristics were measured and were not significantly different to controls. No hormonal correlate able to account for the pathological presentation was identified. Two fMRI measurements suggested neurobiological abnormalities PFS patients. fMRI of PFS patients' brains in response to erotic stimuli was conducted. Worsening IIEF scores correlated to increased activity in the neural areas the authors deemed to correspond with sexual arousal, while activity in brain regions associated with higher level cognitive and motivational networks decreased concomitantly, revealing a dissociation in activity that may be a marker of neural changes following use of finasteride. Blood-oxygen dependent activity in brain areas implicated in major depression were also identified in PFS patients with correlation to BDI scores pertaining to negative affect (Basaria et al., 2016)?. This study included limited gene expression assay of skin taken from the back of symptomatic patients and non-symptomatic finasteride users. Although the paper stated that "we did not find evidence of...significant alterations in expression of AR-dependent genes in the skin", this is not completely reflective of the statement in the study's supplementary appendix: "While the DESeq analysis determined there were statistically significant differences in a few of the androgen-regulated genes, the hierarchical clustering analysis revealed that the symptomatic and non-symptomatic subjects did not share the immediate cluster" (Basaria et al., 2016 appendix: methods).

Melcangi et al. Performed case-controlled clinical evaluations of 16 PFS patients aged 22-44 with a strong focus on the neurological presentation of the syndrome. Mean treatment duration was 1037 days with a range of 451–4697 days between cessation of finasteride and clinical evaluation. 50% of PFS

patients were deemed to suffer from major depressive disorder per screening with the Mini-International Neuropsychiatric Interview, and scores of Beck Depression Inventory and Beck Anxiety Inventory were significantly higher in those with MDD. Ten patients experienced severe ED per the IIEF15, while the remaining 6 exhibited mild to moderate ED. Ultrasound determination of testicular volume was calculated to be normal in patients. Objective markers of neuropathy were determined in 25% of patients via sensory evoked potentials of the pudendal nerve, while 75% of the patients had normal PN_SEPs. No evidence of metabolic, toxic, or inherited disease associated with peripheral nervous system damage was detected. Interestingly, depression scores were not correlated to PN_SEPs while sexual dysfunction scores were. The cerebrospinal fluid of 14 patients was analysed with comparison to 25 healthy age-matched controls. Significant differences were determined. Pregnenolone, isopregnanolone, progesterone and dihydroprogesterone were significantly decreased, while levels of dehydroepiandrosterone (DHEA), testosterone and 3 β -diol were increased. Additionally, 17 β -estradiol and DHT were decreased. Plasma determination showed differences to the CSF findings. In serum, pregnenolone, tetrahydroprogesterone, DHEA and T were significantly increased, while dihydroprogesterone was significantly decreased (Melcangi et al., 2017). This disruption in neurosteroids is notably heterogeneous and differed slightly to findings from their previous pilot study involving 3 PFS patients (Melcangi et al., 2013). Melcangi et al. later reported that the gene promoter of SRD5A2 was methylated in CSF samples of 9 of 16 PFS patients (age 34.5 \pm 8.8 years) compared with 1 of 13 age-matched controls. Interestingly, the single control with SRD5A2 methylation had a diagnosis of normotensive hydrocephalus. Amongst PFS patients the methylation ranged from 15.4 to 100%. Neither depression, anxiety or erectile dysfunction scoring via validated scales were correlated to methylation status. Methylation was not found in blood DNA, demonstrating tissue specificity. SRD5A1 was found to be unmethylated across samples and groups (Melcangi et al., 2019).

Rubin et al. Performed penile duplex Doppler ultrasound examination with a high frequency probe during maximal pharmacologic erection on 27 PFS patients. Patients had a mean age of 31, no known cardiovascular risk factors, and had sexual dysfunction following use of finasteride. 26 of 27 patients (96%) demonstrated lack of homogeneity and hyperechoic/hypoechoic regions in erectile tissue. They concluded induced corporal smooth muscle apoptosis and fibrosis may represent a biologic pathophysiology responsible for impairing tissue expandability resulted in venoocclusive dysfunction and ED (Rubin et al., 2018). Mirabal et al. issued 25 patients with persistent symptoms following 5ari use for AGA and 25 controls a range of validated questionnaires related to self-reported symptomatology including the IIEF, the International Prostate Symptom Score (IPSS), the Patient Health Questionnaire-9 (PHQ-9) and the Androgen Deficiency in the Aging Male (ADAM). Post-5ari patients had significantly higher median scores compared with controls in the IIEF (35 vs 29, p=0.035), the IPSS (10 vs 3, p < 0.01), the PHQ-9 (10 vs 1, p < 0.001), and had significant differences in all questions of the ADAM. Penile duplex doppler ultrasound revealed vascular abnormalities in 17 (68%) post-5ari patients. Alarmingly, 2 (8%) of post-5ari patients committed suicide during and after the study. Mirabal et al. concluded that there may be persistent genitourinary, physical, psycho-cognitive, anti-androgenic and penile vascular changes after 5ARI discontinuation in addition to persistent sexual dysfunction (Mirabal, 2019).

Epidemiological research into PFS has thus far been limited to sexual dysfunction and depressive symptoms. Ali et al. used data mining techniques with FAERS data to conduct a retrospective pharmacovigilance disproportionality analysis. They analysed reports of sexual dysfunction and suicidal ideation between 1998 and 2013 in men aged 18-45 who had used low-dose finasteride. Supportive of survey research previously discussed, the data revealed that a strong signal of persistent sexual dysfunction and disproportional reporting of suicidal ideation. Most sexual dysfunction reports were serious, with 43.5% resulting in disability. 87% of incidences of suicidal ideation occurred in men also experiencing sexual dysfunction from low-dose finasteride. Most of these events were classed as serious (e.g., contributed to the patient's death, hospitalization, or disability). Ali note there is mechanistic plausibility in the link between finasteride and the risks of sexual dysfunction and suicidal ideation, and that the disproportional reporting could be symptoms of Post-Finasteride Syndrome. The authors conclude that, although a causal link cannot be inferred from this study due to the nature of the data, young men receiving low-dose finasteride for AGA are at risk of persistent sexual dysfunction that may lead to suicidal ideation (Ali et al., 2015)?.

Kiguradze et al. have provided a well-designed analysis of a large set of data from the Northwestern Medicine Enterprise Data Warehouse with sole regards to persistent erectile dysfunction following use of finasteride or dutasteride. They conclusively identified a strong and intrinsic association between debilitating persistent sexual dysfunction and exposure to low dose finasteride or dutasteride. Duration of 5-alpha reductase inhibitor exposure was a greater predictive risk factor for ED in young men than all other assessed factors. Of 4,284 young men, without prior sexual dysfunction, taking finasteride at a dose less than 1.25 mg/day, 34 (0.79%) developed persistent erectile dysfunction with a median 1,534 days after drug cessation (interquartile range of 651–2,351 days). Of 103 young men with new ED, 34 (33%) had new persistent erectile dysfunction (Kiguradze et al., 2017)?.

Through obtaining finasteride-related adverse events catalogued by the FAERS reporting system between April 2011 and October 2014, Fiuk et al. identified 105 women with finasteride-associated adverse events following use. These included typical PFS symptoms including dry eyes, sleep disturbances and suicidal ideation as well as hearing loss, renal failure, urosepsis, new incidences of breast cancer, haemorrhagic diathesis. They concluded female PFS patients represent a small but real subset of long term finasteride-related adverse events, and that further etiological investigation of this devastating syndrome is crucial (Fiuk et al., 2016)?.

In addition to the significant primary findings in PFS patients discussed, the subject is far more regularly the focus of literature review. Than et al. concluded that the existing evidence well supports the existence of persistent sexual, physical, neurological and central effects following 5alpha reductase inhibitor exposure, and that a growing understanding of the constellation of symptoms describing PFS can inform prescribing clinicians as to the risk and benefit of prescription (Than et al., 2018)?.

Traish considered that the magnitude of the broad and serious symptomatology of the syndrome is inadequately appreciated. Persistent loss of libido and erectile dysfunction are recognised to be serious issues pertaining to quality

of life and wellbeing, as well as "signs of something terribly amiss with physiological process". Traish further deemed it imperative that the scientific and medical communities act now to seek a better understanding the pathophysiology of this serious and debilitating disorder, expand awareness amongst physicians and patients, and develop tools for treatment (Traish, 2018). Said and Mehta concluded that comprehensive literature review shows a disproportionately high number of men with 5- α reductase inhibitor-associated sexual dysfunction and infertility, and that though uncommon, broad sexual and reproductive symptoms that are both serious and persistent can occur. They note that while methodological concerns have been raised regarding the possibilities of recall and selection bias in the questionnaire-based study of PFS patients, their results parallel scientific observations about the long-term pathophysiological changes induced by finasteride, even after treatment discontinuation. They suggest physicians engage in productive conversation regarding the potential impact of these medications on their health and quality of life before 5 α reductase inhibitor prescription (Said & Mehta, 2018).

Page Bibliography

1. Ali, A. K., Heran, B. S., & Etminan, M. (2015). Persistent Sexual Dysfunction and Suicidal Ideation in Young Men Treated with Low-Dose Finasteride: A Pharmacovigilance Study. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 687–695. <https://doi.org/10.1002/phar.1612>
2. Basaria, S., Jasuja, R., Huang, G., Wharton, W., Pan, H., Pencina, K., Li, Z., Travison, T. G., Bhawan, J., Gonthier, R., Labrie, F., Dury, A. Y., Serra, C., Papazian, A., O'Leary, M., Amr, S., Storer, T. W., Stern, E., & Bhasin, S. (2016). Characteristics of Men Who Report Persistent Sexual Symptoms After Finasteride Use for Hair Loss. *The Journal of Clinical Endocrinology & Metabolism*, 4669–4680. <https://doi.org/10.1210/jc.2016-2726>
3. Cauci, S., Chiriaco, G., Cecchin, E., Toffoli, G., Xodo, S., Stinco, G., & Trombetta, C. (2017). Androgen Receptor (AR) Gene (CAG)_n and (GGN)_n Length Polymorphisms and Symptoms in Young Males With Long-Lasting Adverse Effects After Finasteride Use Against Androgenic Alopecia. *Sexual Medicine*, e61–e71. <https://doi.org/10.1016/j.esxm.2016.11.001>
4. Cecchin, E., De Mattia, E., Mazzon, G., Cauci, S., Trombetta, C., & Toffoli, G. (2014). A Pharmacogenetic Survey of Androgen Receptor (CAG)_N and (GGN)_N Polymorphisms in Patients Experiencing Long Term Side Effects after Finasteride Discontinuation. *The International Journal of Biological Markers*, 310–316. <https://doi.org/10.5301/jbm.5000095>
5. Chiriaco, G., Cauci, S., Mazzon, G., & Trombetta, C. (2016). An observational retrospective evaluation of 79 young men with long-term adverse effects after use of finasteride against androgenetic alopecia. *Andrology*, 245–250. <https://doi.org/10.1111/andr.12147>

6. Di Loreto, C., La Marra, F., Mazzon, G., Belgrano, E., Trombetta, C., & Cauci, S. (2014). Immunohistochemical Evaluation of Androgen Receptor and Nerve Structure Density in Human Prepuce from Patients with Persistent Sexual Side Effects after Finasteride Use for Androgenetic Alopecia. *PLoS ONE*, e100237. <https://doi.org/10.1371/journal.pone.0100237>
7. Drasa, K. (2014). *Sexual and nonsexual problems after finasteride used for hair loss in young men*. American Society for Men's Health annual meeting.
8. Fiuk, J., Butcher, M., Kohler, T., & McVary, K. (2016). 133 Female Post-Finasteride Syndrome: It's Not Just A Man's World. *The Journal of Sexual Medicine*, S62–S63. <https://doi.org/10.1016/j.jsxm.2016.02.139>
9. Ganzer, C. A., Jacobs, A. R., & Iqbal, F. (2014). Persistent Sexual, Emotional, and Cognitive Impairment Post-Finasteride. *American Journal of Men's Health*, 222–228. <https://doi.org/10.1177/1557988314538445>
10. Irwig, M. S. (2012a). Depressive Symptoms and Suicidal Thoughts Among Former Users of Finasteride With Persistent Sexual Side Effects. *The Journal of Clinical Psychiatry*, 1220–1223. <https://doi.org/10.4088/jcp.12m07887>
11. Irwig, M. S. (2012b). Persistent Sexual Side Effects of Finasteride: Could They Be Permanent? *The Journal of Sexual Medicine*, 2927–2932. <https://doi.org/10.1111/j.1743-6109.2012.02846.x>
12. Irwig, M. S. (2013). Decreased Alcohol Consumption Among Former Male Users of Finasteride with Persistent Sexual Side Effects: A Preliminary Report. *Alcoholism: Clinical and Experimental Research*, 1823–1826. <https://doi.org/10.1111/acer.12177>
13. Irwig, M. S. (2014). Androgen Levels and Semen Parameters Among Former Users of Finasteride With Persistent Sexual Adverse Effects. *JAMA Dermatology*, 1361. <https://doi.org/10.1001/jamadermatol.2014.1830>
14. Irwig, M. S. (2020). Finasteride and Suicide: A Postmarketing Case Series. *Dermatology*, 1–6. <https://doi.org/10.1159/000505151>
15. Irwig, M. S., & Kolukula, S. (2011). Persistent Sexual Side Effects of Finasteride for Male Pattern

Hair Loss. *The Journal of Sexual Medicine*, 1747–1753.

<https://doi.org/10.1111/j.1743-6109.2011.02255.x>

16. Kiguradze, T., Temps, W. H., Yarnold, P. R., Cashy, J., Brannigan, R. E., Nardone, B., Micali, G., West, D. P., & Belknap, S. M. (2017). Persistent erectile dysfunction in men exposed to the 5 α -reductase inhibitors, finasteride, or dutasteride. *PeerJ*, e3020. <https://doi.org/10.7717/peerj.3020>
17. La Marra, F. (2010). *The post-finasteride syndrome in patients with alopecia*. University of Udine.
18. Melcangi, R. C., Caruso, D., Abbiati, F., Giatti, S., Calabrese, D., Piazza, F., & Cavaletti, G. (2013). Neuroactive Steroid Levels are Modified in Cerebrospinal Fluid and Plasma of Post-Finasteride Patients Showing Persistent Sexual Side Effects and Anxious/Depressive Symptomatology. *The Journal of Sexual Medicine*, 2598–2603. <https://doi.org/10.1111/jsm.12269>
19. Melcangi, R. C., Casarini, L., Marino, M., Santi, D., Sperduti, S., Giatti, S., Diviccaro, S., Grimoldi, M., Caruso, D., Cavaletti, G., & Simoni, M. (2019). Altered methylation pattern of the SRD5A2 gene in the cerebrospinal fluid of post-finasteride patients: a pilot study. *Endocrine Connections*, 1118–1125. <https://doi.org/10.1530/ec-19-0199>
20. Melcangi, R. C., Santi, D., Spezzano, R., Grimoldi, M., Tabacchi, T., Fusco, M. L., Diviccaro, S., Giatti, S., Carrà, G., Caruso, D., Simoni, M., & Cavaletti, G. (2017). Neuroactive steroid levels and psychiatric and andrological features in post-finasteride patients. *The Journal of Steroid Biochemistry and Molecular Biology*, 229–235. <https://doi.org/10.1016/j.jsbmb.2017.04.003>
21. Mirabal, J. R. (2019, October 24). *The Persistent Effects of 5 α -reductase Inhibitors Used in the Treatment of Androgenic Alopecia on Men's Health*. 20th Annual Fall Scientific Meeting Of Sexual Medicine Society of North America, Omni Nashville Hotel, Nashville, TN, USA.
22. Motofei, I. G., Rowland, D. L., Georgescu, S. R., Tampa, M., Paunica, S., Constantin, V. D., Balalau, C., Manea, M., Baleanu, B. C., & Sinescu, I. (2017). Post-Finasteride Adverse Effects in Male Androgenic Alopecia: A Case Report of Vitiligo. *Skin Pharmacology and Physiology*, 42–45. <https://doi.org/10.1159/000455972>
23. Rubin, R., Winter, A., & Goldstein, I. (2018). 311 Novel Penile Ultrasound Technique to Explain Mechanism of Erectile Dysfunction (ED) in Young Patients using Finasteride for Androgenic Alopecia. *The Journal of Sexual Medicine*, S81. <https://doi.org/10.1016/j.jsxm.2017.11.194>

24. Said, M. A., & Mehta, A. (2018). The Impact of 5 α -Reductase Inhibitor Use for Male Pattern Hair Loss on Men's Health. *Current Urology Reports*. <https://doi.org/10.1007/s11934-018-0814-z>

 25. Sharma, N., Gupta, A., & Shukla, P. (2016). Atypical post-finasteride syndrome: A pharmacological riddle. *Indian Journal of Pharmacology*, 316. <https://doi.org/10.4103/0253-7613.182898>

 26. Than, J. K., Rodriguez, K., & Khera, M. (2018). Post-finasteride Syndrome: A Review of Current Literature. *Current Sexual Health Reports*, 152–157. <https://doi.org/10.1007/s11930-018-0163-4>

 27. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>

 28. Traish, A. M., Hassani, J., Guay, A. T., Zitzmann, M., & Hansen, M. L. (2011). Adverse Side Effects of 5 α -Reductase Inhibitors Therapy: Persistent Diminished Libido and Erectile Dysfunction and Depression in a Subset of Patients. *The Journal of Sexual Medicine*, 872–884. <https://doi.org/10.1111/j.1743-6109.2010.02157.x>

 29. Walf, A. A., Kaurejo, S., & Frye, C. A. (2018). Research Brief: Self-Reports of a Constellation of Persistent Antiandrogenic, Estrogenic, Physical, and Psychological Effects of Finasteride Usage Among Men. *American Journal of Men's Health*, 900–906. <https://doi.org/10.1177/1557988317750989>
-

Finasteride Drug origin, pharmacology, AR structure and androgen action

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/finasteride-drug-origin-pharmacology-ar-structure-and-androgen-action/>

Androgens, the androgen receptor and the expanding understanding of its role in health

Androgens are well appreciated for their critical developmental role in sexual differentiation (Forger, 2018), male characteristics, the development and maintenance of male sexual organs and sexual function (C J Bagatell et al., 1994; Podlasek et al., 2016; A. M. Traish, 2008; Yamada et al., 2006). However, androgens are now known to play a "pleiotropic role...in virtually all body systems" (Gibson et al., 2018). Gibson et al. identify four key areas in which the understanding of the role of androgens has evolved and expanded in the 21st century: Testosterone's recognition as a "Goldilocks" molecule, with too much or too little androgen signalling disrupting cellular homeostasis and proving deleterious to health, a dynamic and tissue specific regulation of intracrine androgen metabolism, an increased understanding of the role of androgens in female reproductive tissue, and the extensive role for androgen-mediated regulation in tissue beyond the reproductive system in both sexes (Gibson et al., 2018). At the tissue level, there are tightly controlled optimum levels for androgen concentrations. Owing to the crucial role of androgen intracrine biosynthesis and metabolism in the physiology of peripheral tissues in males and females, dysregulation can impair both local and systemic metabolic homeostasis (Carrie J. Bagatell & Bremner, 1996; Schiffer et al., 2018).

Nuclear receptors are ancient proteins well conserved across evolutionary time and are present across the Metazoa (King-Jones & Thummel, 2005). The effects of androgen steroids are primarily mediated through the Androgen Receptor (Verhoeven & Swinnen, 1999), a class I steroidal receptor protein which binds androgens as ligand in the cytoplasm, dissociates from chaperones and translocates to the nucleus (Davey & Grossmann, 2016; Ni et al., 2013; Tsai & O'Malley, 1994). The AR is ubiquitously expressed across most bodily tissues including the brain and nervous system, penis, testes, prostate, skeletal muscle, skin, liver, urinary bladder, gastrointestinal tract, arteries, kidneys, breast, uterus, bone, adrenal glands, and teeth (Dale et al., 2002; Fujimoto et al., 1994; Gannon et al., 2019; Heemers & Tindall, 2007; Khalil et al., 2018; Kimura et al., 1993; Mhaouty-Kodja, 2018; Ruizeveld de Winter et al., 1991; Schultheiss et al., 2003; Sinha-Hikim et al., 2004; Vanderschueren et al., 2014; Verhoeven & Swinnen, 1999; Wu et al., 2019; Xia et al., 2019). Significant evidence has demonstrated the AR is expressed across many areas of the brain in both sexes including the temporal, medial preoptic, hypothalamus, amygdala, bed nucleus of the stria terminalis, midbrain, frontal and prefrontal areas, cingulate gyrus, and limbic regions including the hippocampus, where it is critical to important

neurocognitive functions including reproductive behaviour, reward behaviour, learning, memory, spatial awareness and metabolic regulation (Beyenburg et al., 2000; Brock et al., 2015; Lu et al., 1998; Morford et al., 2018; Shah et al., 2004; Simerly et al., 1990; Tobiansky et al., 2018)?. The role of the AR in disease cannot be overstated (Koryakina et al., 2014)? owing to its role as an important hub mediating multiple cellular signals and functions (Lai et al., 2012)?.

The AR is coded from eight exons located in the long arm of the X-chromosome (J. Brand & M. Dehm, 2013; Lubahn et al., 1988)?, lacks a TATA and CCAAT box in the regulatory promotor, and is comprised of four distinct domains acting together to mediate genomic effects of androgens in target tissue. These are the N-terminal domain, the DNA binding domain, the hinge region, and the C-terminal ligand binding domain (Brinkmann et al., 1989; Lanciotti et al., 2019; Mangelsdorf et al., 1995; I J McEwan, 2004)?. The AR is regulated by ligand binding, interaction of functional domains (such as N/C terminal interaction), homodimerization and cofactor interactions (van Royen et al., 2012)?. The N-terminal domain is intrinsically disordered and exists as collections of conformers, allowing rapid impermanent structural alterations in response to the cellular environment and binding of multiple coregulators with distinct outcomes (Kumar & McEwan, 2012; I. McEwan & Monaghan, 2016)?. This region contains polymorphic glutamine and glycine tracts (Wadosky & Koochekpour, 2016)?. The ligand independent AF-1 surface in the N-terminal domain interacts with coregulators (Heinlein & Chang, 2002)?. More than 200 AR-interacting proteins with either coactivator or corepressor activities are known (Chang & McDonnell, 2005)?. The open structure of the ligand binding domain (LBD) adopts a compact structure when bound to agonists, which are then sealed within hydrophobic interior (Iain J. McEwan & Kumar, 2015)?. Helix 12 is repositioned to form a surface for transcription promoters (Hur et al., 2004)?. The LBD contains AF-2 which is pivotal to the ligand-dependent full activation of the androgen receptor (Narayanan et al., 2018)? and is affected by coregulators. The AF-2 has high affinity for a highly conserved 5-residue FQNLF motif in the N-terminal segment of the N-terminal domain. The LBD binding to this region facilitates activation, and molecular chaperones compete for binding and prevent activation of the AR in a delicate balance of protein-protein interaction that is seemingly regulatory of activity, solubility, concentration and AR turnover (Eftekharzadeh et al., 2019)?. A nonclassical zinc finger structure in the DNA Binding Domain functions to recognise and make contact with nucleotide sequences, while a second mediates dimerization on DNA (Iain J. McEwan & Kumar, 2015)?. The activation of the AR and targeting of androgen response elements results in increased transcription of a host of genes, many of which control cell growth, proliferation and regulation of apoptosis (Heemers & Tindall, 2007)?. Liganded AR also activates coregulators distinctly from its DNA binding capability (Slagsvold et al., 2001)?. The human AR has AREs and autoregulates its own gene in a tissue-specific manner (Hunter et al., 2018)?. The AR has functional roles beyond transcription, and nonclassical AR mediated actions occur via the ERK, SRC, PI3K, MEK and AKT pathways (Deng et al., 2017; Vanderschueren et al., 2014)?. Recently, ubiquitously expressed specific G protein-coupled receptors known as membrane androgen receptors have been described by which androgens mediate rapid intracellular actions and diverse nonclassical processes, eliciting significant physiological and behavioural effects in animals and humans within seconds or minutes (Balthazart et al., 2018; Foradori et al., 2008; Geniole et al., 2019; Kalyvianaki et al., 2019; Thomas, 2019; Thomas et al., 2018)?. Testosterone association to membrane AR exerts a rapid regulatory influence over classical genomic AR signaling (Deng et al., 2017; Li et al., 2018)?, and appreciation of these effects are therefore more accurately characterised as nonclassical as opposed to nongenomic (Balthazart et al., 2018)?. The rapid

nonclassical actions of the mAR ZIP9 are vulnerable to disruption by endocrine disrupting chemicals known to interfere with classical androgen signaling, and the toxicological consequences of this are currently unclear (Thomas & Dong, 2019)?.

The primary male steroid hormone and AR ligand testosterone is produced by the Leydig cells of the testes (Schiffer et al., 2018). Testosterone is synthesised from cholesterol (Miller, 1988) through a number of steps originating with p450 side-chain cleavage conversion to pregnenolone in the inner mitochondrial membrane (Selvaraj et al., 2018). Leydig cell production of testosterone is stimulated in response to the anterior pituitary releasing LH in response to a pulsatile release of LHRH by the hypothalamus, and testosterone regulates LHRH release via a negative feedback loop (Heemers & Tindall, 2007). Androgen signalling is amplified in target tissue through the metabolism of T to 5 α -dihydrotestosterone (DHT). DHT is the most potent endogenous androgen (Pretorius et al., 2016; Rege et al., 2013), with a four-fold higher binding affinity for the androgen receptor (Gao et al., 2005) and a three-fold lower dissociation rate than that of testosterone (Wilson & French, 1976). Agonists form hydrogen bonds to the AR with high occupancy, with DHT bonding to residue Thr877 and testosterone bonding at Asn705 (Azhagiya Singam et al., 2019). DHT binding causes the AR to undergo conformational change to its DNA binding state (Kovacs et al., 1984), and increases synthesis and degradation of the AR protein (Syms et al., 1985). However, tissue-level factors regulating metabolism including local intracellular ligand concentrations influence binding in addition to relative ligand affinities, and as such DHT does not always bind preferentially compared with T (Swerdloff et al., 2017).

Circulating T is more important than serum DHT for optimizing the intracellular DHT concentrations due to the presence of a rate-limiting enzyme, 5 α -reductase. Testosterone is metabolised to DHT irreversibly by the catalytic microsomal enzyme 5 α -reductase type 2. 5AR2 is a hydrophobic membrane-bound protein comprised of 254-260 amino acid residues (Russell & Wilson, 1994). The 5 α -reductase family of enzymes are diffusely expressed across a large number of tissues, and exert a profound effect on human health due to their regulation of steroid metabolism and metabolic functions including glucocorticoid clearance (Abdulmageed M. Traish et al., 2014)?. 5 α r enzymes catalyze the reduction of the double bond in the A-ring at 4,5 position in C-19 and C-21 steroids (Azzouni et al., 2012; Abdulmageed M. Traish et al., 2015)?.

Development and pharmacology of Finasteride

Loss of appropriate androgen signaling is associated with diverse detrimental effects in males, as evidenced by the well appreciated side effects of androgen deprivation therapy. ADT is known to induce bone problems, metabolic dysfunction, sexual dysfunction, reduction of penile and testicular size, gynecomastia, fatigue, vasomotor flushing, memory, cognitive and psychosocial impairments (Nguyen

et al., 2015)? Heightened androgen action is directly implicated in pathologies including benign prostate hyperplasia, prostate cancer (Banerjee et al., 2018)?, bladder cancer (Gil et al., 2019; Liu et al., 2018)?, androgenic alopecia (Lai et al., 2012)?, lower urinary tract symptoms and polycystic ovary syndrome (Apparao et al., 2002)?. As 5alpha reductase is largely responsible for tissue DHT levels, 5alpha reductase inhibitor products can alleviate symptoms owing to reducing pathological androgen receptor activation. Significant increases in serum DHT via exogenous DHT administration have little effect on prostate DHT concentrations, prostate size, and lower urinary tract symptoms (Swerdlhoff et al., 2017)?. Considering misconceptions likely arising from the lowered serum levels following 5alpha reductase inhibitor therapy coinciding with symptomatic relief in these domains, Swerdlhoff et al. note this illustrates fundamentally important control mechanisms in androgen target tissues that finely regulate androgen synthesis and degradation pathways to maintain DHT homeostasis, to which circulating DHT levels are of much less importance than that of T (Swerdlhoff et al., 2017)?. Beyond these primary androgens, around 5-10% of serum androgens include dehydroepiandrosterone, androstenediol, and androstenedione, which can be produced by ACTH-regulated adrenal synthesis (Rainey et al., 2002)?.

The prostate is a strictly androgen dependent structure (Banerjee et al., 2018)?. The link between androgens and prostate growth was established in the mid-20th century (Nelson, 2016)?. Concurrently, androgens were understood to be both a strict requirement and driver of male pattern hair loss (Hamilton, 1942)?. The selective 5alpha-reductase type 2 inhibitor Finasteride is a 4-azasteroid that was developed as a treatment for benign prostate hyperplasia (BPH) and androgenic alopecia (AGA) by Merck. This programme followed Imperato-McGinley's identification and profiling of pseudohermaphroditism in males with genetic 5aR deficiency (Imperato-McGinley et al., 1974, 1991)?. These (46XY) males demonstrate at birth a marked ambiguity of external genitalia and are frequently raised as girls. However, a notable change occurred at puberty during which they developed a typical male phenotype, including virilisation of ambiguous genitalia into a functional penis and male psychosexual orientation regardless of prior female designation and rearing (Imperato-McGinley et al., 1974; Imperato-McGinley & Zhu, 2002)?. In adulthood this cohort display little body hair, minimal beard growth, no hairline recession, no acne and significantly smaller prostates. Finasteride clinical research and development leads viewed genetic 5ar2 deficiency as a predictive model for the chronic inhibition of the 5ar2 enzyme in the adult male (Stoner, 1990)? and that enzymatic inhibition with finasteride would mimic a genetic 5ar2 deficiency (GORMLEY et al., 1990)?. Without consideration as to the devastating outcomes for a subset of consumers, finasteride appears to be tolerated in most men.

Finasteride exhibits a highly unusual and nonlinear dose-response. Maximum DHT suppression is achieved after a single 1mg dose. It is markedly suppressive of DHT at all daily doses between 0.04 and 100mg over two weeks. Steady-state DHT levels were reduced to between 0.1-0.15ng/ml at all doses tested by Gormley et al, with DHT levels returning to pre-treatment levels within 14 days of cessation (GORMLEY et al., 1990)?. 0.05 to 5mg finasteride produces a 60% reduction in DHT in scalp skin. Similarly, a dose of approximately 0.2 mg of finasteride is not appreciably different to 5mg in terms of serum DHT reduction, suggesting this drug is profoundly effective at low doses (Frankel, 1999)?. Preferentially binding to the 5ar2 enzyme though with a notable lesser effect on 5ar1, finasteride is a pseudo-irreversible mechanism-based inhibitor that is exceptionally potent, specific, and unusually

efficient. The enzyme-bound inhibitor complex follows parallel reaction coordinates that proceed through closely related enolate intermediates as testosterone's reduction to DHT, with the two reactions proving divergent in the final step, as detailed by Bull et al. (Bull et al., 1996).

No meaningful assessment of lasting sexual dysfunction was published during the clinical development of finasteride or dutasteride (Kiguradze et al., 2017). Meta-analysis of 34 clinical trials of Finasteride for use in androgenic alopecia discovered serious flaws, poor quality reporting and systematic bias (Belknap et al., 2015). None of the 34 articles considered had adequate safety reporting. Of 25 clinical trial reports with a control arm, none reported on blinding adequacy. 18 publications (53%) disclosed authors with conflicts of interest, while 19 articles (56%) received funding from a pharmaceutical manufacturer of finasteride. 12 articles (35%) did not disclose their funding. Nonsexual adverse drug events were not reported in 28 articles. One report found a clinically and statistically significant increase in Beck Depression Inventory Scores after exposure to finasteride but did not adequately assess adverse effects other than depression. Noting the flaws in reporting raised by Belknap, Lee et al. meta-analysed fifteen trials and nevertheless concluded a 1.55 fold increased risk of sexual dysfunction including erectile dysfunction, loss of libido and ejaculatory dysfunction with oral use of finasteride (Lee et al., 2018).

The typical physiological response to finasteride in animals and humans is not sufficient to account for PFS, its remarkable dose-independent severity, or the common worsening and progression following withdrawal. However, it is important that significant basic science evidence illustrates finasteride interacts with the broad physiological systems affected in PFS (Irwig & Kolukula, 2011). Use of finasteride is an identified risk factor for male infertility (Samplaski et al., 2019) and has been associated with a variably reversible depletion in sperm count in humans at 5mg and 1mg doses (Amory et al., 2007; Samplaski et al., 2013). Recent animal research reveals not only lasting decreases in fertility parameters in finasteride exposed animals (Garcia et al., 2012), but a negative impact on the fertility of the next generation (Kolasa-Wolosiuk et al., 2015; Kolasa-Wolosiuk et al., 2018, 2019). Reduced androgen levels in the offspring of finasteride treated adult male rats have been noted as similar to those reported in studies exploring the effects of prenatal exposure to the antiandrogenic endocrine disruptors flutamide and vinclozolin (Kolasa-Wolosiuk et al., 2019; Ostby et al., 1999). In gerbils, low doses of Finasteride have been demonstrated to cause structural alterations in the prostates of both sexes, as well as lasting upregulation of the AR in the prostate epithelium of intrauterine exposed males, suggested to be a compensatory response to the low available DHT (Maldarine et al., 2019). 5alpha reductase inhibition induces erectile dysfunction in rats that is not fully reversed by washout (Öztekin et al., 2012; Pinsky et al., 2011). Histopathological evidence of marked atrophic changes in prostatic epithelial tissues, loss of penile smooth muscle content and prominent collagen deposition in penile cavernosal tissues has been reported in rats treated with either Finasteride or Dutasteride (Sahin Kilic et al., 2018; Shen et al., 2003; Zhang et al., 2013), suggesting direct deleterious effects on the penis and on erectile function.

Rats subchronically treated with finasteride for 20 days showed depressive behaviour and hippocampal alterations one month after withdrawal (Diviccaro et al., 2019). Additionally, disruption of

neurosteroids and steroid receptors, including an upregulation of the AR in the cerebral cortex, persisted a month after 20 days of low-dose finasteride treatment in rats, suggesting lasting structural and functional consequences on brain function (Giatti et al., 2015). Finasteride has broad consequences upon the formation of centrally active steroids and neurosteroids (Soggiu et al., 2016; Abdulmaged M. Traish, 2018). Neurosteroids are important to a range of central functions including HPA regulation and their dysregulation has a determinant role in neuropsychological abnormalities (Belelli & Lambert, 2005; Calogero et al., 1998; Camille Melón & Maguire, 2016; Carver & Reddy, 2013; Maguire, 2019). Allopregnanolone, determined to be low in the central nervous system of PFS patients (Melcangi et al., 2017), has a known role in increasing neurogenesis and neuronal cell survival, as well as reducing cell death in the hippocampus and midbrain (Diotel et al., 2018). Low or absent allopregnanolone is associated with psychological pathology including Post-Traumatic Stress Disorder (PTSD) (Pineles et al., 2018) and major depressive disorder (Maguire, 2019). Finasteride is employed experimentally to abolish the formation of neuroactive steroids including allopregnanolone in models relevant to Tourette syndrome and PTSD (Cadeddu et al., 2019; Nagaya et al., 2015).

Page Bibliography

1. Amory, J. K., Wang, C., Swerdloff, R. S., Anawalt, B. D., Matsumoto, A. M., Bremner, W. J., Walker, S. E., Haberer, L. J., & Clark, R. V. (2007). The Effect of 5 α -Reductase Inhibition with Dutasteride and Finasteride on Semen Parameters and Serum Hormones in Healthy Men. *The Journal of Clinical Endocrinology & Metabolism*, 1659–1665. <https://doi.org/10.1210/jc.2006-2203>
2. Apparao, K. B. C., Lovely, L. P., Gui, Y., Lininger, R. A., & Lessey, B. A. (2002). Elevated Endometrial Androgen Receptor Expression in Women with Polycystic Ovarian Syndrome. *Biology of Reproduction*, 297–304. <https://doi.org/10.1095/biolreprod66.2.297>
3. Azzouni, F., Godoy, A., Li, Y., & Mohler, J. (2012). The 5 Alpha-Reductase Isozyme Family: A Review of Basic Biology and Their Role in Human Diseases. *Advances in Urology*, 1–18. <https://doi.org/10.1155/2012/530121>
4. Bagatell, C J, Heiman, J. R., Rivier, J. E., & Bremner, W. J. (1994). Effects of endogenous testosterone and estradiol on sexual behavior in normal young men. *The Journal of Clinical Endocrinology & Metabolism*, 711–716. <https://doi.org/10.1210/jcem.78.3.8126146>
5. Bagatell, Carrie J., & Bremner, W. J. (1996). Androgens in Men — Uses and Abuses. *New England Journal of Medicine*, 707–715. <https://doi.org/10.1056/nejm199603143341107>

6. Balthazart, J., Choleris, E., & Remage-Healey, L. (2018). Steroids and the brain: 50 years of research, conceptual shifts and the ascent of non-classical and membrane-initiated actions. *Hormones and Behavior*, 1–8. <https://doi.org/10.1016/j.yhbeh.2018.01.002>
7. Banerjee, P., Banerjee, S., Brown, T., & Zirkin, B. (2018). Androgen action in prostate function and disease. *American Journal of Clinical and Experimental Urology*, 6(2), 62–77. <https://www.ncbi.nlm.nih.gov/pubmed/29666834>
8. Belelli, D., & Lambert, J. J. (2005). Neurosteroids: endogenous regulators of the GABAA receptor. *Nature Reviews Neuroscience*, 565–575. <https://doi.org/10.1038/nrn1703>
9. Belknap, S. M., Aslam, I., Kiguradze, T., Temps, W. H., Yarnold, P. R., Cashy, J., Brannigan, R. E., Micali, G., Nardone, B., & West, D. P. (2015). Adverse Event Reporting in Clinical Trials of Finasteride for Androgenic Alopecia. *JAMA Dermatology*, 600. <https://doi.org/10.1001/jamadermatol.2015.36>
10. Beyenburg, S., Watzka, M., Clusmann, H., Blümcke, I., Bidlingmaier, F., Elger, C. E., & Stoffel-Wagner, B. (2000). Androgen receptor mRNA expression in the human hippocampus. *Neuroscience Letters*, 25–28. [https://doi.org/10.1016/s0304-3940\(00\)01542-1](https://doi.org/10.1016/s0304-3940(00)01542-1)
11. Brinkmann, A. O., Faber, P. W., van Rooij, H. C. J., Kuiper, G. G. J. M., Ris, C., Klaassen, P., van der Korput, J. A. G. M., Voorhorst, M. M., van Laar, J. H., Mulder, E., & Trapman, J. (1989). The human androgen receptor: Domain structure, genomic organization and regulation of expression. *Journal of Steroid Biochemistry*, 307–310. [https://doi.org/10.1016/0022-4731\(89\)90098-8](https://doi.org/10.1016/0022-4731(89)90098-8)
12. Brock, O., De Mees, C., & Bakker, J. (2015). Hypothalamic Expression of Oestrogen Receptor ? and Androgen Receptor is Sex-, Age- and Region-Dependent in Mice. *Journal of Neuroendocrinology*, 264–276. <https://doi.org/10.1111/jne.12258>
13. Bull, H. G., Garcia-Calvo, M., Andersson, S., Baginsky, W. F., Chan, H. K., Ellsworth, D. E., Miller, R. R., Stearns, R. A., Bakshi, R. K., Rasmusson, G. H., Tolman, R. L., Myers, R. W., Kozarich, J. W., & Harris, G. S. (1996). Mechanism-Based Inhibition of Human Steroid 5 α -Reductase by Finasteride: Enzyme-Catalyzed Formation of NADP⁺Dihydrofinasteride, a Potent Bisubstrate Analog Inhibitor. *Journal of the American Chemical Society*, 2359–2365. <https://doi.org/10.1021/ja953069t>
14. Cadeddu, R., Bäckström, T., Floris, G., Nordkild, P., Segerdahl, M., & Bortolato, M. (2019).

Isoallopregnanolone reduces tic-like behaviours in the D1 CT 7 mouse model of Tourette syndrome. *Journal of Neuroendocrinology*. <https://doi.org/10.1111/jne.12754>

15. Calogero, A., Palumbo, M., Bosboom, A., Burrello, N., Ferrara, E., Palumbo, G., Petraglia, F., & D'Agata, R. (1998). The neuroactive steroid allopregnanolone suppresses hypothalamic gonadotropin-releasing hormone release through a mechanism mediated by the gamma-aminobutyric acidA receptor. *Journal of Endocrinology*, 121–125. <https://doi.org/10.1677/joe.0.1580121>
16. Camille Melón, L., & Maguire, J. (2016). GABAergic regulation of the HPA and HPG axes and the impact of stress on reproductive function. *The Journal of Steroid Biochemistry and Molecular Biology*, 196–203. <https://doi.org/10.1016/j.jsbmb.2015.11.019>
17. Carver, C. M., & Reddy, D. S. (2013). Neurosteroid interactions with synaptic and extrasynaptic GABAA receptors: regulation of subunit plasticity, phasic and tonic inhibition, and neuronal network excitability. *Psychopharmacology*, 151–188. <https://doi.org/10.1007/s00213-013-3276-5>
18. Chang, C., & McDonnell, D. P. (2005). Androgen receptor–cofactor interactions as targets for new drug discovery. *Trends in Pharmacological Sciences*, 225–228. <https://doi.org/10.1016/j.tips.2005.03.002>
19. Dale, J. B., Sarich, S. L., Bretz, T. M., Hatton, J. F., & Zachow, R. J. (2002). Hormonal Regulation of Androgen Receptor Messenger Ribonucleic Acid Expression in Human Tooth Pulp. *Journal of Dental Research*, 360–365. <https://doi.org/10.1177/154405910208100514>
20. Davey, R., & Grossmann, M. (2016). Androgen Receptor Structure, Function and Biology: From Bench to Bedside. *The Clinical Biochemist. Reviews*, 37(1), 3–15. <https://www.ncbi.nlm.nih.gov/pubmed/27057074>
21. Deng, Q., Zhang, Z., Wu, Y., Yu, W., Zhang, J., Jiang, Z., Zhang, Y., Liang, H., & Gui, Y. (2017). Non-Genomic Action of Androgens is Mediated by Rapid Phosphorylation and Regulation of Androgen Receptor Trafficking. *Cellular Physiology and Biochemistry*, 223–236. <https://doi.org/10.1159/000480343>
22. Diotel, N., Charlier, T. D., Lefebvre d'Hellencourt, C., Couret, D., Trudeau, V. L., Nicolau, J. C., Meilhac, O., Kah, O., & Pellegrini, E. (2018). Steroid Transport, Local Synthesis, and Signaling within the Brain: Roles in Neurogenesis, Neuroprotection, and Sexual Behaviors. *Frontiers in Neuroscience*. <https://doi.org/10.3389/fnins.2018.00084>

23. Diviccaro, S., Giatti, S., Borgo, F., Barcella, M., Borghi, E., Trejo, J. L., Garcia-Segura, L. M., & Melcangi, R. C. (2019). Treatment of male rats with finasteride, an inhibitor of 5alpha-reductase enzyme, induces long-lasting effects on depressive-like behavior, hippocampal neurogenesis, neuroinflammation and gut microbiota composition. *Psychoneuroendocrinology*, 206–215. <https://doi.org/10.1016/j.psyneuen.2018.09.021>
24. Eftekharzadeh, B., Banduseela, V. C., Chiesa, G., Martínez-Cristóbal, P., Rauch, J. N., Nath, S. R., Schwarz, D. M. C., Shao, H., Marin-Argany, M., Di Sanza, C., Giorgetti, E., Yu, Z., Pierattelli, R., Felli, I. C., Brun-Heath, I., García, J., Nebreda, Á. R., Gestwicki, J. E., Lieberman, A. P., & Salvatella, X. (2019). Hsp70 and Hsp40 inhibit an inter-domain interaction necessary for transcriptional activity in the androgen receptor. *Nature Communications*. <https://doi.org/10.1038/s41467-019-11594-y>
25. Foradori, C. D., Weiser, M. J., & Handa, R. J. (2008). Non-genomic actions of androgens. *Frontiers in Neuroendocrinology*, 169–181. <https://doi.org/10.1016/j.yfrne.2007.10.005>
26. Forger, N. G. (2018). Past, present and future of epigenetics in brain sexual differentiation. *Journal of Neuroendocrinology*, e12492. <https://doi.org/10.1111/jne.12492>
27. Frankel, S. (1999). Study of the Food and Drug Administration Files on Propecia. *Archives of Dermatology*. <https://doi.org/10.1001/archderm.135.3.257>
28. Fujimoto, J., Nishigaki, M., Hori, M., Ichigo, S., Itoh, T., & Tamaya, T. (1994). The effect of estrogen and androgen on androgen receptors and mRNA levels in uterine leiomyoma, myometrium and endometrium of human subjects. *The Journal of Steroid Biochemistry and Molecular Biology*, 137–143. [https://doi.org/10.1016/0960-0760\(94\)90020-5](https://doi.org/10.1016/0960-0760(94)90020-5)
29. Gannon, A.-L., O'Hara, L., Mason, J. I., Jørgensen, A., Frederiksen, H., Milne, L., Smith, S., Mitchell, R. T., & Smith, L. B. (2019). Androgen receptor signalling in the male adrenal facilitates X-zone regression, cell turnover and protects against adrenal degeneration during ageing. *Scientific Reports*. <https://doi.org/10.1038/s41598-019-46049-3>
30. Garcia, P. V., Barbieri, M. F., Perobelli, J. E., Consonni, S. R., Mesquita, S. de F. P., Kempinas, W. de G., & Pereira, L. A. V. (2012). Morphometric-stereological and functional epididymal alterations and a decrease in fertility in rats treated with finasteride and after a 30-day post-treatment recovery period. *Fertility and Sterility*, 1444–1451. <https://doi.org/10.1016/j.fertnstert.2012.03.025>

31. Geniole, S. N., Procyshyn, T. L., Marley, N., Ortiz, T. L., Bird, B. M., Marcellus, A. L., Welker, K. M., Bonin, P. L., Goldfarb, B., Watson, N. V., & Carré, J. M. (2019). Using a Psychopharmacogenetic Approach To Identify the Pathways Through Which—and the People for Whom—Testosterone Promotes Aggression. *Psychological Science*, 481–494. <https://doi.org/10.1177/0956797619826970>
32. Giatti, S., Foglio, B., Romano, S., Pesaresi, M., Panzica, G., Garcia-Segura, L. M., Caruso, D., & Melcangi, R. C. (2015). Effects of Subchronic Finasteride Treatment and Withdrawal on Neuroactive Steroid Levels and Their Receptors in the Male Rat Brain. *Neuroendocrinology*, 746–757. <https://doi.org/10.1159/000442982>
33. Gibson, D. A., Saunders, P. T. K., & McEwan, I. J. (2018). Androgens and androgen receptor: Above and beyond. *Molecular and Cellular Endocrinology*, 1–3. <https://doi.org/10.1016/j.mce.2018.02.013>
34. Gil, D., Zarzycka, M., Dulińska-Litewka, J., Ciończyk-Wierzbicka, D., Lekka, M., & Laidler, P. (2019). Dihydrotestosterone increases the risk of bladder cancer in men. *Human Cell*, 379–389. <https://doi.org/10.1007/s13577-019-00255-3>
35. GORMLEY, G. J., STONER, E., RITTMASER, R. S., GREGG, H., THOMPSON, D. L., LASSETER, K. C., VLASSES, P. H., & STEIN, E. A. (1990). Effects of Finasteride (MK-906), a 5 α -Reductase Inhibitor, on Circulating Androgens in Male Volunteers*. *The Journal of Clinical Endocrinology & Metabolism*, 1136–1141. <https://doi.org/10.1210/jcem-70-4-1136>
36. Hamilton, J. B. (1942). Male hormone stimulation is prerequisite and an incitant in common baldness. *American Journal of Anatomy*, 451–480. <https://doi.org/10.1002/aja.1000710306>
37. Heemers, H. V., & Tindall, D. J. (2007). Androgen Receptor (AR) Coregulators: A Diversity of Functions Converging on and Regulating the AR Transcriptional Complex. *Endocrine Reviews*, 778–808. <https://doi.org/10.1210/er.2007-0019>
38. Heinlein, C. A., & Chang, C. (2002). Androgen Receptor (AR) Coregulators: An Overview. *Endocrine Reviews*, 175–200. <https://doi.org/10.1210/edrv.23.2.0460>
39. Hunter, I., Hay, C. W., Esswein, B., Watt, K., & McEwan, I. J. (2018). Tissue control of androgen action: The ups and downs of androgen receptor expression. *Molecular and Cellular*

Endocrinology, 27–35. <https://doi.org/10.1016/j.mce.2017.08.002>

40. Hur, E., Pfaff, S. J., Payne, E. S., Grøn, H., Buehrer, B. M., & Fletterick, R. J. (2004). Recognition and Accommodation at the Androgen Receptor Coactivator Binding Interface. *PLoS Biology*, e274. <https://doi.org/10.1371/journal.pbio.0020274>
41. Imperato-McGinley, J., Guerrero, L., Gautier, T., & Peterson, R. E. (1974). Steroid 5 α -Reductase Deficiency in Man: An Inherited Form of Male Pseudohermaphroditism. *Science*, 1213–1215. <https://doi.org/10.1126/science.186.4170.1213>
42. Imperato-McGinley, J., Miller, M., Wilson, J. D., Peterson, R. E., Shackleton, C., & Gajdusek, D. C. (1991). A cluster of male pseudohermaphrodites with 5 α -reductase deficiency in Papua New Guinea. *Clinical Endocrinology*, 293–298. <https://doi.org/10.1111/j.1365-2265.1991.tb03769.x>
43. Imperato-McGinley, J., & Zhu, Y.-S. (2002). Androgens and male physiology the syndrome of 5 α -reductase-2 deficiency. *Molecular and Cellular Endocrinology*, 51–59. [https://doi.org/10.1016/s0303-7207\(02\)00368-4](https://doi.org/10.1016/s0303-7207(02)00368-4)
44. Irwig, M. S., & Kolukula, S. (2011). Persistent Sexual Side Effects of Finasteride for Male Pattern Hair Loss. *The Journal of Sexual Medicine*, 1747–1753. <https://doi.org/10.1111/j.1743-6109.2011.02255.x>
45. J. Brand, L., & M. Dehm, S. (2013). Androgen Receptor Gene Rearrangements: New Perspectives on Prostate Cancer Progression. *Current Drug Targets*, 441–449. <https://doi.org/10.2174/1389450111314040005>
46. Kalyvianaki, K., Panagiotopoulos, A. A., Malamos, P., Moustou, E., Tzardi, M., Stathopoulos, E. N., Ioannidis, G. S., Marias, K., Notas, G., Theodoropoulos, P. A., Castanas, E., & Kampa, M. (2019). Membrane androgen receptors (OXER1, GPRC6A AND ZIP9) in prostate and breast cancer: A comparative study of their expression. *Steroids*, 100–108. <https://doi.org/10.1016/j.steroids.2019.01.006>
47. Khalil, R., Kim, N. R., Jardi, F., Vanderschueren, D., Claessens, F., & Decallonne, B. (2018). Sex steroids and the kidney: role in renal calcium and phosphate handling. *Molecular and Cellular Endocrinology*, 61–72. <https://doi.org/10.1016/j.mce.2017.11.011>
48. Kiguradze, T., Temps, W. H., Yarnold, P. R., Cashy, J., Brannigan, R. E., Nardone, B., Micali, G.,

- West, D. P., & Belknap, S. M. (2017). Persistent erectile dysfunction in men exposed to the 5 α -reductase inhibitors, finasteride, or dutasteride. *PeerJ*, e3020. <https://doi.org/10.7717/peerj.3020>
49. Kimura, N., Mizokami, A., Oonuma, T., Sasano, H., & Nagura, H. (1993). Immunocytochemical localization of androgen receptor with polyclonal antibody in paraffin-embedded human tissues. *Journal of Histochemistry & Cytochemistry*, 671–678. <https://doi.org/10.1177/41.5.8468448>
50. King-Jones, K., & Thummel, C. S. (2005). Nuclear receptors — a perspective from *Drosophila*. *Nature Reviews Genetics*, 311–323. <https://doi.org/10.1038/nrg1581>
51. Kolasa-Wołoskiuk, A., Misiakiewicz-Has, K., Baranowska-Bosiacka, I., Gutowska, I., Tarnowski, M., Tkacz, M., & Wiszniewska, B. (2018). Connexin 43 expression in the testes during postnatal development of finasteride-treated male rat offspring. *Archives of Medical Science*, 1471–1479. <https://doi.org/10.5114/aoms.2016.63022>
52. Kolasa-Wołoskiuk, A., Misiakiewicz-Has, K., Baranowska-Bosiacka, I., Gutowska, I., & Wiszniewska, B. (2015). Androgen levels and apoptosis in the testis during postnatal development of finasteride-treated male rat offspring. *Folia Histochemica et Cytobiologica*, 53(3), 236–248. <https://doi.org/10.5603/fhc.a2015.0025>
53. Kolasa-Wołoskiuk, A., Tarnowski, M., Baranowska-Bosiacka, I., Chlubek, D., & Wiszniewska, B. (2019). Antioxidant enzyme expression of mRNA and protein in the epididymis of finasteride-treated male rat offspring during postnatal development. *Archives of Medical Science*, 797–810. <https://doi.org/10.5114/aoms.2017.68528>
54. Koryakina, Y., Ta, H. Q., & Gioeli, D. (2014). Androgen receptor phosphorylation: biological context and functional consequences. *Endocrine-Related Cancer*, T131–T145. <https://doi.org/10.1530/erc-13-0472>
55. Kumar, R., & McEwan, I. J. (2012). Allosteric Modulators of Steroid Hormone Receptors: Structural Dynamics and Gene Regulation. *Endocrine Reviews*, 271–299. <https://doi.org/10.1210/er.2011-1033>
56. Lai, J.-J., Chang, P., Lai, K.-P., Chen, L., & Chang, C. (2012). The role of androgen and androgen receptor in skin-related disorders. *Archives of Dermatological Research*, 499–510. <https://doi.org/10.1007/s00403-012-1265-x>

57. Lanciotti, L., Cofini, M., Leonardi, A., Bertozzi, M., Penta, L., & Esposito, S. (2019). Different Clinical Presentations and Management in Complete Androgen Insensitivity Syndrome (CAIS). *International Journal of Environmental Research and Public Health*, 1268. <https://doi.org/10.3390/ijerph16071268>
58. Lee, S., Lee, Y., Choe, S., & Lee, W. (2018). Adverse Sexual Effects of Treatment with Finasteride or Dutasteride for Male Androgenetic Alopecia: A Systematic Review and Meta-analysis. *Acta Dermato Venereologica*, 0. <https://doi.org/10.2340/00015555-3035>
59. Li, J., Fu, X., Cao, S., Li, J., Xing, S., Li, D., Dong, Y., Cardin, D., Park, H.-W., Mauvais-Jarvis, F., & Zhang, H. (2018). Membrane-associated androgen receptor (AR) potentiates its transcriptional activities by activating heat shock protein 27 (HSP27). *Journal of Biological Chemistry*, 12719–12729. <https://doi.org/10.1074/jbc.ra118.003075>
60. Liu, Y., Ding, M., Liao, X., Gao, Q., He, A., Liu, B., Hu, K., Xie, H., Zhou, Q., Zhan, H., Liu, Y., Huang, W., & Mei, H. (2018). High expression of enhancer RNA MARC1 or its activation by DHT is associated with the malignant behavior in bladder cancer. *Experimental Cell Research*, 303–311. <https://doi.org/10.1016/j.yexcr.2018.06.032>
61. Lu, S., McKenna, S. E., Cologer-Clifford, A., Nau, E. A., & Simon, N. G. (1998). Androgen Receptor in Mouse Brain: Sex Differences and Similarities in Autoregulation1. *Endocrinology*, 1594–1601. <https://doi.org/10.1210/endo.139.4.5863>
62. Lubahn, D., Joseph, D., Sullivan, P., Willard, H., French, F., & Wilson, E. (1988). Cloning of human androgen receptor complementary DNA and localization to the X chromosome. *Science*, 327–330. <https://doi.org/10.1126/science.3353727>
63. Maguire, J. (2019). Neuroactive Steroids and GABAergic Involvement in the Neuroendocrine Dysfunction Associated With Major Depressive Disorder and Postpartum Depression. *Frontiers in Cellular Neuroscience*. <https://doi.org/10.3389/fncel.2019.00083>
64. Maldarine, J. S., Sanches, B. D. A., Santos, V. A., Amaro, G. M., Calmon, M. F., Rahal, P., Góes, R. M., Vilamaior, P. S. L., & Taboga, S. R. (2019). Low-dose in utero exposure to finasteride promotes developmental changes in both male and female gerbil prostates. *Environmental Toxicology*, 15–26. <https://doi.org/10.1002/tox.22838>
65. Mangelsdorf, D. J., Thummel, C., Beato, M., Herrlich, P., Schütz, G., Umesono, K., Blumberg, B., Kastner, P., Mark, M., Chambon, P., & Evans, R. M. (1995). The nuclear receptor

superfamily: The second decade. *Cell*, 835–839. [https://doi.org/10.1016/0092-8674\(95\)90199-x](https://doi.org/10.1016/0092-8674(95)90199-x)

66. McEwan, I J. (2004). Molecular mechanisms of androgen receptor-mediated gene regulation: structure-function analysis of the AF-1 domain. *Endocrine-Related Cancer*, 281–293. <https://doi.org/10.1677/erc.0.0110281>
67. McEwan, I., & Monaghan, A. (2016). A sting in the tail: the N-terminal domain of the androgen receptor as a drug target. *Asian Journal of Andrology*, 687. <https://doi.org/10.4103/1008-682x.181081>
68. McEwan, Iain J., & Kumar, R. (2015). Twenty-five Years of Nuclear Receptor Structure Analysis: From the Laboratory to the Clinic. In *Nuclear Receptors: From Structure to the Clinic* (pp. 1–14). Springer International Publishing. https://doi.org/10.1007/978-3-319-18729-7_1
69. Melcangi, R. C., Santi, D., Spezzano, R., Grimoldi, M., Tabacchi, T., Fusco, M. L., Diviccaro, S., Giatti, S., Carrà, G., Caruso, D., Simoni, M., & Cavaletti, G. (2017). Neuroactive steroid levels and psychiatric and andrological features in post-finasteride patients. *The Journal of Steroid Biochemistry and Molecular Biology*, 229–235. <https://doi.org/10.1016/j.jsbmb.2017.04.003>
70. Mhaouty-Kodja, S. (2018). Role of the androgen receptor in the central nervous system. *Molecular and Cellular Endocrinology*, 103–112. <https://doi.org/10.1016/j.mce.2017.08.001>
71. Morford, J. J., Wu, S., & Mauvais-Jarvis, F. (2018). The impact of androgen actions in neurons on metabolic health and disease. *Molecular and Cellular Endocrinology*, 92–102. <https://doi.org/10.1016/j.mce.2017.09.001>
72. Nagaya, N., Acca, G. M., & Maren, S. (2015). Allopregnanolone in the bed nucleus of the stria terminalis modulates contextual fear in rats. *Frontiers in Behavioral Neuroscience*. <https://doi.org/10.3389/fnbeh.2015.00205>
73. Narayanan, R., Coss, C. C., & Dalton, J. T. (2018). Development of selective androgen receptor modulators (SARMs). *Molecular and Cellular Endocrinology*, 134–142. <https://doi.org/10.1016/j.mce.2017.06.013>
74. Nelson, W. G. (2016). Commentary on Huggins and Hodges: “Studies on Prostatic Cancer.” *Cancer Research*, 186–187. <https://doi.org/10.1158/0008-5472.can-15-3172>

75. Nguyen, P. L., Alibhai, S. M. H., Basaria, S., D'Amico, A. V., Kantoff, P. W., Keating, N. L., Penson, D. F., Rosario, D. J., Tombal, B., & Smith, M. R. (2015). Adverse Effects of Androgen Deprivation Therapy and Strategies to Mitigate Them. *European Urology*, 825–836. <https://doi.org/10.1016/j.eururo.2014.07.010>
76. Ni, L., Llewellyn, R., Kesler, C. T., Kelley, J. B., Spencer, A., Snow, C. J., Shank, L., & Paschal, B. M. (2013). Androgen Induces a Switch from Cytoplasmic Retention to Nuclear Import of the Androgen Receptor. *Molecular and Cellular Biology*, 4766–4778. <https://doi.org/10.1128/mcb.00647-13>
77. Ostby, J., Kelce, W. R., Lambright, C., Wolf, C. J., Mann, P., & Gray, L. E., Jr. (1999). The fungicide procymidone alters sexual differentiation in the male rat by acting as an androgen-receptor antagonist in vivo and in vitro. *Toxicology and Industrial Health*, 80–93. <https://doi.org/10.1177/074823379901500108>
78. Öztekin, Ç. V., Gur, S., Abdulkadir, N. A., Lokman, U., Akdemir, A. Ö., Cetinkaya, M., & Hellstrom, W. J. G. (2012). Incomplete Recovery of Erectile Function in Rat after Discontinuation of Dual 5 α Reductase Inhibitor Therapy. *The Journal of Sexual Medicine*, 1773–1781. <https://doi.org/10.1111/j.1743-6109.2012.02759.x>
79. Pineles, S. L., Nillni, Y. I., Pinna, G., Irvine, J., Webb, A., Arditte Hall, K. A., Hauger, R., Miller, M. W., Resick, P. A., Orr, S. P., & Rasmusson, A. M. (2018). PTSD in women is associated with a block in conversion of progesterone to the GABAergic neurosteroids allopregnanolone and pregnanolone measured in plasma. *Psychoneuroendocrinology*, 133–141. <https://doi.org/10.1016/j.psyneuen.2018.04.024>
80. Pinsky, M. R., Gur, S., Tracey, A. J., Harbin, A., & Hellstrom, W. J. G. (2011). The Effects of Chronic 5 α Reductase Inhibitor (Dutasteride) Treatment on Rat Erectile Function. *The Journal of Sexual Medicine*, 3066–3074. <https://doi.org/10.1111/j.1743-6109.2011.02425.x>
81. Podlasek, C. A., Mulhall, J., Davies, K., Wingard, C. J., Hannan, J. L., Bivalacqua, T. J., Musicki, B., Khera, M., González-Cadavid, N. F., & Burnett, A. L., II. (2016). Translational Perspective on the Role of Testosterone in Sexual Function and Dysfunction. *The Journal of Sexual Medicine*, 1183–1198. <https://doi.org/10.1016/j.jsxm.2016.06.004>
82. Rainey, W. E., Carr, B. R., Sasano, H., Suzuki, T., & Mason, J. I. (2002). Dissecting human adrenal androgen production. *Trends in Endocrinology & Metabolism*, 234–239. [https://doi.org/10.1016/s1043-2760\(02\)00609-4](https://doi.org/10.1016/s1043-2760(02)00609-4)

83. Ruizeveld de Winter, J. A., Trapman, J., Vermey, M., Mulder, E., Zegers, N. D., & van der Kwast, T. H. (1991). Androgen receptor expression in human tissues: an immunohistochemical study. *Journal of Histochemistry & Cytochemistry*, 927–936.
<https://doi.org/10.1177/39.7.1865110>
84. Sahin Kilic, Engin Kolukcu, Fikret Erdemir, Ismail Benli, & Akgul Arici. (2018). The Effects of Oral 5-alpha Reductase Inhibitors on Penile Intracavernosal Pressures and Penile Morphology in Rat Model. *Urology Journal*. <https://doi.org/10.22037/uj.v0i0.4164>
85. Samplaski, M. K., Lo, K., Grober, E., & Jarvi, K. (2013). Finasteride use in the male infertility population: effects on semen and hormone parameters. *Fertility and Sterility*, 1542–1546.
<https://doi.org/10.1016/j.fertnstert.2013.07.2000>
86. Samplaski, M. K., Smith, J. F., Lo, K. C., Hotaling, J. M., Lau, S., Grober, E. D., Trussell, J. C., Walsh, T. J., Kolettis, P. N., Chow, V. D. W., Zini, A. S., Spitz, A., Fischer, M. A., Domes, T., Zeitlin, S. I., Fuchs, E. F., Hedges, J. C., Sandlow, J. I., Brannigan, R. E., ... Jarvi, K. A. (2019). Reproductive endocrinologists are the gatekeepers for male infertility care in North America: results of a North American survey on the referral patterns and characteristics of men presenting to male infertility specialists for infertility investigations. *Fertility and Sterility*, 657–662.
<https://doi.org/10.1016/j.fertnstert.2019.06.011>
87. Schiffer, L., Arlt, W., & Storbeck, K.-H. (2018). Intracrine androgen biosynthesis, metabolism and action revisited. *Molecular and Cellular Endocrinology*, 4–26.
<https://doi.org/10.1016/j.mce.2017.08.016>
88. Schultheiss, D., Badalyan, R., Pilatz, A., Gabouev, A. I., Schlote, N., Wefer, J., von Wasielewski, R., Mertsching, H., Sohn, M., Stief, C. G., & Jonas, U. (2003). Androgen and estrogen receptors in the human corpus cavernosum penis: immunohistochemical and cell culture results. *World Journal of Urology*, 320–324. <https://doi.org/10.1007/s00345-003-0371-y>
89. Shah, N. M., Pisapia, D. J., Maniatis, S., Mendelsohn, M. M., Nemes, A., & Axel, R. (2004). Visualizing Sexual Dimorphism in the Brain. *Neuron*, 313–319.
<https://doi.org/10.1016/j.neuron.2004.07.008>
90. Shen, Z., Zhou, X., Lu, Y., & Chen, Z. (2003). Effect of androgen deprivation on penile ultrastructure. *Asian Journal of Andrology*, 5(1), 33–36.
<https://www.ncbi.nlm.nih.gov/pubmed/12647000>

91. Simerly, R. B., Swanson, L. W., Chang, C., & Muramatsu, M. (1990). Distribution of androgen and estrogen receptor mRNA-containing cells in the rat brain: An in situ hybridization study. *The Journal of Comparative Neurology*, 76–95. <https://doi.org/10.1002/cne.902940107>
92. Sinha-Hikim, I., Taylor, W. E., Gonzalez-Cadavid, N. F., Zheng, W., & Bhasin, S. (2004). Androgen Receptor in Human Skeletal Muscle and Cultured Muscle Satellite Cells: Up-Regulation by Androgen Treatment. *The Journal of Clinical Endocrinology & Metabolism*, 5245–5255. <https://doi.org/10.1210/jc.2004-0084>
93. Slagsvold, T., Kraus, I., Frønsdal, K., & Saatcioglu, F. (2001). DNA Binding-independent Transcriptional Activation by the Androgen Receptor through Triggering of Coactivators. *Journal of Biological Chemistry*, 31030–31036. <https://doi.org/10.1074/jbc.m104310200>
94. Soggiu, A., Piras, C., Greco, V., Devoto, P., Urbani, A., Calzetta, L., Bortolato, M., & Roncada, P. (2016). Exploring the neural mechanisms of finasteride: a proteomic analysis in the nucleus accumbens. *Psychoneuroendocrinology*, 387–396. <https://doi.org/10.1016/j.psyneuen.2016.10.001>
95. Stoner, E. (1990). The clinical development of a 5 α -reductase inhibitor, finasteride. *The Journal of Steroid Biochemistry and Molecular Biology*, 375–378. [https://doi.org/10.1016/0960-0760\(90\)90487-6](https://doi.org/10.1016/0960-0760(90)90487-6)
96. Swerdloff, R. S., Dudley, R. E., Page, S. T., Wang, C., & Salameh, W. A. (2017). Dihydrotestosterone: Biochemistry, Physiology, and Clinical Implications of Elevated Blood Levels. *Endocrine Reviews*, 220–254. <https://doi.org/10.1210/er.2016-1067>
97. Thomas, P. (2019). Membrane Androgen Receptors Unrelated to Nuclear Steroid Receptors. *Endocrinology*, 772–781. <https://doi.org/10.1210/en.2018-00987>
98. Thomas, P., Converse, A., & Berg, H. A. (2018). ZIP9, a novel membrane androgen receptor and zinc transporter protein. *General and Comparative Endocrinology*, 130–136. <https://doi.org/10.1016/j.ygcen.2017.04.016>
99. Thomas, P., & Dong, J. (2019). Novel mechanism of endocrine disruption by fungicides through binding to the membrane androgen receptor, ZIP9 (SLC39A9), and antagonizing rapid testosterone induction of the intrinsic apoptotic pathway. *Steroids*, 108415. <https://doi.org/10.1016/j.steroids.2019.05.007>

100. Tobiansky, D. J., Wallin-Miller, K. G., Floresco, S. B., Wood, R. I., & Soma, K. K. (2018). Androgen Regulation of the Mesocorticolimbic System and Executive Function. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2018.00279>

101. Traish, A. M. (2008). Androgens Play a Pivotal Role in Maintaining Penile Tissue Architecture and Erection: A Review. *Journal of Andrology*, 363–369. <https://doi.org/10.2164/jandrol.108.006007>

102. Traish, Abdulmaged M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>

103. Traish, Abdulmaged M., Melcangi, R. C., Bortolato, M., Garcia-Segura, L. M., & Zitzmann, M. (2015). Adverse effects of 5 α -reductase inhibitors: What do we know, don't know, and need to know? *Reviews in Endocrine and Metabolic Disorders*, 177–198. <https://doi.org/10.1007/s11154-015-9319-y>

104. Traish, Abdulmaged M., Mulgaonkar, A., & Giordano, N. (2014). The Dark Side of 5 α -Reductase Inhibitors' Therapy: Sexual Dysfunction, High Gleason Grade Prostate Cancer and Depression. *Korean Journal of Urology*, 367. <https://doi.org/10.4111/kju.2014.55.6.367>

105. Tsai, M., & O'Malley, B. W. (1994). Molecular Mechanisms of Action of Steroid/Thyroid Receptor Superfamily Members. *Annual Review of Biochemistry*, 451–486. <https://doi.org/10.1146/annurev.bi.63.070194.002315>

106. van Royen, M. E., van Cappellen, W. A., de Vos, C., Houtsmuller, A. B., & Trapman, J. (2012). Stepwise androgen receptor dimerization. *Journal of Cell Science*, 1970–1979. <https://doi.org/10.1242/jcs.096792>

107. Vanderschueren, D., Laurent, M. R., Claessens, F., Gielen, E., Lagerquist, M. K., Vandeput, L., Börjesson, A. E., & Ohlsson, C. (2014). Sex Steroid Actions in Male Bone. *Endocrine Reviews*, 906–960. <https://doi.org/10.1210/er.2014-1024>

108. Verhoeven, G., & Swinnen, J. V. (1999). Indirect mechanisms and cascades of androgen action. *Molecular and Cellular Endocrinology*, 205–212. [https://doi.org/10.1016/s0303-7207\(99\)00014-3](https://doi.org/10.1016/s0303-7207(99)00014-3)

109. Wadosky, K. M., & Koochekpour, S. (2016). Therapeutic Rationales, Progresses, Failures, and Future Directions for Advanced Prostate Cancer. *International Journal of Biological Sciences*, 409–426. <https://doi.org/10.7150/ijbs.14090>
110. Wu, J., Henning, P., Sjögren, K., Koskela, A., Tuukkanen, J., Movérare-Skrtic, S., & Ohlsson, C. (2019). The androgen receptor is required for maintenance of bone mass in adult male mice. *Molecular and Cellular Endocrinology*, 159–169. <https://doi.org/10.1016/j.mce.2018.10.008>
111. Xia, T., Sun, H., Huang, H., Bi, H., Pu, R., Zhang, L., Zhang, Y., Liu, Y., Xu, J., Onwuka, J. U., Liu, Y., Cui, B., & Zhao, Y. (2019). Androgen receptor gene methylation related to colorectal cancer risk. *Endocrine Connections*, 979–987. <https://doi.org/10.1530/ec-19-0122>
112. Yamada, G., Suzuki, K., Haraguchi, R., Miyagawa, S., Satoh, Y., Kamimura, M., Nakagata, N., Kataoka, H., Kuroiwa, A., & Chen, Y. (2006). Molecular genetic cascades for external genitalia formation: An emerging organogenesis program. *Developmental Dynamics*, 1738–1752. <https://doi.org/10.1002/dvdy.20807>
113. Zhang, M.-G., Wang, X.-J., Shen, Z.-J., & Gao, P.-J. (2013). Long-term Oral Administration of 5 α -reductase Inhibitor Attenuates Erectile Function by Inhibiting Autophagy and Promoting Apoptosis of Smooth Muscle Cells in Corpus Cavernosum of Aged Rats. *Urology*, 743.e9-743.e15. <https://doi.org/10.1016/j.urology.2013.02.045>
-

PFS: Manifestation of a Post-Androgen Deprivation Syndrome following exposure to substances with antiandrogenic effects

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/pfs-manifestation-of-a-post-androgen-deprivation-syndrome-following-exposure-to-substances-with-antiandrogenic-effects/>

Endocrine disruption

"Endocrine disruption" refers to a specific toxicity whereby natural and/or anthropogenic chemicals cause adverse health effects by disrupting the endogenous hormone system. An endocrine disruptor is defined by the World Health Organisation as "an exogenous substance or mixture that alters the function(s) of the endocrine system and consequently causes adverse effects in an intact organism, or its progeny, or (sub) populations". Potential endocrine disruptors can act on hormone receptors directly or interfere with proteins mediating hormonal delivery to target tissues and cells. They may act at low doses, exhibit non-monotonic dose-response relationships, cause tissue specific effects and differing endpoints (Bergman et al., 2012). There is broad potential for pharmac/toxicodynamic influences from EDCs including alteration of receptor expression and interruption of the critical and complex feedback mechanisms regulating the endocrine system (Lagarde et al., 2015). It has been estimated that, in the EU, the cost associated with disease and disability reasonably attributable to EDC exposure is €157 billion, 1.23% of the European Union's gross domestic product (Trasande et al., 2015). Health risks related to exposure to endocrine disruptors are typically underestimated and poorly characterised (Fucic et al., 2018).

There is now scientific consensus that, as well as disruptive effects during developmental windows, interference with the role of hormones during maintenance of physiological function in adult life can cause adverse effects (Solecki et al., 2016). An adverse effect in this context constitutes "a change in morphology, physiology, growth, reproduction, development or lifespan of an organism which results in impairment of functional capacity or impairment of capacity to compensate for additional stress or increased susceptibility to the harmful effects of other environmental influences" (Bergman et al., 2012). In this context, anthropogenic chemicals can represent pervasive environmental stressors (Latchney et al., 2017), and the marked sensitivity to endocrine-affecting substances common in PFS patients appears to us to be a manifestation of this increased susceptibility. Changes to the epigenome that can persist indefinitely after exposure to pharmaceutical products is an increasing area of consideration (Csoka & Szyf, 2009). Recent publications centring on epigenetics increasingly appreciate Finasteride in the context of endocrine disruptors, with respect to both PFS (Traish, 2018) and in broader animal studies. Finasteride induces hypospadias and a permanent reduction in anogenital distance in adult male rats exposed during late gestation (Bowman et al., 2003). This effect on LABC weight is consistent with the effects of other antiandrogens such as flutamide, procymidone, vinclozolin, and linuron

?(McIntyre, 2001, 2002; Ostby et al., 1999; Wolf et al., 1999)?.

Despite the "clear endocrine disrupting activity" of 5-alpha reductase inhibitors, there is a paucity of information regarding the impact of non-clinical 5-alpha reductase inhibition (Patisaul & Belcher, 2017)?. Even with sole consideration of the known effects of very low doses of finasteride on development, the strong persistence of the drug in the environment and high photostability raises serious concerns about its widespread availability (Sammartino et al., 2013)?. The profound and devastating changes to physiological health manifesting as PFS in an adult subpopulation of fertile age following exposure to as little as 0.2mg of Finasteride should add significant and urgent public health concerns regarding its environmental toxicity as an EDC.

"PFS" following therapeutic use and cessation of other substances?

Importantly, patients are increasingly presenting to us suffering what is ostensibly clinically "post-finasteride syndrome" following use of drugs and substances including Isotretinoin, Serenoa repens (saw palmetto) extract, SSRI antidepressants, topical ketoconazole, topical minoxidil, and high-dose phenolic compounds marketed as health supplements including quercetin and milk thistle extract.

It is recognised that the syndrome termed Post-SSRI Sexual Dysfunction (PSSD) and PFS may share an etiological link. With focus on neurological symptoms, Giatti et al. presented a hypothesis that the impairment of overlapping signals of neuroactive steroids, dopamine and serotonin as potentially underlying the condition(s) (Giatti et al., 2018)?. In another consideration of the potential for a single syndrome underlying these presentations, Healy et al. analysed 300 patient responses to structured questions provided by and submitted to rxrisk.org, an independent drug safety website. The cohort was comprised of patients suffering persistent sexual dysfunction following use of 5-ARIs, Isotretinoin and Serotonin Reuptake Inhibitors, with treatment duration ranging from a single dose to over 16 years. Overlap was seen in symptoms including ED, Libido loss, genital anaesthesia, difficulty achieving orgasm, pleasureless orgasm, premature ejaculation, emotional blunting, loss of nocturnal erections, penile or testicular pain, reduction of penis size, decreased testosterone, watery ejaculate, testicular atrophy, and other skin numbness. Across drug groups, the sexual dysfunction became markedly worse or even began after cessation of treatment in many instances. For all three drug groups there were reports of profound dysfunction appearing within days of stopping, while Finasteride and Isotretinoin are stopped abruptly, SSRIs are often tapered. Interestingly, three subjects on SSRIs reported an increasing loss of sexual function as the dose was tapered, suggesting that PSSD may be equally likely following abrupt or gradual discontinuation of an SSRI or SNRI. They conclude the need for comparative investigation in these cohorts and a systematic approach with structured symptom sets to establish the existence of a single syndrome (David Healy et al., 2018)?.

The antiandrogenic commonality of substances causing an ostensibly similar persistent syndrome

Accutane, Roaccutan, Generics (Isotretinoin)

Retinoids possess important antiandrogenic endocrine disrupting properties. Isotretinoin is a 3-cis-retinoic acid which is marketed under the brand name Accutane, Roaccutan, and as branded generic preparations. The main application is treatment of acne, which is strongly linked to androgenic activity in the skin (Melnik, 2017). Boudou et al. reported that after three months of isotretinoin treatment to six male patients with severe acne, complete resolution of acne was achieved in four patients and the remaining two patients improved significantly. No changes were recorded in serum testosterone but a significant decrease in DHT was observed. Androgen receptor status was investigated in back skin biopsies obtained in acne areas before and after three months of isotretinoin treatment. Treatment induced a 2.6-fold decrease in AR binding capacity constant (62 vs. 24 fmol/mg cytosolic protein), demonstrating a marked sensitivity of androgen receptor in the skin to oral isotretinoin. The authors concluded the data supported previous observations of DHT suppression and were consistent with the key role of the AR and DHT in acne, noting sebum is under androgen control and that androgen responsiveness of the pilosebaceous unit is implicated in acne pathogenesis (Boudou et al., 1995). Boudou et al. had previously illustrated that skin biopsies of eight men with severe acne treated with 3 months of isotretinoin "lost 80% of their ability to form 5 alpha-dihydrotestosterone (P <0.001)" (Boudou et al., 1994).

AR signal transduction is crucial to acne pathogenesis, stimulating the size of sebocytes and sebum production as well as proliferation of keratinocytes (Lai et al., 2012). IGF-1/PI3K/AKT-mediated inactivation of Forkhead box O1 (FoxO1) is vital to androgen receptor transactivation (Fan et al., 2007). FoxO1 is repressive of AR owing to FoxO1's inhibition of AR N/C terminal interaction (Q. Ma et al., 2009). IGF-1 has correlated to acne severity (Cappel, 2005) and isotretinoin decreases IGF-1 (A.S. Karadag et al., 2009). As IGF-1 is inhibitory of AR (Palazzolo et al., 2009; Yanase & Fan, 2009), Karadag et al. hypothesised that a consequential nuclear increase in FoxO1 would significantly contribute to the downregulation of AR and thus a decrease of androgen-responsive gene transcription (Ayse Serap Karadag et al., 2015). As with other anti-acne therapies, Isotretinoin enhances p53 expression (Melnik, 2017), which suppresses AR expression (Alimirah et al., 2007; Shenk et al., 2001). Additionally, p53 activates and increases FoxO1 expression (Pappas et al., 2017). Human primary keratinocytes treated with isotretinoin show an increase in FoxO1 (Shi et al., 2018), and significant increases in nuclear levels of FoxO1 protein are reported in skin biopsies from acne patients following isotretinoin treatment (Agamia et al., 2018). In vitro evidence demonstrates all-trans retinoic acid profoundly downregulates the AR and abolishes the induction of androgen-induced functions (Ubels et al., 2002, 2003), suggesting a common androgen antagonism among retinoids. Taken together, there is significant evidence

for the conclusion that oral Isotretinoin exerts a potent antiandrogenic effect.

Antidepressants

SSRI/SNRI class antidepressants exert significant antiandrogenic activity and have been associated with reproductive toxicity in male rats and humans (Atli et al., 2017; Ilgin et al., 2017; Tanrikut et al., 2010). Fluoxetine is known to be endocrine disruptive, with evidence of nonmonotonic effects (Cunha et al., 2018; Vandenberg et al., 2012). Rats administered Fluoxetine display delayed sexual development and decreased sexual behaviours (Drugs@FDA, 2016). Griffin and Mellon found the enzymatic efficiency of 3 β -HSD conversion of DHT to androstenediol increased 163-fold when the enzyme was incubated with fluoxetine and 63-fold with paroxetine (Griffin & Mellon, 1999), which greatly reduces intracellular DHT.

Using the H295R cell line, Hansen et al. demonstrated that commonly used SSRIs fluoxetine, paroxetine, citalopram, escitalopram, sertraline and fluvoxamine exert significant endocrine disrupting properties in vitro. Despite different steroidogenic enzymes being affected across the six different drugs, the outcome was the same in terms of a marked decrease in testosterone. Observing that the steroidogenic interruptions may partly explain some of the sexual disorders associated with SSRIs, Hansen et al. suggest that the endocrine disrupting potential of these drugs at pharmacologically relevant doses should encourage their careful use in therapy (Hansen et al., 2017). A similar decrease in testosterone in this cell line following exposure to five SSRI drugs had previously been reported (Jacobsen et al., 2015). Munkboel et al. demonstrated that steroidogenesis was significantly disrupted in rats exposed to therapeutic doses of sertraline. The most significant effects observed on testicular sex steroid production, particularly the Delta 4 steroidogenic pathway (comprising progesterone, 17-hydroxyprogesterone, Androstenedione, Testosterone, DHT). Testosterone production was significantly decreased in all 3 exposure groups, and DHT was significantly decreased in the testis, plasma and brain. A 53% decrease of testosterone was reported in testis of rats exposed to 5 mg/kg/day alongside a general decrease on the D4 axis. Munkboel et al. note that this corresponds to the human starting dose of 50mg per day and this pronounced effect suggests the possibility of significant consequences on reproductive and health endpoints. They conclude that men treated with sertraline should be monitored carefully for sexual dysfunction (Munkboel et al., 2018).

Serotonin is recognised to be inhibitory of both male and female sexual behaviour and function (Croft, 2017; Iovino et al., 2019; Olivier et al., 2010). SSRIs increase inhibition of serotonin reuptake (Ferguson, 2001), and increase serotonin by a downregulation of autoreceptors which otherwise act to inhibit serotonin release (Hagan et al., 2012; Neumaier, 1996). Both 5HT1a receptor knockdown and interference using siRNA molecules has demonstrated antidepressant effects accompanied with greater increases in extracellular serotonin in response to either stress or fluoxetine (Ferrés-Coy et al., 2012).

Increased extracellular serotonin levels in the ventral hippocampus of 5HT1b knockout mice were observed in response to SSRI administration (Nautiyal et al., 2016). As well as reuptake inhibition, SSRIs have been observed to upregulate tryptophan hydroxylase (Kim et al., 2002), mediatory of serotonin production in non-neuronal and neuronal tissue (Walther, 2003; X. Zhang, 2004).

There is a well-studied and remarkable antagonism between testosterone and serotonin in terms of their behavioural effects that aligns with the significant impact of androgens on serotonergic activity in the brain (Ambar & Chiavegatto, 2009; Daly et al., 2001; Keleta et al., 2007; Martinez-Conde et al., 1985; Sundblad & Eriksson, 1997; L. Zhang et al., 1999). Testosterone promotes territorial behaviour, impulsivity, sexual behaviour and aggression (Bing et al., 1998; Kimura & Hagiwara, 1985; Svensson, 2003; Wu & Shah, 2011), whereas serotonin appears to exert opposite effects (Batty & Meyerson, 1980; Nelson & Chiavegatto, 2001; Olivier et al., 2010). Studer et al. demonstrated that while the pro-aggressive effect of testosterone is apparently independent of serotonin, the inhibitory effect of serotonin to dampen maladaptive aggression is "irrelevant" in the absence of testosterone. Additionally, inhibition of serotonin production failed to reinstate aggression in mice rendered hypoaggressive by early life brain AR knockout (Studer et al., 2015).

Recent evidence in tissue outside the brain shows that serotonin exerts a powerful downregulatory effect on the androgen receptor. BPH tissue has been observed to demonstrate AR upregulation (Izumi et al., 2013; Nicholson et al., 2013; P. Zhang et al., 2015) as well as a significant depletion of 5-HT (Cockett et al., 1993). Carvalho-Dias et al explored the relationship between 5-HT and androgen signaling, demonstrating a clear inhibitory influence of serotonin on the androgen pathway, providing robust data from a number of elegant in vitro and in vivo observations. In vitro, 5-HT significantly inhibited rat prostate cell growth through a 5-HT1a and 5-HT1b mediated down-regulation of AR either with or without testosterone supplementation. In cultured human cell lines, proliferation of BPH epithelium and normal prostate stroma cells supplemented with testosterone was significantly reduced by 5-HT or specific 5-HT1a and 5HT1b agonists. Proliferation of normal prostate epithelium cells was not affected. Testosterone was observed to upregulate the AR in BPH epithelium and markedly in normal stroma, while 5-HT or specific 5-HT1a and 5HT1b agonists inhibited this upregulation. Importantly, the absence of an inhibitory action of 5HT or an agonist of either autoreceptor on viability and proliferation of normal epithelium cells, with or without testosterone supplementation, was coincidental with a complete absence of AR expression in these cells. They additionally demonstrated that tryptophan hydroxylase type 1 knockout mice exhibit a remarkable 37% higher prostate-to-body weight ratio compared to wild-type at 20 weeks without difference in overall body weight, with prostate histology revealing areas of hyperplasia in epithelium and stroma. These mice displayed significantly larger seminal vesicles than controls, supportive of negative androgenic regulation by 5HT beyond the prostate cell lines. qRT-PCR revealed increased AR levels in the dorsolateral prostate of Tph1^{-/-} mice. Remarkably, 5HT treatment significantly reduced prostate weight and seminal vesicles near to that of controls, and reduced AR mRNA to levels comparable to controls (Carvalho-Dias et al., 2017).

Collectively, these in vitro and in vivo studies demonstrate that the inhibition of 5alpha reductase type 2 with finasteride and steroidogenic dysregulation with SSRIs have a clear mechanistic commonality: A considerable disruption to androgen signaling. As with isotretinoin, SSRIs exert antiandrogenic endocrine disruptive activity through distinct actions. Further supporting this hypothesis, a significantly affected patient registered on propeciahelp.com suffers the syndrome following over the counter use of 5-hydroxytryptophan, a serotonin precursor observed to increase excretion of 5-HIAA with significant interindividual variation (Joy et al., 2008). This is suggestive of increased production of serotonin following 5-HTP intake, which is the rationale underlying its supplemental use (Hallin et al., 2012).

Saw Palmetto (*Serenoa repens*)

Amongst propeciahelp membership, *Serenoa repens* (saw palmetto), an extract with markedly antiandrogenic properties commonly used in treatment of BPH and LUTS (Cicero et al., 2019), is the most prevalent alternative therapy causative of the syndrome. This is usually taken as a "natural" hair loss remedy. Although proportionally rarer, topical antiandrogenic products are causing patients to present with the syndrome, and these include finasteride, ketoconazole, the antiandrogen RU-58841 and minoxidil. In animals, Finasteride has been demonstrated to have significant systemic effects following topical application (Chen et al., 1995). Ketoconazole is antiandrogenic and suppressive of steroid production, exhibiting nonmonotonic activity. As with other imidazole azole class drugs, the extremely potent endocrine disruptive properties of ketoconazole are attracting increasing scrutiny given their prevalence as antifungal treatments (Munkboel et al., 2019). In vitro investigations have demonstrated minoxidil can directly bind to the AR, decrease transcriptional activity and interfere with AR function (Hsu et al., 2014). Additionally, minoxidil has been shown to significantly downregulate 5 alpha reductase type 2 expression in human keratinocytes (Pekmezci & Türkoğlu, 2017). A 28 year old patient member of our site recently received a diagnosis of "5 alpha reductase inhibitor syndrome" after one week of oral quercetin-3-O-rutinoside under physician direction led to the rapid development of persistent symptoms including severe muscle loss, increased adiposity, osteoporosis of the hip and lumbar spine, severe penile atrophy, post orgasmic illness, impotence, anxiety, depression and insomnia. Polyphenols can be potent 5alpha reductase inhibitors (Hiipakka et al., 2002) and antiandrogenic at the receptor level (Boam, 2015; Cicero et al., 2019; Kampa et al., 2017; Xing, 2001). Nordeen et al. noted the lack of data regarding purified concentrated flavonoid supplements, while providing evidence that two flavonoids, luteolin and quercetin, are "promiscuous endocrine disruptors" that demonstrate anti-androgenic effects, suggesting caution regarding the potential "peril" of supplementing these phenols far beyond the intake of a normal, healthy diet (Nordeen et al., 2013).

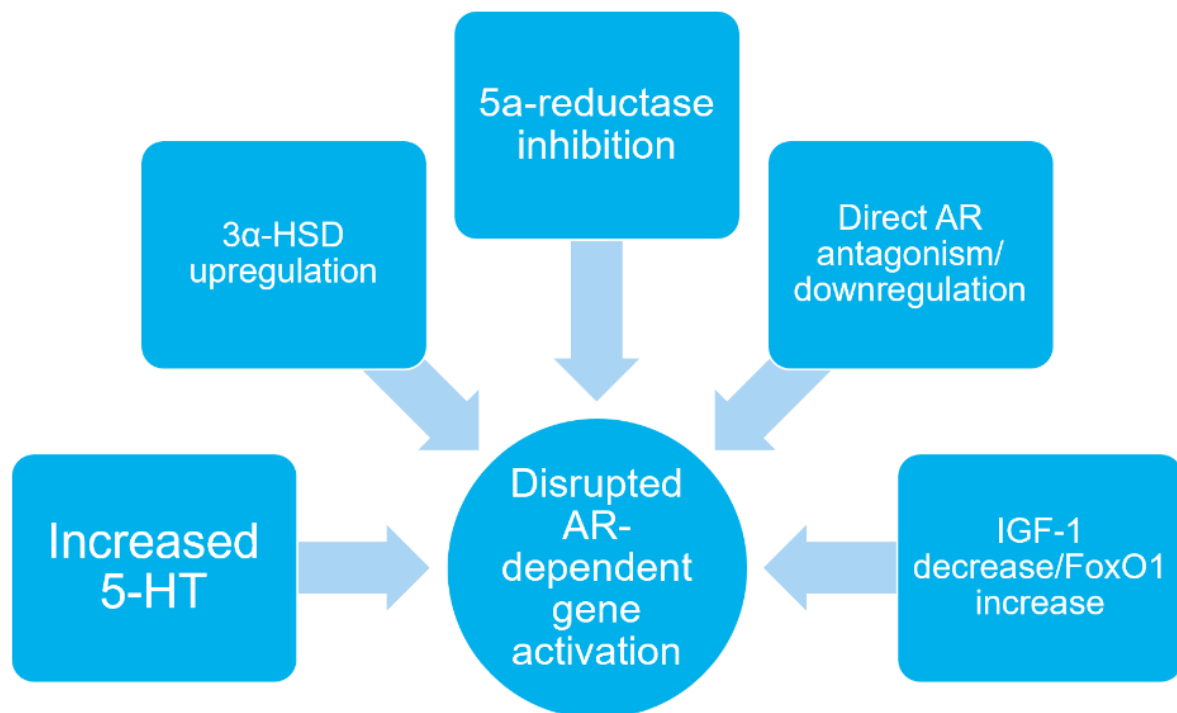


Fig. Finasteride, Accutane and SSRIs are all potent antiandrogenic endocrine disruptors.

While this large range of pharmaceutical and natural substances may seem broad and mechanistically distinct, the notable commonality is dramatic antiandrogenic endocrine disruption. Any treatments targeting the AR or suppressing androgens are known to have adverse effects on other critical physiological functions (Bourke et al., 2011). Narrow mechanistic perspectives often inform substance grouping for analysis of the risk of permanent male reproductive malformations and irreversible sexual disorders in the developing foetus. Through analysis of adverse outcome pathway networks, Kortenkamp illustrated that independent mechanistic effects from a very broad range of substances meet at nodal points in the network to result in common down-stream antiandrogenic effects and adverse outcomes. Kortenkamp suggested that - in addition to phthalates - substances capable of AR antagonism, cholesterol transporter down-regulation, and interruption or inhibition of steroidogenic or cholesterol synthesising enzymes should be included in an expanded consideration of substances capable of inducing male reproductive malformation. A non-exhaustive list of chemicals identified as a starting basis included vinclozolin, bisphenol A, finasteride, paracetamol, ibuprofen, ketoconazole and simvastatin (Kortenkamp, 2020).

Page Bibliography

1. Drugs@FDA, . (2016). *FDA Approved Drug Products. US Food and Drug Administration for All Antidepressants*. FDA.Gov.
<http://www.accessdata.fda.gov/scripts/cder/daf/index.cfm?event=overview.process&ApplNo=021>

[860](#)

2. Agamia, N. F., Roshdy, O. H., Abdelmaksoud, R. E., Abdalla, D. M., Talaat, I. M., Zaki, E. I., El Tawdy, A., & Melnik, B. C. (2018). Effect of oral isotretinoin on the nucleo?cytoplasmic distribution of FoxO1 and FoxO3 proteins in sebaceous glands of patients with acne vulgaris. *Experimental Dermatology*, 1344–1351. <https://doi.org/10.1111/exd.13787>
3. Alimirah, F., Panchanathan, R., Chen, J., Zhang, X., Ho, S.-M., & Choubey, D. (2007). Expression of Androgen Receptor Is Negatively Regulated By p53. *Neoplasia*, 1152–1159. <https://doi.org/10.1593/neo.07769>
4. Ambar, G., & Chiavegatto, S. (2009). Anabolic-androgenic steroid treatment induces behavioral disinhibition and downregulation of serotonin receptor messenger RNA in the prefrontal cortex and amygdala of male mice. *Genes, Brain and Behavior*, 161–173. <https://doi.org/10.1111/j.1601-183x.2008.00458.x>
5. Atli, O., Baysal, M., Aydogan-Kilic, G., Kilic, V., Ucarcan, S., Karaduman, B., & Ilgin, S. (2017). Sertraline-induced reproductive toxicity in male rats: evaluation of possible underlying mechanisms. *Asian Journal of Andrology*, 672. <https://doi.org/10.4103/1008-682x.192637>
6. Batty, J., & Meyerson, B. J. (1980). The effects of p-Chlorophenylalanine, fenfluramine and ?-methyltyrosine on marking responses in the male Mongolian gerbil (*Meriones unguiculatus*). *Pharmacology Biochemistry and Behavior*, 181–184. [https://doi.org/10.1016/0091-3057\(80\)90352-4](https://doi.org/10.1016/0091-3057(80)90352-4)
7. Bergman, Å., Heindel, J., Jobling, S., Kidd, K., & Zoeller, R. T. (2012). State-of-the-science of endocrine disrupting chemicals, 2012. *Toxicology Letters*, S3. <https://doi.org/10.1016/j.toxlet.2012.03.020>
8. Bing, O., Heilig, M., Kakoulidis, P., Sundblad, C., Wiklund, L., & Eriksson, E. (1998). High doses of testosterone increase anticonflict behaviour in rat. *European Neuropsychopharmacology*, 321–323. [https://doi.org/10.1016/s0924-977x\(97\)00095-3](https://doi.org/10.1016/s0924-977x(97)00095-3)
9. Boam, T. (2015). Anti-androgenic effects of flavonols in prostate cancer. *Ecancermedicalscience*. <https://doi.org/10.3332/ecancer.2015.585>
10. Boudou, P., Chivot, M., Vexiau, P., Soliman, H., Villette, J. M., Julien, R., Belanger, A., & Fiet, J.

- (1994). Evidence for decreased androgen 5 alpha-reduction in skin and liver of men with severe acne after 13-cis-retinoic acid treatment. *The Journal of Clinical Endocrinology & Metabolism*, 1064–1069. <https://doi.org/10.1210/jcem.78.5.8175961>
11. Boudou, P., Soliman, H., Chivot, M., Villette, J. M., Vexiau, P., Belanger, A., & Fiet, J. (1995). Effect of oral isotretinoin treatment on skin androgen receptor levels in male acneic patients. *The Journal of Clinical Endocrinology & Metabolism*, 1158–1161. <https://doi.org/10.1210/jcem.80.4.7714084>
12. Bourke, L., Chico, T. J. A., Albertsen, P. C., Hamdy, F. C., & Rosario, D. J. (2011). Cardiovascular risk in androgen suppression: underappreciated, under-researched and unresolved: Figure 1. *Heart*, 345–348. <https://doi.org/10.1136/heartjnl-2011-300893>
13. Bowman, C. J., Barlow, N. J., Turner, K. J., Wallace, D. G., & Foster, P. M. D. (2003). Effects of in Utero Exposure to Finasteride on Androgen-Dependent Reproductive Development in the Male Rat. *Toxicological Sciences*, 393–406. <https://doi.org/10.1093/toxsci/kfg128>
14. Cappel, M. (2005). Correlation Between Serum Levels of Insulin-like Growth Factor 1, Dehydroepiandrosterone Sulfate, and Dihydrotestosterone and Acne Lesion Counts in Adult Women. *Archives of Dermatology*, 333–338. <https://doi.org/10.1001/archderm.141.3.333>
15. Carvalho-Dias, E., Miranda, A., Martinho, O., Mota, P., Costa, Â., Nogueira-Silva, C., Moura, R. S., Alenina, N., Bader, M., Autorino, R., Lima, E., & Correia-Pinto, J. (2017). Serotonin regulates prostate growth through androgen receptor modulation. *Scientific Reports*. <https://doi.org/10.1038/s41598-017-15832-5>
16. Chen, C., Puy, L. A., Simard, J., Li, X., Singh, S. M., & Labrie, F. (1995). Local and Systemic Reduction by Topical Finasteride or Flutamide of Hamster Flank Organ Size and Enzyme Activity. *Journal of Investigative Dermatology*, 678–682. <https://doi.org/10.1111/1523-1747.ep12324390>
17. Cicero, A. F. G., Allkanjari, O., Busetto, G. M., Cai, T., Larganà, G., Magri, V., Perletti, G., Robustelli Della Cuna, F. S., Russo, G. I., Stamatiou, K., Trinchieri, A., & Vitalone, A. (2019). Nutraceutical treatment and prevention of benign prostatic hyperplasia and prostate cancer. *Archivio Italiano Di Urologia e Andrologia*. <https://doi.org/10.4081/aiua.2019.3.139>
18. Cockett, A. T. K., di Sant'Agnes, P. A., Gopinath, P., Schoen, S. R., & Abrahamsson, P.-A. (1993). Relationship of neuroendocrine cells of prostate and serotonin to benign prostatic

hyperplasia. *Urology*, 512–519. [https://doi.org/10.1016/0090-4295\(93\)90260-h](https://doi.org/10.1016/0090-4295(93)90260-h)

19. Croft, H. A. (2017). Understanding the Role of Serotonin in Female Hypoactive Sexual Desire Disorder and Treatment Options. *The Journal of Sexual Medicine*, 1575–1584. <https://doi.org/10.1016/j.jsxm.2017.10.068>
20. Csoka, A. B., & Szyf, M. (2009). Epigenetic side-effects of common pharmaceuticals: A potential new field in medicine and pharmacology. *Medical Hypotheses*, 770–780. <https://doi.org/10.1016/j.mehy.2008.10.039>
21. Cunha, V., Rodrigues, P., Santos, M. M., Moradas-Ferreira, P., & Ferreira, M. (2018). Fluoxetine modulates the transcription of genes involved in serotonin, dopamine and adrenergic signalling in zebrafish embryos. *Chemosphere*, 954–961. <https://doi.org/10.1016/j.chemosphere.2017.10.100>
22. Daly, R. C., Su, T.-P., Schmidt, P. J., Pickar, D., Murphy, D. L., & Rubinow, D. R. (2001). Cerebrospinal Fluid and Behavioral Changes After Methyltestosterone Administration. *Archives of General Psychiatry*, 172. <https://doi.org/10.1001/archpsyc.58.2.172>
23. David Healy, Joanna Le Noury, & Derelie Mangin. (2018). Enduring sexual dysfunction after treatment with antidepressants, 5 α -reductase inhibitors and isotretinoin: 300 cases. *International Journal of Risk & Safety in Medicine*, 125–134. <https://doi.org/10.3233/JRS-180744>
24. Fan, W., Yanase, T., Morinaga, H., Okabe, T., Nomura, M., Daitoku, H., Fukamizu, A., Kato, S., Takayanagi, R., & Nawata, H. (2007). Insulin-like Growth Factor 1/Insulin Signaling Activates Androgen Signaling through Direct Interactions of Foxo1 with Androgen Receptor. *Journal of Biological Chemistry*, 7329–7338. <https://doi.org/10.1074/jbc.m610447200>
25. Ferguson, J. M. (2001). SSRI Antidepressant Medications. *The Primary Care Companion to The Journal of Clinical Psychiatry*, 22–27. <https://doi.org/10.4088/pcc.v03n0105>
26. Ferrés-Coy, A., Santana, N., Castañé, A., Cortés, R., Carmona, M. C., Toth, M., Montefeltro, A., Artigas, F., & Bortolozzi, A. (2012). Acute 5-HT_{1A} autoreceptor knockdown increases antidepressant responses and serotonin release in stressful conditions. *Psychopharmacology*, 61–74. <https://doi.org/10.1007/s00213-012-2795-9>
27. Fucic, A., Galea, K., Duca, R., El Yamani, M., Frery, N., Godderis, L., Halldorsson, T., Iavicoli, I., Ndaw, S., Ribeiro, E., Viegas, S., & Moshammer, H. (2018). Potential Health Risk of

Endocrine Disruptors in Construction Sector and Plastics Industry: A New Paradigm in Occupational Health. *International Journal of Environmental Research and Public Health*, 1229. <https://doi.org/10.3390/ijerph15061229>

28. Giatti, S., Diviccaro, S., Panzica, G., & Melcangi, R. C. (2018). Post-finasteride syndrome and post-SSRI sexual dysfunction: two sides of the same coin? *Endocrine*, 180–193. <https://doi.org/10.1007/s12020-018-1593-5>
29. Griffin, L. D., & Mellon, S. H. (1999). Selective serotonin reuptake inhibitors directly alter activity of neurosteroidogenic enzymes. *Proceedings of the National Academy of Sciences*, 13512–13517. <https://doi.org/10.1073/pnas.96.23.13512>
30. Hagan, C. E., Mcdevitt, R. A., Liu, Y., Furay, A. R., & Neumaier, J. F. (2012). 5-HT1Bautoreceptor regulation of serotonin transporter activity in synaptosomes. *Synapse*, 1024–1034. <https://doi.org/10.1002/syn.21608>
31. Hallin, M. L. P., Mahmoud, K., Viswanath, A., & Gama, R. (2012). “Sweet Dreams”, “Happy Days” and elevated 24-h urine 5-hydroxyindoleacetic acid excretion. *Annals of Clinical Biochemistry*, 80–82. <https://doi.org/10.1258/acb.2012.012041>
32. Hansen, C. H., Larsen, L. W., Sørensen, A. M., Halling-Sørensen, B., & Styrihave, B. (2017). The six most widely used selective serotonin reuptake inhibitors decrease androgens and increase estrogens in the H295R cell line. *Toxicology in Vitro*, 1–11. <https://doi.org/10.1016/j.tiv.2017.02.001>
33. Hiipakka, R. A., Zhang, H.-Z., Dai, W., Dai, Q., & Liao, S. (2002). Structure–activity relationships for inhibition of human 5 α -reductases by polyphenols. *Biochemical Pharmacology*, 1165–1176. [https://doi.org/10.1016/s0006-2952\(02\)00848-1](https://doi.org/10.1016/s0006-2952(02)00848-1)
34. Hsu, C.-L., Liu, J.-S., Lin, A.-C., Yang, C.-H., Chung, W.-H., & Wu, W.-G. (2014). Minoxidil may suppress androgen receptor-related functions. *Oncotarget*. <https://doi.org/10.18632/oncotarget.1886>
35. Ilgin, S., Kilic, G., Baysal, M., Kilic, V., Korkut, B., Ucarcan, S., & Atli, O. (2017). Citalopram Induces Reproductive Toxicity in Male Rats. *Birth Defects Research*, 475–485. <https://doi.org/10.1002/bdr2.1010>

36. Iovino, M., Messina, T., Iovino, E., De Pergola, G., Guastamacchia, E., Giagulli, V. A., & Triggiani, V. (2019). Neuroendocrine Mechanisms Involved in Male Sexual and Emotional Behavior. *Endocrine, Metabolic & Immune Disorders - Drug Targets*, 472–480. <https://doi.org/10.2174/1871530319666190131155310>
37. Izumi, K., Mizokami, A., Lin, W.-J., Lai, K.-P., & Chang, C. (2013). Androgen Receptor Roles in the Development of Benign Prostate Hyperplasia. *The American Journal of Pathology*, 1942–1949. <https://doi.org/10.1016/j.ajpath.2013.02.028>
38. Jacobsen, N. W., Hansen, C. H., Nellemann, C., Styrisshave, B., & Halling-Sørensen, B. (2015). Effects of selective serotonin reuptake inhibitors on three sex steroids in two versions of the aromatase enzyme inhibition assay and in the H295R cell assay. *Toxicology in Vitro*, 1729–1735. <https://doi.org/10.1016/j.tiv.2015.07.005>
39. Joy, T., Walsh, G., Tokmakejian, S., & Van Uum, S. H. (2008). Increase of Urinary 5-Hydroxyindoleacetic Acid Excretion but Not Serum Chromogranin a Following Over-The-Counter 5-Hydroxytryptophan Intake. *Canadian Journal of Gastroenterology*, 49–53. <https://doi.org/10.1155/2008/472159>
40. Kampa, M., Notas, G., & Castanas, E. (2017). Natural extranuclear androgen receptor ligands as endocrine disruptors of cancer cell growth. *Molecular and Cellular Endocrinology*, 43–48. <https://doi.org/10.1016/j.mce.2017.02.021>
41. Karadag, A.S., Ertugrul, D. T., Tural, E., & Akin, K. O. (2009). Short-term isotretinoin treatment decreases insulin-like growth factor-1 and insulin-like growth factor binding protein-3 levels: does isotretinoin affect growth hormone physiology? *British Journal of Dermatology*, 798–802. <https://doi.org/10.1111/j.1365-2133.2009.09618.x>
42. Karadag, Ayse Serap, Takci, Z., Ertugrul, D. T., Bilgili, S. G., Balahoroglu, R., & Takir, M. (2015). The Effect of Different Doses of Isotretinoin on Pituitary Hormones. *Dermatology*, 354–359. <https://doi.org/10.1159/000375370>
43. Keleta, Y. B., Lumia, A. R., Anderson, G. M., & McGinnis, M. Y. (2007). Behavioral effects of pubertal anabolic androgenic steroid exposure in male rats with low serotonin. *Brain Research*, 129–138. <https://doi.org/10.1016/j.brainres.2006.10.097>
44. Kim, S. W., Park, S. Y., & Hwang, O. (2002). Up-Regulation of Tryptophan Hydroxylase Expression and Serotonin Synthesis by Sertraline. *Molecular Pharmacology*, 778–785.

<https://doi.org/10.1124/mol.61.4.778>

45. Kimura, T., & Hagiwara, Y. (1985). Regulation of urine marking in male and female mice: Effects of sex steroids. *Hormones and Behavior*, 64–70.
[https://doi.org/10.1016/0018-506x\(85\)90006-6](https://doi.org/10.1016/0018-506x(85)90006-6)
46. Lagarde, F., Beausoleil, C., Belcher, S. M., Belzunces, L. P., Emond, C., Guerbet, M., & Rousselle, C. (2015). Non-monotonic dose-response relationships and endocrine disruptors: a qualitative method of assessment. *Environmental Health*.
<https://doi.org/10.1186/1476-069x-14-13>
47. Lai, J.-J., Chang, P., Lai, K.-P., Chen, L., & Chang, C. (2012). The role of androgen and androgen receptor in skin-related disorders. *Archives of Dermatological Research*, 499–510.
<https://doi.org/10.1007/s00403-012-1265-x>
48. Latchney, S. E., Fields, A. M., & Susiarjo, M. (2017). Linking inter-individual variability to endocrine disruptors: insights for epigenetic inheritance. *Mammalian Genome*, 141–152.
<https://doi.org/10.1007/s00335-017-9729-0>
49. Martinez-Conde, E., Leret, M. L., & Diaz, S. (1985). The influence of testosterone in the brain of the male rat on levels of serotonin (5-HT) and hydroxyindole-acetic acid (5-HIAA). *Comparative Biochemistry and Physiology Part C: Comparative Pharmacology*, 411–414.
[https://doi.org/10.1016/0742-8413\(85\)90077-5](https://doi.org/10.1016/0742-8413(85)90077-5)
50. McIntyre, B. S. (2001). Androgen-Mediated Development in Male Rat Offspring Exposed to Flutamide in Utero: Permanence and Correlation of Early Postnatal Changes in Anogenital Distance and Nipple Retention with Malformations in Androgen-Dependent Tissues. *Toxicological Sciences*, 236–249. <https://doi.org/10.1093/toxsci/62.2.236>
51. McIntyre, B. S. (2002). Male Rats Exposed to Linuron in Utero Exhibit Permanent Changes in Anogenital Distance, Nipple Retention, and Epididymal Malformations That Result in Subsequent Testicular Atrophy. *Toxicological Sciences*, 62–70. <https://doi.org/10.1093/toxsci/65.1.62>
52. Melnik, B. C. (2017). p53: key conductor of all anti-acne therapies. *Journal of Translational Medicine*. <https://doi.org/10.1186/s12967-017-1297-2>
53. Munkboel, C. H., Larsen, L. W., Weisser, J. J., Møbjerg Kristensen, D., & Styrihave, B. (2018).

Sertraline Suppresses Testis and Adrenal Steroid Production and Steroidogenic Gene Expression While Increasing LH in Plasma of Male Rats Resulting in Compensatory Hypogonadism. *Toxicological Sciences*, 609–619. <https://doi.org/10.1093/toxsci/kfy059>

54. Munkboel, C. H., Rasmussen, T. B., Elgaard, C., Olesen, M.-L. K., Kretschmann, A. C., & Styrisshave, B. (2019). The classic azole antifungal drugs are highly potent endocrine disruptors in vitro inhibiting steroidogenic CYP enzymes at concentrations lower than therapeutic Cmax. *Toxicology*, 152247. <https://doi.org/10.1016/j.tox.2019.152247>
55. Nautiyal, K. M., Tritschler, L., Ahmari, S. E., David, D. J., Gardier, A. M., & Hen, R. (2016). A Lack of Serotonin 1B Autoreceptors Results in Decreased Anxiety and Depression-Related Behaviors. *Neuropsychopharmacology*, 2941–2950. <https://doi.org/10.1038/npp.2016.109>
56. Nelson, R. J., & Chiavegatto, S. (2001). Molecular basis of aggression. *Trends in Neurosciences*, 713–719. [https://doi.org/10.1016/s0166-2236\(00\)01996-2](https://doi.org/10.1016/s0166-2236(00)01996-2)
57. Neumaier, J. (1996). Chronic Fluoxetine Reduces Serotonin Transporter mRNA and 5-HT1B mRNA in a Sequential Manner in the Rat Dorsal Raphe Nucleus. *Neuropsychopharmacology*, 515–522. [https://doi.org/10.1016/s0893-133x\(96\)00095-4](https://doi.org/10.1016/s0893-133x(96)00095-4)
58. Nicholson, T. M., Sehgal, P. D., Drew, S. A., Huang, W., & Ricke, W. A. (2013). Sex steroid receptor expression and localization in benign prostatic hyperplasia varies with tissue compartment. *Differentiation*, 140–149. <https://doi.org/10.1016/j.diff.2013.02.006>
59. Nordeen, S. K., Bona, B. J., Jones, D. N., Lambert, J. R., & Jackson, T. A. (2013). Endocrine Disrupting Activities of the Flavonoid Nutraceuticals Luteolin and Quercetin. *Hormones and Cancer*, 293–300. <https://doi.org/10.1007/s12672-013-0150-1>
60. Olivier, B., Chan, J. S. W., Snoeren, E. M., Olivier, J. D. A., Veening, J. G., Vinkers, C. H., Waldinger, M. D., & Oosting, R. S. (2010). Differences in Sexual Behaviour in Male and Female Rodents: Role of Serotonin. In *Biological Basis of Sex Differences in Psychopharmacology* (pp. 15–36). Springer Berlin Heidelberg. https://doi.org/10.1007/7854_2010_116
61. Ostby, J., Kelce, W. R., Lambright, C., Wolf, C. J., Mann, P., & Gray, L. E., Jr. (1999). The fungicide procymidone alters sexual differentiation in the male rat by acting as an androgen-receptor antagonist in vivo and in vitro. *Toxicology and Industrial Health*, 80–93. <https://doi.org/10.1177/074823379901500108>

62. Palazzolo, I., Stack, C., Kong, L., Musaro, A., Adachi, H., Katsuno, M., Sobue, G., Taylor, J. P., Sumner, C. J., Fischbeck, K. H., & Pennuto, M. (2009). Overexpression of IGF-1 in Muscle Attenuates Disease in a Mouse Model of Spinal and Bulbar Muscular Atrophy. *Neuron*, 316–328. <https://doi.org/10.1016/j.neuron.2009.07.019>
63. Pappas, K., Xu, J., Zairis, S., Resnick-Silverman, L., Abate, F., Steinbach, N., Ozturk, S., Saal, L. H., Su, T., Cheung, P., Schmidt, H., Aaronson, S., Hibshoosh, H., Manfredi, J., Rabadan, R., & Parsons, R. (2017). p53 Maintains Baseline Expression of Multiple Tumor Suppressor Genes. *Molecular Cancer Research*, 1051–1062. <https://doi.org/10.1158/1541-7786.mcr-17-0089>
64. Patisaul, H. B., & Belcher, S. M. (2017). Endocrine Disruptors, Brain, and Behavior. In *Oxford Scholarship Online*. Oxford University Press. <https://doi.org/10.1093/acprof:oso/9780199935734.001.0001>
65. Pekmezci, E., & Türko?lu, M. (2017). Minoxidil Acts as an Antiandrogen: A Study of 5?-reductase Type 2 Gene Expression in a Human Keratinocyte Cell Line. *Acta Dermatovenerologica Croatica : ADC*, 25(4), 271–275. <https://www.ncbi.nlm.nih.gov/pubmed/30064598>
66. Sammartino, M. P., Castrucci, M., Ruiu, D., Visco, G., & Campanella, L. (2013). Photostability and toxicity of finasteride, diclofenac and naproxen under simulating sunlight exposure: evaluation of the toxicity trend and of the packaging photoprotection. *Chemistry Central Journal*. <https://doi.org/10.1186/1752-153x-7-181>
67. Shenk, J. L., Fisher, C. J., Chen, S.-Y., Zhou, X.-F., Tillman, K., & Shemshedini, L. (2001). p53 Represses Androgen-induced Transactivation of Prostate-specific Antigen by Disrupting hAR Amino- to Carboxyl-terminal Interaction. *Journal of Biological Chemistry*, 38472–38479. <https://doi.org/10.1074/jbc.m103652200>
68. Shi, G., Liao, P.-Y., Cai, X.-L., Pi, X.-X., Zhang, M.-F., Li, S.-J., Quan, J.-H., & Fan, Y.-M. (2018). FoxO1 enhances differentiation and apoptosis in human primary keratinocytes. *Experimental Dermatology*, 1254–1260. <https://doi.org/10.1111/exd.13775>
69. Solecki, R., Kortenkamp, A., Bergman, Å., Chahoud, I., Degen, G. H., Dietrich, D., Greim, H., Håkansson, H., Hass, U., Husoy, T., Jacobs, M., Jobling, S., Mantovani, A., Marx-Stoelting, P., Piersma, A., Ritz, V., Slama, R., Stahlmann, R., van den Berg, M., ... Boobis, A. R. (2016). Scientific principles for the identification of endocrine-disrupting chemicals: a consensus statement. *Archives of Toxicology*, 1001–1006. <https://doi.org/10.1007/s00204-016-1866-9>

70. Studer, E., Näslund, J., Andersson, E., Nilsson, S., Westberg, L., & Eriksson, E. (2015). Serotonin Depletion-Induced Maladaptive Aggression Requires the Presence of Androgens. *PLOS ONE*, e0126462. <https://doi.org/10.1371/journal.pone.0126462>
71. Sundblad, C., & Eriksson, E. (1997). Reduced extracellular levels of serotonin in the amygdala of androgenized female rats. *European Neuropsychopharmacology*, 253–259. [https://doi.org/10.1016/s0924-977x\(97\)00031-x](https://doi.org/10.1016/s0924-977x(97)00031-x)
72. Svensson, A. (2003). Testosterone treatment induces behavioral disinhibition in adult male rats. *Pharmacology Biochemistry and Behavior*, 481–490. [https://doi.org/10.1016/s0091-3057\(03\)00137-0](https://doi.org/10.1016/s0091-3057(03)00137-0)
73. Tanrikut, C., Feldman, A. S., Altemus, M., Paduch, D. A., & Schlegel, P. N. (2010). Adverse effect of paroxetine on sperm. *Fertility and Sterility*, 1021–1026. <https://doi.org/10.1016/j.fertnstert.2009.04.039>
74. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>
75. Trasande, L., Zoeller, R. T., Hass, U., Kortenkamp, A., Grandjean, P., Myers, J. P., DiGangi, J., Bellanger, M., Hauser, R., Legler, J., Skakkebaek, N. E., & Heindel, J. J. (2015). Estimating Burden and Disease Costs of Exposure to Endocrine-Disrupting Chemicals in the European Union. *The Journal of Clinical Endocrinology & Metabolism*, 1245–1255. <https://doi.org/10.1210/jc.2014-4324>
76. Ubels, J. L., Veenstra, E., Ditlev, J., & Ingersoll, K. (2003). Interactions of testosterone and all-trans retinoic acid in regulation of androgen receptor expression in rat lacrimal gland. *Experimental Eye Research*, 741–748. <https://doi.org/10.1016/j.exer.2003.07.006>
77. Ubels, J. L., Wertz, J. T., Ingersoll, K. E., Jackson II, R. S., & Aupperlee, M. D. (2002). Down-regulation of Androgen Receptor Expression and Inhibition of Lacrimal Gland Cell Proliferation by Retinoic Acid. *Experimental Eye Research*, 561–571. <https://doi.org/10.1006/exer.2002.2054>
78. Vandenberg, L. N., Colborn, T., Hayes, T. B., Heindel, J. J., Jacobs, D. R., Jr., Lee, D.-H., Shioda, T., Soto, A. M., vom Saal, F. S., Welshons, W. V., Zoeller, R. T., & Myers, J. P. (2012).

Hormones and Endocrine-Disrupting Chemicals: Low-Dose Effects and Nonmonotonic Dose Responses. *Endocrine Reviews*, 378–455. <https://doi.org/10.1210/er.2011-1050>

79. Walther, D. J. (2003). Synthesis of Serotonin by a Second Tryptophan Hydroxylase Isoform. *Science*, 76–76. <https://doi.org/10.1126/science.1078197>
80. Wolf, C., Lambright, C., Mann, P., Price, M., Cooper, R. L., Ostby, J., & Gray, L. E., Jr. (1999). Administration of potentially antiandrogenic pesticides (procymidone, linuron, iprodione, chlozolinate, p,p'-DDE, and ketoconazole) and toxic substances (dibutyl- and diethylhexyl phthalate, PCB 169, and ethane dimethane sulphonate) during sexual differentiation produces diverse profiles of reproductive malformations in the male rat. *Toxicology and Industrial Health*, 94–118. <https://doi.org/10.1177/074823379901500109>
81. Wu, M. V., & Shah, N. M. (2011). Control of masculinization of the brain and behavior. *Current Opinion in Neurobiology*, 116–123. <https://doi.org/10.1016/j.conb.2010.09.014>
82. Xing, N. (2001). Quercetin inhibits the expression and function of the androgen receptor in LNCaP prostate cancer cells. *Carcinogenesis*, 409–414. <https://doi.org/10.1093/carcin/22.3.409>
83. Yanase, T., & Fan, W. (2009). Chapter 23 Modification of Androgen Receptor Function by Igf?1 Signaling. In *Vitamins & Hormones* (pp. 649–666). Elsevier. [https://doi.org/10.1016/s0083-6729\(08\)00623-7](https://doi.org/10.1016/s0083-6729(08)00623-7)
84. Zhang, L., Ma, W., Barker, J. L., & Rubinow, D. R. (1999). Sex differences in expression of serotonin receptors (subtypes 1A and 2A) in rat brain: a possible role of testosterone. *Neuroscience*, 251–259. [https://doi.org/10.1016/s0306-4522\(99\)00234-1](https://doi.org/10.1016/s0306-4522(99)00234-1)
85. Zhang, P., Hu, W.-L., Cheng, B., Zeng, Y.-J., Wang, X.-H., Liu, T.-Z., & Zhang, W.-B. (2015). Which play a more important role in the development of large-sized prostates (?80 ml), androgen receptors or oestrogen receptors? A comparative study. *International Urology and Nephrology*, 325–333. <https://doi.org/10.1007/s11255-015-1181-z>
86. Zhang, X. (2004). Tryptophan Hydroxylase-2 Controls Brain Serotonin Synthesis. *Science*, 217–217. <https://doi.org/10.1126/science.1097540>
-

AR CAG Repeats and Spinal and Bulbar Muscular Atrophy

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/ar-cag-repeats-and-spinal-and-bulbar-muscular-atrophy/>

The AR CAG repeat polymorphism influences tissue response to androgens

An increase in repeats of the cytosine-adenine-guanine (CAG) trinucleotide sequence in the N-terminal domain of the androgen receptor is inhibitory of appropriate transactivation function (Chamberlain et al., 1994), entailing weaker transcriptional activity (Singh et al., 2007). Patrizio et al reported a statistically significant association between longer CAG repeats and infertility (mean length 25) when compared with healthy controls (mean length 22), particularly apparent in those with extremely severe oligozoospermia (Patrizio et al., 2001). AR CAG repeat sequence length is associated with a higher risk of symptomatic late-onset hypogonadism in men (Hong et al., 2018; Kim et al., 2018). As well as physiological outcomes, the CAGn has been associated with evolutionary-relevant male life history strategies (Gettler et al., 2017).

Huhtaniemi et al. analysed valuable and unique data from the European Male Ageing Study, comprising AR CAG repeat lengths and endocrine and clinical characteristics of nearly 3000 men aged 40–79. Coordinated by centres across Europe (Lee et al., 2009), this dataset benefits distinctly from standardisation and large sample size. Analysis revealed that, while below the 40 CAGn threshold considered denotive of SBMA (Spada et al., 1991), as the Exon 1 CAG repeat length extended, the length of the AR polyglutamine tract repeat correlated directly to all measures of serum testosterone (total, bioavailable, free) and strongly positively correlated to T and E2 in circulation. No symptoms or signs of androgen deficiency correlated to the CAG repeat length, suggesting that in the presence of greater polyQ expansion, deficiency of androgen action may be compensated for by a concomitant increase in the production of androgens under normal hypothalamic-pituitary-testicular axis conditions (Huhtaniemi et al., 2009). This compensation had similarly been hypothesised by Skjærpe et al. who also reported a positive association between CAG repeat length and free and total testosterone (Skjærpe et al., 2008).

Although not universal, assumedly due to reasons including fluctuations in testosterone levels and the cross-sectional nature of some studies (Harkonen et al., 2003), this positive correlation of longer CAG stretches with free and total testosterone is well established (Crabbe et al., 2007; Gong et al., 2014; Harkonen et al., 2003; Krithivas et al., 1999; Owens et al., 2018; Stanworth et al., 2008). Khan et al. additionally observed this in a large cohort of 400 men (Khan et al., 2018). Their study noted that the

IIEF-15 scores negatively correlated to long CAGn repeats despite higher testosterone levels, concluding that long CAGn repeats impair the effects of testosterone, particularly on erectile function. Liu et al had previously reported, in a cohort of 478 Taiwanese males aged 41 to 83, that long CAGn repeats were an independent risk factor for erectile dysfunction in men with testosterone above 3.3ng/mL but, interestingly, not 3.3ng/mL or below (Liu et al., 2015). This finding was additionally corroborated by Tirabassi et al (Tirabassi et al., 2016). Speculatively, this evidence suggests higher testosterone may exert a negative physiological effect on tissues expressing expanded CAGn AR before reaching the repeat threshold of SBMA, in which toxicity is ligand dependent. The relationship between AR CAGn and optimal function is not strictly linear: Low repeat lengths are also associated with suboptimal function. In vitro investigations by Nenonen et al. revealed a 22 CAG genotype had the highest AR-mediated transcription with the least protein compared with 16 CAG and 28 CAG. (H. Nenonen et al., 2009). In agreement, analysis of 4000 men revealed lengths close to this median confine a lower risk of infertility (H. A. Nenonen et al., 2010).

Despite the vital role of testosterone centrally (Santi et al., 2018) and peripherally for male sexual function and maintenance (Corona et al., 2016; Traish, 2008), studies of healthy men have failed to denote a relevant testosterone threshold for erectile dysfunction (Lackner et al., 2011). Androgen-induced target activities are attenuated corresponding to the length of triplet residues (Zitzmann, 2008) and the result of exogenous testosterone treatment is markedly modulated by CAG repeat polymorphism (Francomano et al., 2013). Owing to this relationship, it has been suggested that existing thresholds of hypogonadism and consequential indications are likely to be replaced with a continuum spanned by genetics and symptom specificity (Zitzmann, 2009). Recently, Escobedo et al. demonstrated the tandem CAG repeat sequence folds into a helical structure, with propensity of helicity correlating positively to sequence length. An accumulation of unconventional hydrogen bond donations from glutamine side chains to the main chain carbonyl of the residue at relative position $i+4$ confers a gain of stability to the polyQ helix and could provide a rationale for length-dependent impairment of transactivation function (Escobedo et al., 2019). Collectively, research illustrates that the available level of ligand is not an absolute determinant of optimum androgenic function, and much is dependent on its transcription factor in target tissues. The effect of agonists as beneficial or detrimental is determined specifically by the tissue of action (Narayanan et al., 2018).

SBMA

X-linked Spinal and Bulbar Muscular Atrophy, also known as Kennedy's disease, is a condition which effects multiple bodily systems and organs (Manzano et al., 2018; Sperfeld et al., 2002). SBMA is caused by an expansion of the CAG trinucleotide repeat polyglutamine tract in the first exon of the androgen receptor (Spada et al., 1991), with an excess of 38 repeats denotive of the pathogenesis (G. Querin et al., 2017).

SBMA is rare, occurring in 1 per 400,000 men per year (Fischbeck, 1997). This rarity has led to calls for the establishment of international multi-center networks to speed understanding and progress (Fratta et al., 2014; G. Querin et al., 2017). Poor clinical awareness, frequent improper diagnosis and confusion with other diseases likely result in an underestimated prevalence (G. Querin et al., 2017). In cohorts of 47 and 46 patients considered, 32% and 30% respectively had received an alternative diagnosis at first (Fratta et al., 2014; Rhodes et al., 2009). SBMA usually becomes notably symptomatic in middle age or later (Katsuno et al., 2012), however initial symptoms often begin in adolescence, long before clinical assessment (Sperfeld et al., 2002). In line with the inhibitory action of the polyglutamine tract on AR transactivation, tandem CAG repeat length has been correlated to androgen insensitivity in SBMA (Dejager et al., 2002). CAG repeat length correlates inversely with age at onset but does not always correlate to disease severity or progression (Doyu et al., 1992; Fratta et al., 2014; Andrew P. Lieberman et al., 2014; Rhodes et al., 2009). Epigenetic contributions to the late onset nature of SBMA are likely (Kondo et al., 2019). Progression is gradual and life expectancy is averagely insignificantly decreased (Chahin et al., 2008). The breadth of the clinical spectrum and involvement of testosterone target tissue likely reflects the ubiquitous expression of the androgen receptor throughout the central nervous system and peripheral tissues (H. Adachi, 2005). The complex clinical picture that results has been described by Manzano et al. as an "interplay between differentially affected tissues, which struggle to cooperate to maintain homeostasis" (Manzano et al., 2018).

Characteristic symptoms are proximal and distal weakness and proximal muscle atrophy. Bulbar muscle involvement accounts for dysarthria, dysphagia, hypernasality and decreased range of pitch and loudness (Pennuto & Rinaldi, 2018; G. Querin et al., 2017). Other common symptoms include fasciculations, cramps, tremor, reduced or absent deep tendon reflexes, loss of sensory functions in extremities, loss of vibratory sensation, tongue wasting, gynecomastia, sexual dysfunction, testicular atrophy and fertility problems including oligospermia/azoospermia. (Dahlqvist et al., 2019; Dejager et al., 2002; Fratta et al., 2014; Kennedy et al., 1968; Polo et al., 1996; G. Querin et al., 2017; Udd et al., 2009). Symptoms including gynecomastia, hand tremors, muscular cramps, myalgias, premature exhaustion during physical exercise and feet numbness can present before the onset of weakness (Finsterer & Scorza, 2019; Finsterer & Soraru, 2015). Libido loss presents and can be unappreciated due to the late onset (G. Querin et al., 2017). Abdominal obesity, dyslipidemia, glucose intolerance and liver problems represent a commonly seen metabolic involvement and these patients can frequently develop metabolic syndrome (Dejager et al., 2002; Pennuto & Rinaldi, 2018; G. Querin et al., 2017; Rosenbohm et al., 2018). Heart rhythm abnormalities including Brugada syndrome can occur (Araki et al., 2014; Giorgia Querin et al., 2015). Alterations in bone mineral density including lumbar density scores above controls, lumbar and/or femoral osteopenia, and osteoporosis are reported without correlation to LH, testosterone or vitamin D determinations. The frequency of lower urinary tract symptoms exceeds that of the general population significantly (Giorgia Querin et al., 2015). Interestingly, AR133Q knock-in mouse models exhibit significant atrophy and abnormal spontaneous myotonic discharges in the levator ani/bulbocavernosus (LABC) muscles, suggesting alteration to lower urinary tract muscle membrane excitability that could be responsible for the obstructive LUTS and associated death in these models (Yu, 2006). Hypospadias has been suggested as potentially underreported feature of the SBMA phenotype (Nordenvall et al., 2016).

While traditional focus has been on the muscular symptoms and long-associated motor neuron degeneration (Lombardi, Querin, et al., 2019), this can be misleading (Finsterer & Scorza, 2019; Sperfeld et al., 2002). The systemic, endocrinological and neuropsychological effects are now known to be of equal importance to both the clinical picture and the quality of life of patients (G. Querin et al., 2017; Giorgia Querin et al., 2018). SBMA can manifest in absence of neuromuscular complaints or symptoms, presenting with an endocrine phenotype comprising of symptoms including gynecomastia, testicular atrophy, hypercholesterolemia and diabetes mellitus (Battaglia et al., 2003). Nonclassical symptoms including erectile dysfunction can be cited by patients as amongst their most disabling symptoms (Fratta et al., 2014). Sexual dysfunction across domains including orgasm function, erectile function and satisfaction is commonly reported (Dahlqvist et al., 2019). In a large cohort of 73 patients, excluding ten patients who refused to answer, all patients were found to have mild-to-severe erectile dysfunction per IIEF (mean 15.9 ± 7.6 ; range 0–25) (Giorgia Querin et al., 2015).

SBMA patients can display peculiar psychological characteristics including diffidence, marked emotional sensitivity and concentration problems (G. Querin et al., 2017). Soukup et al. reported systematic evidence of differing frontotemporal cognitive functioning in SBMA patients compared to age and education matched controls (Soukup et al., 2009). Despite similar intelligence per IQ assessment, SBMA patients were found to significantly underperform in a battery of neuropsychological tests. Interestingly, while this varied from mild to severe impairment and "astonishingly widespread", most were subclinical in expression. Executive function and short- and long-term memory were found to be domains exhibiting pronounced deficits, while attentional control was also deficient. Consistent with prefrontal deficits, Di Rosa et al. utilised control matched neuropsychological testing, reporting a significant weakness in cognitive empathy but not in areas of affective empathy in SBMA patients. They suggest even mild impairment in mentalising may have profound implications for interpersonal relations, particularly when such changes are not recognized as the consequence of neural processes (Di Rosa et al., 2014). In a small cohort of SBMA patients, Romigi et al. reported a decrease in both subjective and objective sleep quality parameters compared with healthy age and sex matched controls. 77.8% of SBMA patients subjectively experienced disturbed sleep per the Pittsburgh Sleep Quality Index. Objectively, time in bed, total sleep time and sleep efficiency were significantly lower in SBMA patients, with a significantly higher apnea-hypopnea index. SBMA patients showed periodic limb movements. Obstructive sleep apnea was evident in a majority of patients, REM sleep without atonia was observed in 22% of patients (Romigi et al., 2014).

Although CAG repeat length is not held to be strictly associated with severity, individual case reports of patients with abnormally long CAG repeat lengths present with severe phenotypes that have expanded the clinical understanding of SBMA. Grunseich et al. reported a 29-year-old SBMA patient with a long 68 CAG repeat expansion. The patient experienced early onset of multisystemic symptoms. He had been born with penile congenital abnormality. He developed gynecomastia by 16 and muscle weakness, fatigue after exercise, fasciculations, cramping, and tremor by age 18. He experienced ejaculation difficulties, testicular atrophy, burning neuropathic pain and dysesthesia in the feet and fingertips, reduced sweating and decreased facial hair growth. Tongue atrophy was noted, and weakness was observed in the

orbicularis oculi and orbicularis oris. He exhibited perioral fasciculations, severe limb weakness bilaterally, difficulty standing on his heels and ankles, and loss of temperature and vibratory sensation in the fingers and toes. Abnormalities were seen on muscle MRI. Evidence of autonomic dysfunction suggestive of small fiber dysfunction was determined, including negligible sweat responses and orthostatic tachycardia without blood pressure changes or symptoms (Grunseich et al., 2014). Similarly, Madeira et al., reported a phenotype of an exceptional 72 CAG repeat length. This man was 53 years old and underweight. He complained of shortness of breath, difficulty breathing while lying down and paroxysmal nocturnal dyspnea. He had a micropenis, small testicles and progressive testicular failure. Deep tendon reflexes were absent. Fasciculations, weakness and atrophy were apparent in the tongue, masseter muscles and limb muscles. Neck muscles were severely weakened. He had osteopenia, with low bone mass densities in the lumbar spine and femoral neck. He additionally had dyslipidaemia (Madeira et al., 2017). These phenotypical presentations highlight the broad effects associated with alteration of androgen-dependant signaling pathways.

Reliable biomarkers for SBMA remain a challenge (Manzano et al., 2018; Giorgia Querin et al., 2018), but common findings have been established. Creatine-Kinase will often be elevated (G. Querin et al., 2017). Testosterone, LH and FSH are generally found to be within normal bounds, although T and DHT can be high or low in some patients (Hashizume et al., 2012; Ni et al., 2015; Giorgia Querin et al., 2015; Rhodes et al., 2009). Patterns of androgen insensitivity per biomarkers are seen in some patients as per the Androgen Sensitivity Index, and have been reported to correlate positively with CAG repeats (Dejager et al., 2002; Giorgia Querin et al., 2015). High proportions of patients will show lipid and metabolic abnormality, including elevated total cholesterol, triglycerides, fasting glucose and insulin (Dejager et al., 2002; Francini-Pesenti et al., 2018; Guber et al., 2017; Giorgia Querin et al., 2015). Signs of non-alcoholic fatty liver disease including excess deposition of triacylglycerol in the liver have been reported as a near universal finding, even in patients with normal BMI (Guber et al., 2017). The observation that hepatic AR knockout models that develop steatosis and insulin resistance (Lin et al., 2008), as well as multisystem disruption of metabolic homeostasis, is suggestive of direct disease involvement in the observed NAFLD in SBMA patients. Serum hydroxyvitamin D was reported as low in a majority of patients in a large cohort (Giorgia Querin et al., 2015). Interestingly, the markers of neuronal damage phosphorylated neurofilament heavy chain and neurofilament light chain levels are not elevated in serum of SBMA patients or animal models and do not correlate with phenotypical severity (Lombardi, Bombaci, et al., 2019; Lombardi, Querin, et al., 2019).

Muscle involvement is diffuse. Myopathic evidence present upon muscle biopsy (Manzano et al., 2018) is supportive of a conserved pathological mechanism that likely underlies a vast proportion of clinical manifestations (Baniahmad, 2015; G. Querin et al., 2017). In a 40-patient cohort, muscle fat content was significantly higher than controls in the semitendinosus, semimembranosus, biceps femoris longus, triceps surae and spared sartorius, gracilis, biceps femoris brevis, and tibialis anterior. Affected leg muscles showed greater involvement than arm muscles, and muscle fat content correlated to muscle strength and function tests, disease duration and severity, and creatine kinase and testosterone levels (Dahlqvist et al., 2019). White matter alterations in the corticospinal tracts, limbic system, brainstem and cerebellum have been demonstrated via quantitative brain imaging (Kassubek et al., 2007; Unrath et

al., 2010)?, while voxel based morphometry has identified gray matter atrophy in the frontal lobes and brainstem (Pieper et al., 2012)?. Skeletal muscle, known to be AR enriched, is a notable site of toxicity and tissue biopsy has demonstrated denervation, muscle fiber degeneration and myogenic changes in addition to neurogenic atrophy (Giorgia Querin et al., 2015; Sorarù et al., 2008)?. Somatosensory evoked potentials are regularly abnormal, while electromyography and nerve conduction study will often reveal low sensory nerve amplitudes, decreased compound motor action potentials and evidence of diffuse denervation (BUECKING, 2000; Kachi et al., 1992; Pennuto & Rinaldi, 2018; Polo et al., 1996)?. Broad involvement of sensory neurons and autonomic skin denervation were reported with abnormal sweat test results (Manganelli et al., 2007)?. These findings align with AR accumulation and degeneration in autonomic regions including the dorsal root ganglia (Antonini et al., 2000)?.

The mechanisms of PolyQ AR toxicity are yet to be fully elucidated but it appears that levels of AR expression are directly correlated to muscular atrophy (Manzano et al., 2018)?. Both testosterone or DHT binding to the polyQ AR and its subsequent translocation of the expanded protein to the nucleus is required for toxicity as demonstrated in vivo (Katsuno et al., 2002; Nedelsky et al., 2010; Takeyama et al., 2002)? and in vitro (Becker et al., 2000; Darrington et al., 2002; Stenoien et al., 1999; Walcott & Merry, 2002)?. Higher androgen levels in males are therefore responsible for the symptomatic presentation, and female carriers will ordinarily remain asymptomatic (Chevalier-Larsen, 2004; Schmidt et al., 2002)?. In humans, exogenous androgen administration does not usually relieve clinical symptoms (Neuschmid-Kaspar et al., 1996)? and has been reported to have reversibly worsened symptoms (Kinirons & Rouleau, 2008)?. Administering testosterone to previously asymptomatic transgene SBMA female mice induces a distinct increase of symptoms similar to the level of untreated males, including progressive emaciation and motor dysfunction, pathological markers and nuclear localisation of pathogenic AR (Katsuno et al., 2002)?, demonstrating the androgen dependency of the pathology.

AR polyQ expansion involves a partial loss of the normal transcriptional activity of the AR (Chamberlain et al., 1994; Kazemi-Esfarjani et al., 1995; A. P. Lieberman, 2002; Mhatre et al., 1993)? and this contributes to the pathology. However, neither loss of AR function nor androgen ablation is adequate for the pathology, and men with complete androgen insensitivity syndrome do not exhibit neurological symptoms (Chivet et al., 2019; Quigley et al., 1992)?. As such, the disease entails a proteotoxic gain of function (A. P. Lieberman, 2002; Manzano et al., 2018; Nath et al., 2018; Pennuto & Rinaldi, 2018)?. The mutant AR disrupts many downstream pathways, and alteration of diverse cellular processes including transcription, RNA splicing, axonal transport, ion homeostasis, and mitochondrial function likely coalesce to cause toxicity (Borgia et al., 2017; Chua et al., 2015; Eftekharzadeh et al., 2019; Malik et al., 2019)?. Diffuse nuclear accumulation of mutant AR is frequent and extensive in SBMA, occurring in a wide array of CNS nuclei and visceral organs (H. Adachi, 2005; Doi et al., 2013; Katsuno et al., 2002)?. Nuclear accumulation of AR is reported to be important to the pathology (Nedelsky et al., 2010)?. Animal models have revealed export of the pathogenic AR protein is impaired in the absence of any cell-wide disruption of nucleocytoplasmic transport (Arnold et al., 2019)?. Significant age, hormone and CAG repeat length dependent impairment of multiple ubiquitin-proteasome genes have been demonstrated to result from a toxic gain of AR function, progressively compounding toxicity through a failure of polyQ AR clearance. Diminished expression of numerous components of the

ubiquitin-proteasome pathway including ubiquitin receptors, proteolytic subunits and assembly scaffold proteins were recently reported in skeletal muscle of AR113Q male mice. This involved significant reduction of ~30% of constitutive proteasome subunits and ~20% of E2 ubiquitin conjugating enzymes, with no upregulation observed and a non-significant trend towards reduced expression in many more subunits (Nath et al., 2018). This differentiates AR-mediated toxicity from skeletal muscle atrophy following cachexia, renal failure, surgical denervation, fasting, tumors, and diabetes, which all exhibit an up-regulation of proteasome subunits (Sacheck et al., 2006).

Using cell culture and animal models, androgen axis targeted therapeutic strategies have been explored. Androgen ablation and treatment with AR antagonists are beneficial and ameliorate the SBMA pathogenicity (Baniahmad, 2015), demonstrating phenotypical improvement beyond simply a slowing of the disease progression. The antiandrogen flutamide was protective of androgen-mediated toxicity across several SBMA models, preventing or reversing motor dysfunction of transgene models and extending the life of knock-in males significantly (Renier et al., 2014). Similarly, castration of AR97Q males dramatically prevented phenotypical presentation, with these mice showing significantly extended life, ameliorated muscle atrophy and body size reduction, virtually absent motor impairment, and markedly reduced nuclear AR staining intensities as compared to sham operated AR97Q mice displaying significant pathology (Katsuno et al., 2002). Castration was also remarkably effective in 112 and 113 glutamine models (Chevalier-Larsen, 2004; Nath et al., 2018). Leuprorelin has also been demonstrated as effective in transgenic mice (Katsuno et al., 2003). 14 years of prospective quantitative measurement of a single SBMA patient who underwent leuprolide acetate treatment for the initial 7 years before undergoing orchiectomy indicated that long term androgen deprivation slows disease progression when compared to existing control data (Hijikata et al., 2019). In transgenic mice, SBMA symptoms have been shown to be ameliorated through IGF-1 treatment or overexpression in muscle, which promotes AR degradation through phosphorylation by Akt (Palazzolo et al., 2009; Rinaldi et al., 2012). Treatment with genistein, an antiandrogenic soy isoflavone, was demonstrated to promote dissociation of the AR from the co-regulator ARA70 and attenuate pathology and improved survival in 97Q mouse models (Qiang et al., 2013). Modulation of activation function-2 of the AR with the compound MEPB rescued toxicity in a drosophila model of SBMA and showed a dose-dependent rescue from loss of body weight, rotarod activity and grip strength, neuronal loss, neurogenic atrophy and reversed testicular atrophy in a SBMA mouse model (Badders et al., 2018). It is likely that the new generation of Selective Androgen Receptor Degraders in development for use in castration resistant prostate cancer (Han et al., 2019; Ponnusamy et al., 2017) will be of interest with regard to a potential treatment for SBMA. ASC-J9, an AR degrader enhancer with structural similarity to curcumin (Cheng et al., 2018), has already been shown to rescue SBMA motor symptoms and improve sexual function in transgenic 97Q mice (Yang et al., 2007).

In cell models, targeting the heat shock protein families, molecular chaperones to the AR, suppresses aggregation and enhances polyQ AR degradation, making them a potential therapeutic target (Bailey, 2002). Mutant AR forms a Hsp90 chaperone complex preferentially compared to wild type AR, and use of a Hsp90 inhibitor, Tanespimycin, has been demonstrated to be effective at degrading polyQ AR in vitro and in vivo modelling, markedly ameliorating motor impairment (Waza et al., 2005).

Tanespimycin, however, has broad interruptive effects and is not well tolerated (Yang et al., 2007), and Hsp90 inhibitors can induce the degradation of hundreds of client proteins that are likely needed for diverse processes (Eftekharzadeh et al., 2019). Recently, Eftekharzadeh et al. suggested that Hsp70 activation with small molecules such as JG-98 or JG-294 is a safer potential approach to leveraging protein quality control mechanisms to degrade the AR in SBMA and other androgen-mediated conditions (Eftekharzadeh et al., 2019). Hsp70 overexpression is similarly seen to significantly ameliorate SBMA symptoms in a transgenic mouse model by reducing the amount of nuclear-localized mutant AR protein (Hiroaki Adachi et al., 2003). Arimoclomol, a co-inducer of the heat shock response limited to stressed cells, has been observed to delay disease progression in a mouse model of SBMA through the prevention of motor neuron degeneration and alleviation of muscle atrophy (Rinaldi et al., 2015). Trehalose has been suggested as a potential therapeutic agent, and in vitro studies suggest beneficial effects resulting from increased autophagic clearance of the mutant AR (Cicardi et al., 2019). Sodium butyrate, a histone deacetylase inhibitor capable of modulating AR expression (Paskova et al., 2013), showed improvement in motor deficits and histopathological impairment of neurons and muscle within an narrow optimum dose window in transgenic mice (Minamiyama, 2004). Inhibition of Src kinase, a pathway upregulated by polyglutamine expansion and AR overexpression, has been demonstrated to mitigate toxicity in SBMA animal and cell models (Iida et al., 2019).

Given the significant advancement in the understanding of the pathological mechanisms, a move towards targeted molecular therapies addressing the systemic pathological processes is likely in the near future (Giorgia Querin et al., 2018). To achieve a disease modifying therapy for SBMA, Rinaldi et al. suggest a coordinated, collaborative effort of researchers with multiple areas of expertise, clinicians, the pharmaceutical industry and the involvement of patient groups (Rinaldi et al., 2015).

Page Bibliography

1. Adachi, H. (2005). Widespread nuclear and cytoplasmic accumulation of mutant androgen receptor in SBMA patients. *Brain*, 659–670. <https://doi.org/10.1093/brain/awh381>
2. Adachi, Hiroaki, Katsuno, M., Minamiyama, M., Sang, C., Pagoulatos, G., Angelidis, C., Kusakabe, M., Yoshiki, A., Kobayashi, Y., Doyu, M., & Sobue, G. (2003). Heat Shock Protein 70 Chaperone Overexpression Ameliorates Phenotypes of the Spinal and Bulbar Muscular Atrophy Transgenic Mouse Model by Reducing Nuclear-Localized Mutant Androgen Receptor Protein. *The Journal of Neuroscience*, 2203–2211. <https://doi.org/10.1523/jneurosci.23-06-02203.2003>
3. Antonini, G., Gragnani, F., Romaniello, A., Pennisi, E. M., Morino, S., Ceschin, V., Santoro, L., & Cruccu, G. (2000). Sensory involvement in spinal-bulbar muscular atrophy (Kennedy's disease). *Muscle & Nerve*, 252–258. [https://doi.org/10.1002/\(sici\)1097-4598\(200002\)23:2<252::aid-mus17>3.0.co;2-p](https://doi.org/10.1002/(sici)1097-4598(200002)23:2<252::aid-mus17>3.0.co;2-p)

4. Araki, A., Katsuno, M., Suzuki, K., Banno, H., Suga, N., Hashizume, A., Mano, T., Hijikata, Y., Nakatsuji, H., Watanabe, H., Yamamoto, M., Makiyama, T., Ohno, S., Fukuyama, M., Morimoto, S. -i., Horie, M., & Sobue, G. (2014). Brugada syndrome in spinal and bulbar muscular atrophy. *Neurology*, 1813–1821. <https://doi.org/10.1212/wnl.0000000000000434>
5. Arnold, F. J., Pluciennik, A., & Merry, D. E. (2019). Impaired Nuclear Export of Polyglutamine-Expanded Androgen Receptor in Spinal and Bulbar Muscular Atrophy. *Scientific Reports*. <https://doi.org/10.1038/s41598-018-36784-4>
6. Badders, N. M., Korff, A., Miranda, H. C., Vuppala, P. K., Smith, R. B., Winborn, B. J., Quemin, E. R., Sopher, B. L., Dearman, J., Messing, J., Kim, N. C., Moore, J., Freibaum, B. D., Kanagaraj, A. P., Fan, B., Tillman, H., Chen, P.-C., Wang, Y., III, B. B. F., ... Taylor, J. P. (2018). Selective modulation of the androgen receptor AF2 domain rescues degeneration in spinal bulbar muscular atrophy. *Nature Medicine*, 427–437. <https://doi.org/10.1038/nm.4500>
7. Bailey, C. K. (2002). Molecular chaperones enhance the degradation of expanded polyglutamine repeat androgen receptor in a cellular model of spinal and bulbar muscular atrophy. *Human Molecular Genetics*, 515–523. <https://doi.org/10.1093/hmg/11.5.515>
8. Baniahmad, A. (2015). Inhibition of the Androgen Receptor by Antiandrogens in Spinobulbar Muscle Atrophy. *Journal of Molecular Neuroscience*, 343–347. <https://doi.org/10.1007/s12031-015-0681-8>
9. Battaglia, F., Le Galudec, V., Cossee, M., Tranchant, C., Warter, J. M., & Echaniz-Laguna, A. (2003). Kennedy's Disease Initially Manifesting as an Endocrine Disorder. *Journal of Clinical Neuromuscular Disease*, 165–167. <https://doi.org/10.1097/00131402-200306000-00001>
10. Becker, M., Martin, E., Schneikert, J., Krug, H. F., & Cato, A. C. B. (2000). Cytoplasmic Localization and the Choice of Ligand Determine Aggregate Formation by Androgen Receptor with Amplified Polyglutamine Stretch. *Journal of Cell Biology*, 255–262. <https://doi.org/10.1083/jcb.149.2.255>
11. Borgia, D., Malena, A., Spinazzi, M., Andrea Desbats, M., Salviati, L., Russell, A. P., Miotto, G., Tosatto, L., Pegoraro, E., Sorarù, G., Pennuto, M., & Vergani, L. (2017). Increased mitophagy in the skeletal muscle of spinal and bulbar muscular atrophy patients. *Human Molecular Genetics*, ddx019. <https://doi.org/10.1093/hmg/ddx019>

12. BUECKING, A. (2000). Sensory ataxia as the initial clinical symptom in X-linked recessive bulbospinal neuronopathy. *Journal of Neurology, Neurosurgery & Psychiatry*, 277–277. <https://doi.org/10.1136/jnmp.69.2.277>
13. Chahin, N., Klein, C., Mandrekar, J., & Sorenson, E. (2008). Natural history of spinal-bulbar muscular atrophy. *Neurology*, 1967–1971. <https://doi.org/10.1212/01.wnl.0000312510.49768.eb>
14. Chamberlain, N. L., Driver, E. D., & Miesfeld, R. L. (1994). The length and location of CAG trinucleotide repeats in the androgen receptor N-terminal domain affect transactivation function. *Nucleic Acids Research*, 3181–3186. <https://doi.org/10.1093/nar/22.15.3181>
15. Cheng, M. A., Chou, F.-J., Wang, K., Yang, R., Ding, J., Zhang, Q., Li, G., Yeh, S., Xu, D., & Chang, C. (2018). Androgen receptor (AR) degradation enhancer ASC-J9® in an FDA-approved formulated solution suppresses castration resistant prostate cancer cell growth. *Cancer Letters*, 182–191. <https://doi.org/10.1016/j.canlet.2017.11.038>
16. Chevalier-Larsen, E. S. (2004). Castration Restores Function and Neurofilament Alterations of Aged Symptomatic Males in a Transgenic Mouse Model of Spinal and Bulbar Muscular Atrophy. *Journal of Neuroscience*, 4778–4786. <https://doi.org/10.1523/jneurosci.0808-04.2004>
17. Chivet, M., Marchioretta, C., Pirazzini, M., Piol, D., Scaramuzzino, C., Polanco, J., Nath, S., Zuccaro, E., Nogara, L., Canato, M., Marcucci, L., Parodi, S., Romanello, V., Armani, A., D'Antonio, M., Sambataro, F., Dassi, E., Pegoraro, E., Sorarù, G., ... Pennuto, M. (2019). *Polyglutamine-expanded androgen receptor disrupts muscle triad, calcium dynamics and the excitation-contraction coupling gene expression program*. Cold Spring Harbor Laboratory. <https://doi.org/10.1101/618405>
18. Chua, J. P., Reddy, S. L., Yu, Z., Giorgetti, E., Montie, H. L., Mukherjee, S., Higgins, J., McEachin, R. C., Robins, D. M., Merry, D. E., Iñiguez-Lluhí, J. A., & Lieberman, A. P. (2015). Disrupting SUMOylation enhances transcriptional function and ameliorates polyglutamine androgen receptor-mediated disease. *Journal of Clinical Investigation*, 831–845. <https://doi.org/10.1172/jci73214>
19. Cicardi, M. E., Cristofani, R., Crippa, V., Ferrari, V., Tedesco, B., Casarotto, E., Chierichetti, M., Galbiati, M., Piccolella, M., Messi, E., Carra, S., Pennuto, M., Rusmini, P., & Poletti, A. (2019). Autophagic and Proteasomal Mediated Removal of Mutant Androgen Receptor in Muscle Models of Spinal and Bulbar Muscular Atrophy. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2019.00569>

20. Corona, G., Isidori, A. M., Aversa, A., Burnett, A. L., & Maggi, M. (2016). Endocrinologic Control of Men's Sexual Desire and Arousal/Erection. *The Journal of Sexual Medicine*, 317–337. <https://doi.org/10.1016/j.jsxm.2016.01.007>

21. Crabbe, P., Bogaert, V., De Bacquer, D., Goemaere, S., Zmierzak, H., & Kaufman, J. M. (2007). Part of the Interindividual Variation in Serum Testosterone Levels in Healthy Men Reflects Differences in Androgen Sensitivity and Feedback Set Point: Contribution of the Androgen Receptor Polyglutamine Tract Polymorphism. *The Journal of Clinical Endocrinology & Metabolism*, 3604–3610. <https://doi.org/10.1210/jc.2007-0117>

22. Dahlqvist, J. R., Oestergaard, S. T., Poulsen, N. S., Thomsen, C., & Vissing, J. (2019). Refining the spinobulbar muscular atrophy phenotype by quantitative MRI and clinical assessments. *Neurology*, e548–e559. <https://doi.org/10.1212/wnl.0000000000006887>

23. Darrington, R. S., Butler, R., Leigh, P. N., McPhaul, M. J., & Gallo, J.-M. (2002). Ligand-dependent aggregation of polyglutamine-expanded androgen receptor in neuronal cells. *NeuroReport*, 2117–2120. <https://doi.org/10.1097/00001756-200211150-00025>

24. Dejager, S., Bry-Gauillard, H., Bruckert, E., Eymard, B., Salachas, F., LeGuern, E., Tardieu, S., Chadarevian, R., Giral, P., & Turpin, G. (2002). A Comprehensive Endocrine Description of Kennedy's Disease Revealing Androgen Insensitivity Linked to CAG Repeat Length. *The Journal of Clinical Endocrinology & Metabolism*, 3893–3901. <https://doi.org/10.1210/jcem.87.8.8780>

25. Di Rosa, E., Sorarù, G., Kleinbub, J. R., Calvo, V., Vallesi, A., Querin, G., Marcato, S., Grasso, I., & Palmieri, A. (2014). Theory of mind, empathy and neuropsychological functioning in X-linked Spinal and Bulbar Muscular Atrophy: a controlled study of 20 patients. *Journal of Neurology*, 394–401. <https://doi.org/10.1007/s00415-014-7567-5>

26. Doi, H., Adachi, H., Katsuno, M., Minamiyama, M., Matsumoto, S., Kondo, N., Miyazaki, Y., Iida, M., Tohnai, G., Qiang, Q., Tanaka, F., Yanagawa, T., Warabi, E., Ishii, T., & Sobue, G. (2013). p62/SQSTM1 Differentially Removes the Toxic Mutant Androgen Receptor via Autophagy and Inclusion Formation in a Spinal and Bulbar Muscular Atrophy Mouse Model. *Journal of Neuroscience*, 7710–7727. <https://doi.org/10.1523/jneurosci.3021-12.2013>

27. Doyu, M., Sobue, G., Mukai, E., Kachi, T., Yasuda, T., Mitsuma, T., & Takahashi, A. (1992). Severity of X-linked recessive bulbospinal neuronopathy correlates with size of the tandem cag repeat in androgen receptor gene. *Annals of Neurology*, 707–710.

<https://doi.org/10.1002/ana.410320517>

28. Eftekharzadeh, B., Banduseela, V. C., Chiesa, G., Martínez-Cristóbal, P., Rauch, J. N., Nath, S. R., Schwarz, D. M. C., Shao, H., Marin-Argany, M., Di Sanza, C., Giorgetti, E., Yu, Z., Pierattelli, R., Felli, I. C., Brun-Heath, I., García, J., Nebreda, Á. R., Gestwicki, J. E., Lieberman, A. P., & Salvatella, X. (2019). Hsp70 and Hsp40 inhibit an inter-domain interaction necessary for transcriptional activity in the androgen receptor. *Nature Communications*. <https://doi.org/10.1038/s41467-019-11594-y>
29. Escobedo, A., Topal, B., Kunze, M. B. A., Aranda, J., Chiesa, G., Mungianu, D., Bernardo-Seisdedos, G., Eftekharzadeh, B., Gairí, M., Pierattelli, R., Felli, I. C., Diercks, T., Millet, O., García, J., Orozco, M., Crehuet, R., Lindorff-Larsen, K., & Salvatella, X. (2019). Side chain to main chain hydrogen bonds stabilize a polyglutamine helix in a transcription factor. *Nature Communications*. <https://doi.org/10.1038/s41467-019-09923-2>
30. Finsterer, J., & Scorza, F. A. (2019). Central nervous system abnormalities in spinal and bulbar muscular atrophy (Kennedy's disease). *Clinical Neurology and Neurosurgery*, 105426. <https://doi.org/10.1016/j.clineuro.2019.105426>
31. Finsterer, J., & Soraru, G. (2015). Onset Manifestations of Spinal and Bulbar Muscular Atrophy (Kennedy's Disease). *Journal of Molecular Neuroscience*, 321–329. <https://doi.org/10.1007/s12031-015-0663-x>
32. Fischbeck, K. H. (1997). *Journal of Inherited Metabolic Disease*, 152–158. <https://doi.org/10.1023/a:1005344403603>
33. Francini-Pesenti, F., Querin, G., Martini, C., Mareso, S., & Sacerdoti, D. (2018). Prevalence of metabolic syndrome and non-alcoholic fatty liver disease in a cohort of Italian patients with spinal-bulbar muscular atrophy. *Acta Myologica : Myopathies and Cardiomyopathies : Official Journal of the Mediterranean Society of Myology*, 37(3), 204–209. <https://www.ncbi.nlm.nih.gov/pubmed/30838350>
34. Francomano, D., Greco, E. A., Lenzi, A., & Aversa, A. (2013). CAG Repeat Testing of Androgen Receptor Polymorphism: Is This Necessary for the Best Clinical Management of Hypogonadism? *The Journal of Sexual Medicine*, 2373–2381. <https://doi.org/10.1111/jsm.12268>
35. Fratta, P., Nirmalanathan, N., Masset, L., Skorupinska, I., Collins, T., Cortese, A., Pemble, S., Malaspina, A., Fisher, E. M. C., Greensmith, L., & Hanna, M. G. (2014). Correlation of clinical

and molecular features in spinal bulbar muscular atrophy. *Neurology*, 2077–2084.

<https://doi.org/10.1212/wnl.0000000000000507>

36. Gettler, L. T., Ryan, C. P., Eisenberg, D. T. A., Rzhetskaya, M., Hayes, M. G., Feranil, A. B., Bechayda, S. A., & Kuzawa, C. W. (2017). The role of testosterone in coordinating male life history strategies: The moderating effects of the androgen receptor CAG repeat polymorphism. *Hormones and Behavior*, 164–175. <https://doi.org/10.1016/j.yhbeh.2016.10.012>
37. Gong, Y.-G., He, D.-L., Ma, Y.-M., Wu, K.-J., Ning, L., Zeng, J., Kou, B., Xie, H.-J., Ma, Z.-K., & Wang, X.-Y. (2014). Relationships among androgen receptor CAG repeat polymorphism, sex hormones and penile length in Han adult men from China: a cross-sectional study. *Asian Journal of Andrology*, 478. <https://doi.org/10.4103/1008-682x.124560>
38. Grunseich, C., Kats, I. R., Bott, L. C., Rinaldi, C., Kokkinis, A., Fox, D., Chen, K., Schindler, A. B., Mankodi, A. K., Shrader, J. A., Schwartz, D. P., Lehky, T. J., Liu, C.-Y., & Fischbeck, K. H. (2014). Early onset and novel features in a spinal and bulbar muscular atrophy patient with a 68 CAG repeat. *Neuromuscular Disorders*, 978–981. <https://doi.org/10.1016/j.nmd.2014.06.441>
39. Guber, R. D., Takyar, V., Kokkinis, A., Fox, D. A., Alao, H., Kats, I., Bakar, D., Remaley, A. T., Hewitt, S. M., Kleiner, D. E., Liu, C.-Y., Hadigan, C., Fischbeck, K. H., Rotman, Y., & Grunseich, C. (2017). Nonalcoholic fatty liver disease in spinal and bulbar muscular atrophy. *Neurology*, 2481–2490. <https://doi.org/10.1212/wnl.00000000000004748>
40. Han, X., Wang, C., Qin, C., Xiang, W., Fernandez-Salas, E., Yang, C.-Y., Wang, M., Zhao, L., Xu, T., Chinnaswamy, K., Delproposto, J., Stuckey, J., & Wang, S. (2019). Discovery of ARD-69 as a Highly Potent Proteolysis Targeting Chimera (PROTAC) Degradar of Androgen Receptor (AR) for the Treatment of Prostate Cancer. *Journal of Medicinal Chemistry*, 941–964. <https://doi.org/10.1021/acs.jmedchem.8b01631>
41. Harkonen, K., Huhtaniemi, I., Makinen, J., Hubler, D., Irjala, K., Koskenvuo, M., Oettel, M., Raitakari, O., Saad, F., & Pollanen, P. (2003). The polymorphic androgen receptor gene CAG repeat, pituitary-testicular function and andropausal symptoms in ageing men. *International Journal of Andrology*, 187–194. <https://doi.org/10.1046/j.1365-2605.2003.00415.x>
42. Hashizume, A., Katsuno, M., Banno, H., Suzuki, K., Suga, N., Mano, T., Atsuta, N., Oe, H., Watanabe, H., Tanaka, F., & Sobue, G. (2012). Longitudinal changes of outcome measures in spinal and bulbar muscular atrophy. *Brain*, 2838–2848. <https://doi.org/10.1093/brain/aws170>

43. Hijikata, Y., Hashizume, A., Yamada, S., Ito, D., Banno, H., Suzuki, K., Sobue, G., & Katsuno, M. (2019). Long-term Effects of Androgen Deprivation in a Patient with Spinal and Bulbar Muscular Atrophy - A Case Report with 14 Years of Follow-up. *Internal Medicine*, 2231–2234. <https://doi.org/10.2169/internalmedicine.1592-18>
44. Hong, Z., Xu, Q., Mao, Y., Ye, Y., Mao, J., Wan, M., & Jiang, M. (2018). Polymorphism of the androgen receptor gene CAG repeat sequence and male climacteric syndrome. *Journal of Biological Regulators and Homeostatic Agents*, 32(4), 915–921. <https://www.ncbi.nlm.nih.gov/pubmed/30043577>
45. Huhtaniemi, I. T., Pye, S. R., Limer, K. L., Thomson, W., O'Neill, T. W., Platt, H., Payne, D., John, S. L., Jiang, M., Boonen, S., Borghs, H., Vanderschueren, D., Adams, J. E., Ward, K. A., Bartfai, G., Casanueva, F., Finn, J. D., Forti, G., ... Giwercman, A. (2009). Increased Estrogen Rather Than Decreased Androgen Action Is Associated with Longer Androgen Receptor CAG Repeats. *The Journal of Clinical Endocrinology & Metabolism*, 277–284. <https://doi.org/10.1210/jc.2008-0848>
46. Iida, M., Sahashi, K., Kondo, N., Nakatsuji, H., Tohnai, G., Tsutsumi, Y., Noda, S., Murakami, A., Onodera, K., Okada, Y., Nakatochi, M., Tsukagoshi Okabe, Y., Shimizu, S., Mizuno, M., Adachi, H., Okano, H., Sobue, G., & Katsuno, M. (2019). Src inhibition attenuates polyglutamine-mediated neuromuscular degeneration in spinal and bulbar muscular atrophy. *Nature Communications*. <https://doi.org/10.1038/s41467-019-12282-7>
47. Kachi, T., Sobue, G., & Sobue, I. (1992). Central motor and sensory conduction in X-linked recessive bulbospinal neuronopathy. *Journal of Neurology, Neurosurgery & Psychiatry*, 394–397. <https://doi.org/10.1136/jnnp.55.5.394>
48. Kassubek, J., Juengling, F. D., & Sperfeld, A.-D. (2007). Widespread white matter changes in Kennedy disease: a voxel based morphometry study. *Journal of Neurology, Neurosurgery & Psychiatry*, 1209–1212. <https://doi.org/10.1136/jnnp.2006.112532>
49. Katsuno, M., Adachi, H., Doyu, M., Minamiyama, M., Sang, C., Kobayashi, Y., Inukai, A., & Sobue, G. (2003). Leuprorelin rescues polyglutamine-dependent phenotypes in a transgenic mouse model of spinal and bulbar muscular atrophy. *Nature Medicine*, 768–773. <https://doi.org/10.1038/nm878>
50. Katsuno, M., Adachi, H., Kume, A., Li, M., Nakagomi, Y., Niwa, H., Sang, C., Kobayashi, Y., Doyu, M., & Sobue, G. (2002). Testosterone Reduction Prevents Phenotypic Expression in a Transgenic Mouse Model of Spinal and Bulbar Muscular Atrophy. *Neuron*, 843–854.

[https://doi.org/10.1016/s0896-6273\(02\)00834-6](https://doi.org/10.1016/s0896-6273(02)00834-6)

51. Katsuno, M., Tanaka, F., Adachi, H., Banno, H., Suzuki, K., Watanabe, H., & Sobue, G. (2012). Pathogenesis and therapy of spinal and bulbar muscular atrophy (SBMA). *Progress in Neurobiology*, 246–256. <https://doi.org/10.1016/j.pneurobio.2012.05.007>
52. Kazemi-Esfarjani, P., Trifiro, M. A., & Pinsky, L. (1995). Evidence for a repressive function of the long polyglutamine tract in the human androgen receptor: possible pathogenetic relevance for the (CAG)_n-expanded neuronopathies. *Human Molecular Genetics*, 523–527. <https://doi.org/10.1093/hmg/4.4.523>
53. Kennedy, W. R., Alter, M., & Sung, J. H. (1968). Progressive proximal spinal and bulbar muscular atrophy of late onset: A sex-linked recessive trait. *Neurology*, 671–671. <https://doi.org/10.1212/wnl.18.7.671>
54. Khan, H. L., Bhatti, S., Abbas, S., Khan, Y. L., Gonzalez, R. M. M., Aslamkhan, M., Gonzalez, G. R., & Aydin, H. H. (2018). Longer trinucleotide repeats of androgen receptor are associated with higher testosterone and low oxytocin levels in diabetic premature ejaculatory dysfunction patients. *Basic and Clinical Andrology*. <https://doi.org/10.1186/s12610-018-0068-0>
55. Kim, J. W., Bae, Y. D., Ahn, S. T., Kim, J. W., Kim, J. J., & Moon, D. G. (2018). Androgen Receptor CAG Repeat Length as a Risk Factor of Late-Onset Hypogonadism in a Korean Male Population. *Sexual Medicine*, 203–209. <https://doi.org/10.1016/j.esxm.2018.04.002>
56. Kinirons, P., & Rouleau, G. A. (2008). Administration of testosterone results in reversible deterioration in Kennedy's disease. *Journal of Neurology, Neurosurgery & Psychiatry*, 106–107. <https://doi.org/10.1136/jnnp.2006.101899>
57. Kondo, N., Tohnai, G., Sahashi, K., Iida, M., Kataoka, M., Nakatsuji, H., Tsutsumi, Y., Hashizume, A., Adachi, H., Koike, H., Shinjo, K., Kondo, Y., Sobue, G., & Katsuno, M. (2019). DNA methylation inhibitor attenuates polyglutamine-induced neurodegeneration by regulating Hes5. *EMBO Molecular Medicine*. <https://doi.org/10.15252/emmm.201708547>
58. Krithivas, K., Yurgalevitch, S., Mohr, B., Wilcox, C., Batter, S., Brown, M., Longcope, C., McKinlay, J., & Kantoff, P. (1999). Evidence that the CAG repeat in the androgen receptor gene is associated with the age-related decline in serum androgen levels in men. *Journal of Endocrinology*, 137–142. <https://doi.org/10.1677/joe.0.1620137>

59. Lackner, J. E., Rücklinger, E., Schatzl, G., Lunglmayr, G., & Kratzik, C. W. (2011). Are there symptom-specific testosterone thresholds in aging men? *BJU International*, 1310–1315. <https://doi.org/10.1111/j.1464-410x.2010.09986.x>
60. Lee, D. M., O'Neill, T. W., Pye, S. R., Silman, A. J., Finn, J. D., Pendleton, N., Tajar, A., Bartfai, G., Casanueva, F., Forti, G., Giwercman, A., Huhtaniemi, I. T., Kula, K., Punab, M., Boonen, S., Vanderschueren, D., & Wu, F. C. W. (2009). The European Male Ageing Study (EMAS): design, methods and recruitment. *International Journal of Andrology*, 11–24. <https://doi.org/10.1111/j.1365-2605.2008.00879.x>
61. Lieberman, A. P. (2002). Altered transcriptional regulation in cells expressing the expanded polyglutamine androgen receptor. *Human Molecular Genetics*, 1967–1976. <https://doi.org/10.1093/hmg/11.17.1967>
62. Lieberman, Andrew P., Yu, Z., Murray, S., Peralta, R., Low, A., Guo, S., Yu, X. X., Cortes, C. J., Bennett, C. F., Monia, B. P., La Spada, A. R., & Hung, G. (2014). Peripheral Androgen Receptor Gene Suppression Rescues Disease in Mouse Models of Spinal and Bulbar Muscular Atrophy. *Cell Reports*, 774–784. <https://doi.org/10.1016/j.celrep.2014.02.008>
63. Lin, H.-Y., Yu, I.-C., Wang, R.-S., Chen, Y.-T., Liu, N.-C., Altuwajri, S., Hsu, C.-L., Ma, W.-L., Jokinen, J., Sparks, J. D., Yeh, S., & Chang, C. (2008). Increased hepatic steatosis and insulin resistance in mice lacking hepatic androgen receptor. *Hepatology*, 1924–1935. <https://doi.org/10.1002/hep.22252>
64. Liu, C.-C., Lee, Y.-C., Tsai, V. F. S., Cheng, K.-H., Wu, W.-J., Bao, B.-Y., Huang, C.-N., Yeh, H.-C., Tsai, C.-C., Wang, C.-J., & Huang, S.-P. (2015). The interaction of serum testosterone levels and androgen receptor CAG repeat polymorphism on the risk of erectile dysfunction in aging Taiwanese men. *Andrology*, 902–908. <https://doi.org/10.1111/andr.12068>
65. Lombardi, V., Bombaci, A., Zampedri, L., Lu, C.-H., Malik, B., Zetterberg, H., Heslegrave, A. J., Rinaldi, C., Greensmith, L., Hanna, M. G., Malaspina, A., & Fratta, P. (2019). Plasma pNfH levels differentiate SBMA from ALS. *Journal of Neurology, Neurosurgery & Psychiatry*, 215–217. <https://doi.org/10.1136/jnnp-2019-320624>
66. Lombardi, V., Querin, G., Ziff, O. J., Zampedri, L., Martinelli, I., Heller, C., Foiani, M., Bertolin, C., Lu, C.-H., Malik, B., Allen, K., Rinaldi, C., Zetterberg, H., Heslegrave, A., Greensmith, L., Hanna, M., Soraru, G., Malaspina, A., & Fratta, P. (2019). Muscle and not neuronal biomarkers correlate with severity in spinal and bulbar muscular atrophy. *Neurology*,

10.1212/WNL.0000000000007097. <https://doi.org/10.1212/wnl.0000000000007097>

67. Madeira, J. L. O., Souza, A. B. C., Cunha, F. S., Batista, R. L., Gomes, N. L., Rodrigues, A. S., Mennucci de Haidar Jorge, F., Chadi, G., Callegaro, D., Mendonca, B. B., Costa, E. M. F., & Domenice, S. (2017). A severe phenotype of Kennedy disease associated with a very large CAG repeat expansion. *Muscle & Nerve*, E95–E97. <https://doi.org/10.1002/mus.25952>
68. Malik, B., Devine, H., Patani, R., La Spada, A. R., Hanna, M. G., & Greensmith, L. (2019). Gene expression analysis reveals early dysregulation of disease pathways and links Chmp7 to pathogenesis of spinal and bulbar muscular atrophy. *Scientific Reports*. <https://doi.org/10.1038/s41598-019-40118-3>
69. Manganelli, F., Iodice, V., Provitera, V., Pisciotta, C., Nolano, M., Perretti, A., & Santoro, L. (2007). Small-fiber involvement in spinobulbar muscular atrophy (Kennedy's disease). *Muscle & Nerve*, 816–820. <https://doi.org/10.1002/mus.20872>
70. Manzano, R., Sorarú, G., Grunseich, C., Fratta, P., Zuccaro, E., Pennuto, M., & Rinaldi, C. (2018). Beyond motor neurons: expanding the clinical spectrum in Kennedy's disease. *Journal of Neurology, Neurosurgery & Psychiatry*, 808–812. <https://doi.org/10.1136/jnnp-2017-316961>
71. Mhatre, A. N., Trifiro, M. A., Kaufman, M., Kazemi-Esfarjani, P., Figlewicz, D., Rouleau, G., & Pinsky, L. (1993). Reduced transcriptional regulatory competence of the androgen receptor in X-linked spinal and bulbar muscular atrophy. *Nature Genetics*, 184–188. <https://doi.org/10.1038/ng1093-184>
72. Minamiyama, M. (2004). Sodium butyrate ameliorates phenotypic expression in a transgenic mouse model of spinal and bulbar muscular atrophy. *Human Molecular Genetics*, 1183–1192. <https://doi.org/10.1093/hmg/ddh131>
73. Narayanan, R., Coss, C. C., & Dalton, J. T. (2018). Development of selective androgen receptor modulators (SARMs). *Molecular and Cellular Endocrinology*, 134–142. <https://doi.org/10.1016/j.mce.2017.06.013>
74. Nath, S. R., Yu, Z., Gipson, T. A., Marsh, G. B., Yoshidome, E., Robins, D. M., Todi, S. V., Housman, D. E., & Lieberman, A. P. (2018). Androgen receptor polyglutamine expansion drives age-dependent quality control defects and muscle dysfunction. *Journal of Clinical Investigation*, 3630–3641. <https://doi.org/10.1172/jci99042>

75. Nedelsky, N. B., Pennuto, M., Smith, R. B., Palazzolo, I., Moore, J., Nie, Z., Neale, G., & Taylor, J. P. (2010). Native Functions of the Androgen Receptor Are Essential to Pathogenesis in a *Drosophila* Model of Spinobulbar Muscular Atrophy. *Neuron*, 936–952.
<https://doi.org/10.1016/j.neuron.2010.08.034>
76. Nenonen, H. A., Giwercman, A., Hallengren, E., & Giwercman, Y. L. (2010). Non-linear association between androgen receptor CAG repeat length and risk of male subfertility - a meta-analysis. *International Journal of Andrology*, 327–332.
<https://doi.org/10.1111/j.1365-2605.2010.01084.x>
77. Nenonen, H., Bjork, C., Skjaerpe, P.-A., Giwercman, A., Rylander, L., Svartberg, J., & Giwercman, Y. L. (2009). CAG repeat number is not inversely associated with androgen receptor activity in vitro. *Molecular Human Reproduction*, 153–157.
<https://doi.org/10.1093/molehr/gap097>
78. Neuschmid-Kaspar, F., Gast, A., Peterziel, H., Schneikert, J., Muigg, A., Ransmayr, G., Klocker, H., Bartsch, G., & Cato, A. C. B. (1996). CAG-repeat expansion in androgen receptor in Kennedy's disease is not a loss of function mutation. *Molecular and Cellular Endocrinology*, 149–156. [https://doi.org/10.1016/0303-7207\(95\)03741-1](https://doi.org/10.1016/0303-7207(95)03741-1)
79. Ni, W., Chen, S., Qiao, K., Wang, N., & Wu, Z.-Y. (2015). Genotype-Phenotype Correlation in Chinese Patients with Spinal and Bulbar Muscular Atrophy. *PLOS ONE*, e0122279.
<https://doi.org/10.1371/journal.pone.0122279>
80. Nordenvall, A. S., Paucar, M., Almqvist, C., Nordenström, A., Frisé, L., & Nordenskjöld, A. (2016). Hypospadias as a novel feature in spinal bulbar muscle atrophy. *Journal of Neurology*, 703–706. <https://doi.org/10.1007/s00415-016-8038-y>
81. Owens, S. J., Weickert, T. W., Purves-Tyson, T. D., Ji, E., White, C., Galletly, C., Liu, D., O'Donnell, M., & Shannon Weickert, C. (2018). Sex-Specific Associations of Androgen Receptor CAG Trinucleotide Repeat Length and of Raloxifene Treatment with Testosterone Levels and Perceived Stress in Schizophrenia. *Molecular Neuropsychiatry*, 28–41.
<https://doi.org/10.1159/000495062>
82. Palazzolo, I., Stack, C., Kong, L., Musaro, A., Adachi, H., Katsuno, M., Sobue, G., Taylor, J. P., Sumner, C. J., Fischbeck, K. H., & Pennuto, M. (2009). Overexpression of IGF-1 in Muscle Attenuates Disease in a Mouse Model of Spinal and Bulbar Muscular Atrophy. *Neuron*, 316–328.
<https://doi.org/10.1016/j.neuron.2009.07.019>

83. Paskova, L., Smesny Trtkova, K., Fialova, B., Benedikova, A., Langova, K., & Kolar, Z. (2013). Different effect of sodium butyrate on cancer and normal prostate cells. *Toxicology in Vitro*, 1489–1495. <https://doi.org/10.1016/j.tiv.2013.03.002>
84. Patrizio, P., Leonard, D., Chen, K., Hernandez-Ayup, S., & Trounson, A. (2001). Larger trinucleotide repeat size in the androgen receptor gene of infertile men with extremely severe oligozoospermia. *Journal of Andrology*, 22(3), 444–448. <https://www.ncbi.nlm.nih.gov/pubmed/11330644>
85. Pennuto, M., & Rinaldi, C. (2018). From gene to therapy in spinal and bulbar muscular atrophy: Are we there yet? *Molecular and Cellular Endocrinology*, 113–121. <https://doi.org/10.1016/j.mce.2017.07.005>
86. Pieper, C. C., Konrad, C., Sommer, J., Teismann, I., & Schiffbauer, H. (2012). Structural changes of central white matter tracts in Kennedy's disease - a diffusion tensor imaging and voxel-based morphometry study. *Acta Neurologica Scandinavica*, 323–328. <https://doi.org/10.1111/ane.12018>
87. Polo, A., Teatini, F., D'Anna, S., Manganotti, P., Salviati, A., Dallapiccola, B., Zanette, G., & Rizzuto, N. (1996). Sensory involvement in X-linked spino-bulbar muscular atrophy (Kennedy's syndrome): An electrophysiological study. *Journal of Neurology*, 388–392. <https://doi.org/10.1007/bf00868997>
88. Ponnusamy, S., Coss, C. C., Thiyagarajan, T., Watts, K., Hwang, D.-J., He, Y., Selth, L. A., McEwan, I. J., Duke, C. B., Pagadala, J., Singh, G., Wake, R. W., Ledbetter, C., Tilley, W. D., Moldoveanu, T., Dalton, J. T., Miller, D. D., & Narayanan, R. (2017). Novel Selective Agents for the Degradation of Androgen Receptor Variants to Treat Castration-Resistant Prostate Cancer. *Cancer Research*, 6282–6298. <https://doi.org/10.1158/0008-5472.can-17-0976>
89. Qiang, Q., Adachi, H., Huang, Z., Jiang, Y.-M., Katsuno, M., Minamiyama, M., Doi, H., Matsumoto, S., Kondo, N., Miyazaki, Y., Iida, M., Tohnai, G., & Sobue, G. (2013). Genistein, a natural product derived from soybeans, ameliorates polyglutamine-mediated motor neuron disease. *Journal of Neurochemistry*, 122–130. <https://doi.org/10.1111/jnc.12172>
90. Querin, G., Sorarù, G., & Pradat, P.-F. (2017). Kennedy disease (X-linked recessive bulbospinal neuronopathy): A comprehensive review from pathophysiology to therapy. *Revue Neurologique*, 326–337. <https://doi.org/10.1016/j.neurol.2017.03.019>

91. Querin, Giorgia, Bede, P., Marchand-Pauvert, V., & Pradat, P.-F. (2018). Biomarkers of Spinal and Bulbar Muscle Atrophy (SBMA): A Comprehensive Review. *Frontiers in Neurology*. <https://doi.org/10.3389/fneur.2018.00844>
92. Querin, Giorgia, Bertolin, C., Da Re, E., Volpe, M., Zara, G., Pegoraro, E., Caretta, N., Foresta, C., Silvano, M., Corrado, D., Iafrate, M., Angelini, L., Sartori, L., Pennuto, M., Gaiani, A., Bello, L., Semplicini, C., Pareyson, D., Silani, V., ... Sorarù, G. (2015). Non-neural phenotype of spinal and bulbar muscular atrophy: results from a large cohort of Italian patients. *Journal of Neurology, Neurosurgery & Psychiatry*, 810–816. <https://doi.org/10.1136/jnnp-2015-311305>
93. Quigley, C., Friedman, K., Johnson, A., Lafreniere, R., Silverman, L., Lubahn, D., Brown, T., Wilson, E., Willard, H., & French, F. (1992). Complete deletion of the androgen receptor gene: definition of the null phenotype of the androgen insensitivity syndrome and determination of carrier status. *The Journal of Clinical Endocrinology and Metabolism*, 74(4), 927–933. <https://doi.org/10.1210/jcem.74.4.1347772>
94. Renier, K. J., Troxell-Smith, S. M., Johansen, J. A., Katsuno, M., Adachi, H., Sobue, G., Chua, J. P., Sun Kim, H., Lieberman, A. P., Breedlove, S. M., & Jordan, C. L. (2014). Antiandrogen Flutamide Protects Male Mice From Androgen-Dependent Toxicity in Three Models of Spinal Bulbar Muscular Atrophy. *Endocrinology*, 2624–2634. <https://doi.org/10.1210/en.2013-1756>
95. Rhodes, L. E., Freeman, B. K., Auh, S., Kokkinis, A. D., La Pean, A., Chen, C., Lehky, T. J., Shrader, J. A., Levy, E. W., Harris-Love, M., Di Prospero, N. A., & Fischbeck, K. H. (2009). Clinical features of spinal and bulbar muscular atrophy. *Brain*, 3242–3251. <https://doi.org/10.1093/brain/awp258>
96. Rinaldi, C., Bott, L. C., Chen, K., Harmison, G. G., Katsuno, M., Sobue, G., Pennuto, M., & Fischbeck, K. H. (2012). Insulinlike Growth Factor (IGF)-1 Administration Ameliorates Disease Manifestations in a Mouse Model of Spinal and Bulbar Muscular Atrophy. *Molecular Medicine*, 1261–1268. <https://doi.org/10.2119/molmed.2012.00271>
97. Rinaldi, C., Malik, B., & Greensmith, L. (2015). Targeted Molecular Therapies for SBMA. *Journal of Molecular Neuroscience*, 335–342. <https://doi.org/10.1007/s12031-015-0676-5>
98. Romigi, A., Liguori, C., Placidi, F., Albanese, M., Izzi, F., Uasone, E., Terracciano, C., Marciani, M. G., Mercuri, N. B., Ludovisi, R., & Massa, R. (2014). Sleep disorders in spinal and bulbar muscular atrophy (Kennedy's disease): a controlled polysomnographic and self-reported questionnaires study. *Journal of Neurology*, 889–893. <https://doi.org/10.1007/s00415-014-7293-z>

99. Rosenbohm, A., Hirsch, S., Volk, A. E., Grehl, T., Grosskreutz, J., Hanisch, F., Herrmann, A., Kollwe, K., Kress, W., Meyer, T., Petri, S., Prudlo, J., Wessig, C., Müller, H.-P., Dreyhaupt, J., Weishaupt, J., Kubisch, C., Kassubek, J., Weydt, P., & Ludolph, A. C. (2018). The metabolic and endocrine characteristics in spinal and bulbar muscular atrophy. *Journal of Neurology*, 1026–1036. <https://doi.org/10.1007/s00415-018-8790-2>
100. Sacheck, J. M., Hyatt, J. K., Raffaello, A., Thomas Jagoe, R., Roy, R. R., Reggie Edgerton, V., Lecker, S. H., & Goldberg, A. L. (2006). Rapid disuse and denervation atrophy involve transcriptional changes similar to those of muscle wasting during systemic diseases. *The FASEB Journal*, 140–155. <https://doi.org/10.1096/fj.06-6604com>
101. Santi, D., Spaggiari, G., Gilioli, L., Potì, F., Simoni, M., & Casarini, L. (2018). Molecular basis of androgen action on human sexual desire. *Molecular and Cellular Endocrinology*, 31–41. <https://doi.org/10.1016/j.mce.2017.09.007>
102. Schmidt, B. J., Greenberg, C. R., Allingham-Hawkins, D. J., & Spriggs, E. L. (2002). Expression of X-linked bulbospinal muscular atrophy (Kennedy disease) in two homozygous women. *Neurology*, 770–772. <https://doi.org/10.1212/wnl.59.5.770>
103. Singh, R., Singh, L., & Thangaraj, K. (2007). Phenotypic heterogeneity of mutations in androgen receptor gene. *Asian Journal of Andrology*, 147–179. <https://doi.org/10.1111/j.1745-7262.2007.00250.x>
104. Skjaerpe, P. A., Giwercman, Y. L., Giwercman, A., & Svartberg, J. (2008). Androgen receptor gene polymorphism and the metabolic syndrome in 60-80 years old Norwegian men. *International Journal of Andrology*, 500–506. <https://doi.org/10.1111/j.1365-2605.2008.00942.x>
105. Sorarù, G., D'Ascenzo, C., Polo, A., Palmieri, A., Baggio, L., Vergani, L., Gellera, C., Moretto, G., Pegoraro, E., & Angelini, C. (2008). Spinal and bulbar muscular atrophy: Skeletal muscle pathology in male patients and heterozygous females. *Journal of the Neurological Sciences*, 100–105. <https://doi.org/10.1016/j.jns.2007.08.012>
106. Soukup, G. R., Sperfeld, A.-D., Uttner, I., Karitzky, J., Ludolph, A. C., Kassubek, J., & Schreiber, H. (2009). Frontotemporal cognitive function in X-linked spinal and bulbar muscular atrophy (SBMA): a controlled neuropsychological study of 20 patients. *Journal of Neurology*, 1869–1875. <https://doi.org/10.1007/s00415-009-5212-5>

107. Spada, A. R. L., Wilson, E. M., Lubahn, D. B., Harding, A. E., & Fischbeck, K. H. (1991). Androgen receptor gene mutations in X-linked spinal and bulbar muscular atrophy. *Nature*, 77–79. <https://doi.org/10.1038/352077a0>
108. Sperfeld, A. D., Karitzky, J., Brummer, D., Schreiber, H., Häussler, J., Ludolph, A. C., & Hanemann, C. O. (2002). X-linked Bulbospinal Neuronopathy. *Archives of Neurology*, 1921. <https://doi.org/10.1001/archneur.59.12.1921>
109. Stanworth, R. D., Kapoor, D., Channer, K. S., & Jones, T. H. (2008). Androgen receptor CAG repeat polymorphism is associated with serum testosterone levels, obesity and serum leptin in men with type 2 diabetes. *European Journal of Endocrinology*, 739–746. <https://doi.org/10.1530/eje-08-0266>
110. Stenoiien, D. L., Cummings, C. J., Adams, H. P., Mancini, M. G., Patel, K., DeMartino, G. N., Marcelli, M., Weigel, N. L., & Mancini, M. A. (1999). Polyglutamine-Expanded Androgen Receptors Form Aggregates That Sequester Heat Shock Proteins, Proteasome Components and SRC-1, and Are Suppressed by the HDJ-2 Chaperone. *Human Molecular Genetics*, 731–741. <https://doi.org/10.1093/hmg/8.5.731>
111. Takeyama, K., Ito, S., Yamamoto, A., Tanimoto, H., Furutani, T., Kanuka, H., Miura, M., Tabata, T., & Kato, S. (2002). Androgen-Dependent Neurodegeneration by Polyglutamine-Expanded Human Androgen Receptor in *Drosophila*. *Neuron*, 855–864. [https://doi.org/10.1016/s0896-6273\(02\)00875-9](https://doi.org/10.1016/s0896-6273(02)00875-9)
112. Tirabassi, G., Corona, G., Falzetti, S., delli Muti, N., Maggi, M., & Balercia, G. (2016). Influence of Androgen Receptor Gene CAG and GGC Polymorphisms on Male Sexual Function: A Cross-Sectional Study. *International Journal of Endocrinology*, 1–7. <https://doi.org/10.1155/2016/5083569>
113. Traish, A. M. (2008). Androgens Play a Pivotal Role in Maintaining Penile Tissue Architecture and Erection: A Review. *Journal of Andrology*, 363–369. <https://doi.org/10.2164/jandrol.108.006007>
114. Udd, B., Juvonen, V., Hakamies, L., Nieminen, A., Wallgren-Pettersson, C., Cederquist, K., & Savontaus, M.-L. (2009). High prevalence of Kennedy's disease in Western Finland - is the syndrome underdiagnosed? *Acta Neurologica Scandinavica*, 128–133. <https://doi.org/10.1111/j.1600-0404.1998.tb01732.x>

115. Unrath, A., Müller, H.-P., Riecker, A., Ludolph, A. C., Sperfeld, A.-D., & Kassubek, J. (2010). Whole brain-based analysis of regional white matter tract alterations in rare motor neuron diseases by diffusion tensor imaging. *Human Brain Mapping*, NA-NA. <https://doi.org/10.1002/hbm.20971>
116. Walcott, J. L., & Merry, D. E. (2002). Ligand Promotes Intranuclear Inclusions in a Novel Cell Model of Spinal and Bulbar Muscular Atrophy. *Journal of Biological Chemistry*, 50855–50859. <https://doi.org/10.1074/jbc.m209466200>
117. Waza, M., Adachi, H., Katsuno, M., Minamiyama, M., Sang, C., Tanaka, F., Inukai, A., Doyu, M., & Sobue, G. (2005). 17-AAG, an Hsp90 inhibitor, ameliorates polyglutamine-mediated motor neuron degeneration. *Nature Medicine*, 1088–1095. <https://doi.org/10.1038/nm1298>
118. Yang, Z., Chang, Y.-J., Yu, I.-C., Yeh, S., Wu, C.-C., Miyamoto, H., Merry, D. E., Sobue, G., Chen, L.-M., Chang, S.-S., & Chang, C. (2007). ASC-J9 ameliorates spinal and bulbar muscular atrophy phenotype via degradation of androgen receptor. *Nature Medicine*, 348–353. <https://doi.org/10.1038/nm1547>
119. Yu, Z. (2006). Androgen-dependent pathology demonstrates myopathic contribution to the Kennedy disease phenotype in a mouse knock-in model. *Journal of Clinical Investigation*, 2663–2672. <https://doi.org/10.1172/jci28773>
120. Zitzmann, M. (2008). The Role of the CAG Repeat Androgen Receptor Polymorphism in Andrology. In *Frontiers of Hormone Research* (pp. 52–61). KARGER. <https://doi.org/10.1159/000175843>
121. Zitzmann, M. (2009). Pharmacogenetics of testosterone replacement therapy. *Pharmacogenomics*, 1341–1349. <https://doi.org/10.2217/pgs.09.58>
-

AR deregulation as a key pathological driver of PFS?

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/ar-deregulation-as-a-key-pathological-driver-of-pfs/>

Induced Wild Type AR Overexpression and the potential relevance of SBMA to PFS

As discussed, as the CAG trinucleotide sequence extends in the N-terminal domain of the AR, there is a consequent functional decline in transcriptional efficiency which is seemingly associated with a compensatory increase in androgen levels. However, at longer repeat lengths, high androgen levels can exert a deleterious and ultimately toxic response. Crucially, polyglutamine expansion is not the only way ligand-dependent toxicity can be conferred to the AR protein, and overexpression of the wild type AR can cause a paradoxical loss of function and toxic gain of function. This is reflective of evidence in other polyglutamine diseases that point to gain of native protein function underlying pathology (Paulson et al., 2017). It is now appreciated that balanced gene expression is vital for homeostasis, and overexpression of wild-type proteins causes disease states in humans (Ohshima et al., 2017; Shastry, 1995). Multiple studies demonstrate that, while seemingly paradoxical, sufficient increases in AR expression converge with loss of function phenotypes, with an inverse U-shaped curve representative of AR gene dose response in tissues. The pathological consequence of overexpression of the AR is therefore coherent with Prelich's observation that overexpression of proteins mimics a loss of function and interferes with its function antimorphically. The mechanisms by which overexpression causes a mutant phenotype is therefore of great importance to further understand (Prelich, 2012).

Generating mice overexpressing AR solely in skeletal muscle, Monks et al. reported the striking and seemingly counter-intuitive observation that overexpression of the Wild-Type androgen receptor recapitulates the pathological consequence of polyglutamine expansion despite a polyglutamine repeat tract comprised of 22 glutamines. Decreased viability was observed in males of all seven transgene lines but not in females. Interestingly, administration of flutamide to pregnant dams enhanced perinatal survival, suggesting prenatal androgen activation of the overexpressed AR, not the overexpression *per se*, is causative of death. Two transgenic mouse lines of differing WT AR copy number (L78 < L141) were characterised. L141 males exhibited a far more severe phenotype, corresponding to a significantly higher AR expression at the mRNA and protein level. Surviving L78 males were functionally comparable to wild type despite a lower body weight. However, L141 males exhibited a marked phenotype of lower body weight, curvature of the thoracic spine, severe deficits in motor function and muscle strength, and early death. Castration dramatically restored function in L141 mice, illustrating the androgen dependency of the toxicity. Remarkably, although L141 females were apparently unaffected by AR overexpression *per se*, when administered testosterone to the approximate circulating level of male mice, they rapidly developed a comparable disease phenotype to male L141 mice including motor dysfunction and muscle

atrophy. Over 9 days of T treatment was fatal to female L141 mice. L78 female mice did not become symptomatic or atrophic with T treatment, even for prolonged periods. This parallels the asymptomatic L78 male. This is strongly indicative that the degree of overexpression dictates severity of androgen-mediated toxicity and, as Monks et al. observe in several contexts, that overexpressed AR confers toxicity once activated by hormonal ligand (Monks et al., 2007)?.

Monks et al. compared differentially regulated genes in myogenic transgene mice and the SBMA AR97 and AR113Q models. Gene expression in the transgene AR-overexpressing muscle revealed similar deregulation to AR Knock-out muscle, further suggesting that a paradoxical loss of AR function results from overexpression of the androgen receptor. The finding of overexpression of WT AR reproducing a phenotype comparable to polyglutamine expansion was noted to be surprising and puzzling considering SBMA is associated with a loss of AR function whereas overexpression of the AR would typically be expected to enhance the function of androgen signaling (Mo et al., 2010)?. Further striking findings were provided through investigation of the contributions of native AR interactions to polyglutamine-expanded AR toxicity in *Drosophila* models. Nedelsky et al. determined that native interactions at AF-1 of the AR modify toxicity while AF-2 coregulator interaction and function is essential for toxicity. Expressing AR in the photoreceptor neurons and accessory pigment cells of the eyes of the developing flies, they demonstrated a polyglutamine length and ligand-dependent degenerative phenotype. While flies reared on normal food did not demonstrate pathology, a degenerative phenotype in the posterior margin of the eye occurred in flies reared on food containing DHT. This androgen and polyQ length dependent degenerative phenotype of atrophy and functional deficit was further demonstrated in larval tissues including salivary glands and motor neurons. Crucially, Nedelsky et al. reported that wild-type AR of a 12Q polyglutamine-length, when expressed at very high levels, resulted in a degenerative phenotype indistinguishable from that caused by expansion of the AR polyglutamine tract (Nedelsky et al., 2010)?. This reflected the dose dependency and pathological consequence of wild-type AR overexpression well reported by Monks et al. Furthermore, though generally weaker, expression analysis revealed a similar dysregulation in both AR12Q+DHT and AR52Q+DHT flies, lending further support to a link between an amplification of native function and the toxicity induced by polyglutamine expansion and is supportive of a conserved mechanism. Interestingly, quantitative analysis did not reveal a correlation between the amount of high molecular weight species and neurodegeneration in their *Drosophila* model. This is in line with the lack of AR positive aggregates reported in transgenic mice that recapitulated the SBMA phenotype (Monks et al., 2007)?. The presence of aggregates was in previous decades presumed to be a driving factor in pathogenesis of SBMA, however this is no longer the case and a direct mechanistic involvement is controversial (Todd & Lim, 2013)?. Providing another parallel between the effect of polyglutamine expansion and wild type overexpression, Halievski et al. demonstrated that in mice expressing a human androgen receptor of 97 CAGs and the wild-type overexpressing myogenic mice, several common transcriptional effects were seen, such as robust downregulation of BDNF and NT-4 transcripts. Remarkably, similar effects were seen indistinctly across both synaptic and extrasynaptic domains, suggesting a broad effect and involvement of common deleterious AR-mediated mechanisms across cell types (Halievski et al., 2019)?.

While it might be expected AR overexpression would result in a hyper-masculine socio-sexual

phenotype, Swift-Gallant et al. demonstrated significant reductions of male-typical aggressive and sexual behaviours in transgene AR overexpressing mice. This non-linear androgen response was curiously reflective of loss of AR function. Interestingly, same-sex anogenital investigation was increased and male-typical preferences for female olfactory cues were disrupted in globally overexpressing mice but not mice only overexpressing AR in neural tissue, suggesting a direct role of non-neuronal AR in mediation of socio-sexual behaviours. A decrease in testosterone production is not a sufficient explanation for the mechanistic consequences of overexpression on masculine physiological and behavioural phenotypes and the many convergences with loss of function models, and reduced testosterone was not routinely observed in models of overexpression (Swift-Gallant et al., 2016). Monks and Swift-Gallant considered a uniform global loss of AR function unlikely, proposing a cellular mechanism that would be differentiated according to affected neurological or physiological tissue and system. This would implicate regional variations, possibly including site-specific cofactor influences and differential transcriptional effects resulting from regional epigenetic changes. Additionally, overexpression of AR has been suggested as a plausible mechanistic route to alteration in neurosteroid synthesis (Monks & Swift-Gallant, 2018).

Considerable evidence exists to support an overlapping androgen dependent toxicity in the contexts of AR polyglutamine tract expansion and overexpression of the wild-type AR, and a loss of function coincident to both insufficient and excessive AR signaling. It is therefore highly significant that both hypogonadism (Seftel, 2005) and the multi-systemic symptom profile of SBMA (Querin et al., 2017) bear a clear resemblance to the broad symptomatology of PFS. However, it is important to consider that there are notable areas of presentation and progression in which the disease states of PFS and SBMA differ. Neurocognitive symptoms are profoundly more severe in PFS than are reported in SBMA, although these domains of disease involvement are not without overlap as we have illustrated. Tongue atrophy is not reported in PFS. These differences are likely inherent to the aetiologies of the respective diseases: An endocrine disruption leading to epigenetic dysregulation in PFS and a genetic glutamine repeat sequence as causative factor in SBMA. While SBMA is a characteristically slow progressing condition, PFS can, in many cases, onset extremely rapidly with the discussed "crash". After this onset, an initial period of weeks or months during which the pathology is often rapidly progressive to what patients refer to as a "baseline" state occurs. Atrophy of androgen dependent tissue and physiological changes are often reported over this time. Beyond this, PFS is not always markedly progressive, with some patients experiencing improvement or stabilisation of their symptoms to variable points over subsequent months or years. As we will discuss, exogenous testosterone can sometimes cause symptomatic intensification, and significant and rapid phenotypical deterioration with additional symptomatic physiological domains can occur following exposure to further antiandrogenic endocrine disrupting substances. Previously discussed as the "crash", the majority patient experience of a intensification or development of symptoms after cessation of the drug may reflect the return of 5 α -dihydrotestosterone to physiological levels in the presence of the newly uninhibited 5-alpha reductase enzymes. In the myogenic models discussed, when male physiological levels of androgens were administered to female L141 mice a severe disease state is rapidly induced, while the L78 mice were largely asymptomatic. A site-specificity and expression level-dependency of induced AR overexpression therefore serves as a compelling explanation for the large variation in the toxic post-drug phenotype, manifesting as either a continuation of on-drug side effects or, more commonly, the crash, which can vary from an onset of sexual dysfunction, libido loss, anxiety and depression to a devastating and disabling physiological and psychological alteration including cognitive impairment of executive function, derealisation, anhedonia, panic attacks, memory loss, total insomnia,

dysautonomia, atrophy of androgen-responsive tissue and metabolic changes.

A "malignancy switch" for the androgen receptor?

In the absence of serum endocrine or other toxicological findings that could account for the pathological features of PFS (Irwig, 2014; Melcangi et al., 2017)? we suggest a biological event during use of finasteride is causing an often permanent change in the ordinary metabolic function of cells through epigenetic alteration. Although this is controversial to suggest, the potential severity of the disease cannot be overstated, and in a significant number of cases the health problems are severe, progressive, do not resolve with time and entail a peculiar endocrine fragility. We hypothesise underlying pre-existing genetic and/or epigenetic factors differentiate those who are prone to developing PFS, and this predisposition effects deleterious epigenetic modifications by means of a conserved mechanism upon significant reduction of intracellular androgen-dependent transactivation through various modes of action including but not limited to 5alpha reductase inhibition. These vectors include downregulation of AR mRNA, an induced increase of protein degradation, upregulation of enzymes capable of reducing endogenous AR ligand to inactive androgen metabolites and suppression of steroidogenic enzymes. We further suggest the necessary exposure and severity of symptomatic outcomes are dependent on interindividual differences within this/these underlying predisposing factor(s) and the resulting degree of persistent dysregulation of the androgen receptor on a site-specific basis.

The epigenetically determined fate of somatic cells is not terminal. Epigenetic barriers preservative of cellular integrity were famously visualised by Conrad Waddington's epigenetic landscape, which described a ball running down valleys in determination of its ultimate differentiated state (Slack, 2002)?. However, these can be overcome given the correct stimuli, and the past decades have seen rapid advancements in cellular reprogramming methods (MacArthur et al., 2009)?. As chemicals are capable of inducing reversal of cell lineage, Kanherkar et al. investigated the possibility of permanent epigenetic alterations occurring following exposure to pharmacological agents. HEK-293 cells cultured in the SSRI antidepressant citalopram revealed significant differential methylation in hundreds of genes. They proposed the term "pharmaceutical reprogramming" to describe a partial dysdifferentiation event resulting from drug-induced methylation changes that consequently alter cellular function and integrity (Kanherkar et al., 2018)?. Evidence demonstrates adult sex typical behaviour can be altered in mammals under certain conditions and may be a function of epigenetic maintenance and gene expression with behavioural impacts (McCarthy, 2019)?. In relation to androgen signalling, significant recent work has suggested that biologically meaningful differences that directly influence behaviour and function pertaining to sexual traits can arise from epigenetic alteration to the program of the androgen receptor (Schuppe et al., 2020)?.

As well as fibrotic changes in the penis, Enatsu et al. reported a reduction of AR and an increase in ER expression in the prostate of young rats administered dutasteride, speculating that an improper response to androgens upon restoration could underlie the sexual dysfunction in PFS owing to altered local receptor expression (Enatsu et al., 2016). This study demonstrated a deleterious influence exerted by 5 α reductase inhibition in young rats that entailed morphological alterations to sexual organs and epigenetic remodelling that trended towards the effect of castration. However, many factors exclude the typical response to prolonged 5 α reductase inhibition from being an applicable model for the behaviour of PFS. These include the rarity of PFS amongst 5 α users, the clinical picture of PFS including pathological development and/or progression of the disease following cessation, a prevalence in younger men using a lower dose, the brevity of exposure in some of the most severely affected cases, the commonly reported responses of PFS patients to trialled therapies, and the previously reported determination of persistent and significant upregulation of the AR in prepuce tissue of PFS patients. Nevertheless, the parabolic nature of AR expression would suggest Enatsu's hypothesis of an induced dysfunction in local androgen response owing to epigenetic remodelling is plausible. Finasteride has previously been shown to upregulate prostate epithelial AR significantly in BPH patients after 30 days of exposure (Hsieh et al., 2011). Corradi et al. demonstrated that Finasteride induced a persisting overexpression of the AR and important alterations in the tissue microenvironment of the prostate gland in young gerbils. Across three stages of postnatal development, the content and intensity of AR immunostaining were noticeably elevated, particularly in epithelial cell nuclei. Both the tissue changes and AR overexpression proved persistent. Interestingly, when contrasted with their respective control groups, a greater increase in AR nuclear intensity could be observed in the young (8% to 61.5%) Finasteride administered experimental group as opposed to the old Finasteride administered experimental group (66% to 72.5%) at the conclusion of the post-treatment phase (Corradi et al., 2009).

Coskuner et al, reviewing literature on persistent sexual symptoms in a subset of 5 α reductase inhibitor users, considered tissue-specific epigenetic effects likely given the persistence of symptoms (Coskuner et al., 2019). In considering the mechanistic origins of the development of PFS following endocrine disruption with Finasteride, Traish proposed that androgen deprivation and depletion of the substrate precursors for the 3 β -hydroxy-steroid dehydrogenases causative of a block in neurosteroidogenesis, attenuating the function of steroid and neurotransmitter receptors and inducing changes in the expression of a host of gene products, eliciting epigenetic changes manifested in histone acetylation, DNA methylation and upregulation of the AR. Traish thus suggests these changes, together with the consequent depletion of neurosteroids, manifest in the development of PFS in susceptible individuals (Traish, 2018). Di Loreto et al had previously suggested that it was tempting to speculate that PFS patients have triggered processes associated with advanced age by pharmaceutical androgen deprivation (Di Loreto et al., 2014). The natural decline of testosterone values with ageing has been well established (Kaufman & Vermeulen, 2005). PFS may thus represent an aberration of such processes, resulting as an adaptive epigenetic response to the pre-receptor disruption of androgen signaling during finasteride use.

Our stated hypothesis for PFS as an epigenetic adaptation induced by pharmaceutically interrupted androgen signalling accounts for a deregulated epigenome and the onset and/or symptomatic

intensification following finasteride withdrawal, often after a brief resolution of symptoms, which standardised questionnaires including our own data indicate is an intrinsic feature of the syndrome (Propeciahelp Post-Drug Syndrome Survey: Data not provided). Cessation of finasteride will result in a surge in androgen production owing to the newly uninhibited 5 α -reductase enzyme. Presuming a 60% reduction of basal DHT levels during finasteride use, cells epigenetically adapted to a depletion of androgenic signaling owing to the pharmacological reduction of DHT would be exposed to a 300% increase in DHT upon cessation. As molecular level investigation has revealed a persistent elevation in expression of the androgen receptor in symptomatic tissue of a PFS cohort, this may entail a deleterious ligand-dependent effect in alignment with the demonstrated *in vitro* and *in vivo* models discussed. Application of such a conceptual framework to the pathology of PFS is not unprecedented. Professor Charles Ryan explained the tissue response to testosterone in terms of a "bell curve" in his book *The Virility Paradox*. He wrote of PFS: "I think this is what we are seeing here. With a greater concentration of receptors, the organ becomes more sensitive to testosterone and at a certain point, paradoxically, that sensitivity may shut down" (Ryan, 2018)?.

We hypothesise that a loss of function and toxic gain of function manifests tissue specifically in a broad spectrum of clinical endpoints, from functional impairment to atrophy in affected tissues. In consideration of this, we would expect future gene expression analysis of symptomatic tissue in severely affected patients to reveal widespread dysregulation of gene expression. A consideration of how a dysregulation of the AR and associated epigenetic remodelling might occur as an aberrant result of antiandrogenic endocrine disruption, and how it may influence broader gene expression, is therefore necessary. This can be contextualised via known molecular mechanisms.

The most well recognised epigenetic adaptations occurring as a result of androgen deprivation therapy is in the context of castration resistant prostate cancer. As a driver of epithelial cell growth and proliferation as well as a fundamental aspect of prostate cancer progression, the androgen receptor axis has been the predominant therapeutic target in prostate cancer for over 75 years (Kim & Ryan, 2012; Takeda et al., 2018)?. Patients develop resistance to androgen deprivation therapy after a period of this first line treatment, a state with very poor prognosis known as castration resistant prostate cancer. Second generation antiandrogen treatments have been developed, however nearly all men also develop resistance to this, suggestive of a mechanistic response irrespective of the agent (Robinson et al., 2015)?. Although not always observed, amplification of the AR is the most common mechanism of castration resistance (Takeda et al., 2018)? and is the only consistent gene expression change associated with hormone refractory disease (Chen et al., 2003)?. The amplification of the AR occurs during androgen deprivation therapy (Friedlander et al., 2011; Visakorpi et al., 1995)? or antiandrogen treatment (Coutinho et al., 2016)? and represents an adaptive response to the low androgen environment (Perner et al., 2015; Ruggero et al., 2018; Teply et al., 2018)? that sensitizes cells to lower levels of hormone (Waltering et al., 2009)?. Interestingly, low, rather than high, endogenous testosterone levels have been associated with poor prognostic features in prostate cancer and disease reclassification during active surveillance (Amadi et al., 2018; San Francisco et al., 2014)?. Several lines of evidence suggest low levels of androgen may predispose to more aggressive tumours (Swerdlhoff et al., 2017)?. Gravina et al. provided evidence that epigenetic mechanisms can contribute to castration resistant phenotypes, demonstrating that

pca cell models in androgen-deprived medium or bicalutamide progressively increased DNMT expression, which increased in proportion to AR upregulation. These findings were verified in patient tissue. DNMT was additionally shown to be regulated by AR, as siRNA AR interference greatly reduced DNMT modulation (Gravina et al., 2011).

Chen et al. hypothesised that any one of a number of primary molecular events that alter AR activity and increase AR mRNA could represent a common final pathway for castration resistance in PCa. In support of this, it was demonstrated that LNCaP cells altered to express a threefold greater level of AR grew in low androgen concentrations whereas LNCaP cells did not unless supplemented with androgen, confirming that AR overexpression alone confers castration resistance. In addition, they demonstrated that the androgen receptor must bind its ligand to confer hormone-refractory growth. LBD mutant LNCaP constructs did not exhibit hormone-refractory growth beyond vector controls even at ten-fold increases of AR expression levels. Interestingly, AR antagonists, in the circumstance of overexpression, induced certain androgen regulated genes (Chen et al., 2003). This paradoxical response is reflected in the apparent vulnerability CRPC cells exhibit to supraphysiological androgens. Teply et al. demonstrated clinical response and short-lived resensitisation to enzalutamide through bipolar androgen therapy using exogenous testosterone (Teply et al., 2018). Similarly, Christensen et al. reported a remarkable clinical and prostate-specific antigen response to a combination of high doses of testosterone and radium 223 in a patient with metastatic CRPC whose disease had progressed while receiving a number of antiandrogenic therapies (Christensen et al., 2019). ctDNA consistently showed a high degree of AR amplification. These findings suggest that the switch to a hormone refractory state entails a markedly different response to ligand.

Large scale sequencing studies have shown over 90% of cases of advanced CRPC exhibit overexpressed or altered AR, alongside significant alteration to genes involved in histone rearrangement and chromatin modification (Barbieri et al., 2012; Braadland & Urbanucci, 2019; Grasso et al., 2012; Robinson et al., 2015; Taylor et al., 2010). Chromatin structure is at least partially definitive of a cell's transcriptional program, and determines vast networks of regulatory elements tissue-specifically (Pihlajamaa et al., 2015). Chromatin relaxation is part of an adaptive response that increases the probability of genomic access and transcription, and enables continued function in a situation in which sufficient androgens and androgen signaling are therapeutically reduced (Braadland & Urbanucci, 2019). Patterns of open chromatin differ in CRPC to BPH or PCa samples, with large interindividual variance in CRPC (Alfonso Urbanucci et al., 2017). Braadland and Urbanucci suggest that selective or adaptive remodelling occurs mainly upon treatment challenge with AR-targeted therapies (Braadland & Urbanucci, 2019). Sequencing in independent AR overexpressing models by Urbanucci et al. revealed genome wide increases in open confirmations of chromatin and an increased opening at androgen responsive binding sites. Androgens further increased this chromatin opening, suggesting ligand potentiates an AR-driven chromatin remodelling in the context of AR overexpression (Alfonso Urbanucci et al., 2017). This represents a potential "feed forward" mechanism in which the overexpressed AR further facilitates chromatin remodelling that allows the AR greater access and increased binding at the genome (Braadland & Urbanucci, 2019). Additionally, progression to CRPC entails a significant reprogramming of the AR cistrome (Pomerantz et al., 2015; Sharma et al., 2013).

Mechanistic alteration of master regulators of the epigenome have been established to play a key role via increasing AR transcriptional activity (Ruggero et al., 2018), and their behaviour can be context sensitive. The chromatin remodelling enzyme lysine-specific demethylase 1 has emerged as having a dual role given its context-sensitive promotive or repressive effects on AR (Cai et al., 2011; Metzger et al., 2005). High androgen levels have been demonstrated to cause AR-mediated recruitment of LSD1 to facilitate gene silencing via negative autoregulation of the AR gene (Cai et al., 2011), while in the context of CRPC this feedback loop is apparently broken given that low androgen levels drive AR overexpression (Ruggero et al., 2018). LSD1 coactivator or corepressor activity is influenced by post-transcriptional modifications, such as its phosphorylation status which can switch the enzymes substrate (Metzger et al., 2007; Shi et al., 2004). The tyrosine kinase Src, upregulated in CRPC (Siu et al., 2016), inactivates the AR corepressor LCoR that ordinarily downregulates AR in response to ligand. This subsequently activates AR at the chromatin level in CRPC (Asim et al., 2011). A large number of micro-RNAs have been identified to act as post-transcriptional regulators of the AR (Perner et al., 2015). The miRNA miR137 regulates an androgen-mediated feedback loop that inhibits a large network of crucial AR coregulators in normal prostate epithelia, while epigenetic loss of miR137 in CRPC leads to coregulator and, consequently, AR overexpression (Nilsson et al., 2015).

It is notable that, in contrast with other DNA binding elements, the AR is able to initiate epigenetic modification of chromatin by itself (Tewari et al., 2012). Higher AR levels increase AR's genome-wide binding to chromatin upon stimulation with low concentration of ligand (Urbanucci et al., 2011). AR overexpression recruits AR and the basic epigenetic machinery to the chromatin to alter histones at AR binding sites and favour chromatin accessibility in the presence of low androgen levels (Alfonso Urbanucci et al., 2011). Chromatin remodeling proteins such as FOXA1 and HOXB13 are also known to co-localise with AR (Stelloo et al., 2017) and are capable of recruiting acetylating and methylating coregulators including CBP/p300 and MLL (Braadland & Urbanucci, 2019). Many coregulators of the AR exert chromatin remodelling effects themselves (Bannister & Kouzarides, 2011), and there is evidence that the AR upregulates a number of its coregulators gene-specifically through varied mechanisms, including AIB1, CBP, MAK, BRCA1, β -catenin, ATAD2, and MID1 (Perner et al., 2015; Alfonso Urbanucci et al., 2008, 2017). Several coregulators of the AR including p300, CBP and TIF2 have been shown to increase as a result of androgen deprivation (Agoulnik et al., 2006; Comuzzi et al., 2004; Heemers et al., 2007). Even a modest overexpression of AR can alter expression and amounts of AR coregulators (Chen et al., 2003), many of which are histone acetylating (Alfonso Urbanucci et al., 2011). Key bromodomain proteins, which locus-specifically affect chromatin opening, are androgen regulated and upregulated in AR overexpressing cells. These proteins participate in an AR deregulation-driven feedback loop that increases AR chromatin accessibility (Alfonso Urbanucci et al., 2017). The Jumonji C KDM4 histone lysine demethylases are overexpressed in CRPC, and KDM4B expression has been significantly correlated with AR. KDM4B influences chromatin and may induce relaxation in conditions of androgen deprivation that are relevant to progression to CRPC (Duan et al., 2019).

Gritsina et al. reviewed current knowledge regarding the function of AR signaling in driving target gene

repression and silencing by regulation of the epigenetic machinery. Ligand-bound AR binds to the enhancers and/or promoter elements of target genes and mediates assembly and recruitment of the repressive complexes, including histone deacetylases, lysine-specific demethylase 1, and enhancer of zeste homolog 2. AR directly and indirectly induces cascades involving the stabilisation of protein-protein interactions and recruitment of complexes responsible for the removal of acyl groups, demethylation, inhibition of transcriptional activators, and trimethylation, resulting in chromatin modifications that render gene regulatory elements inaccessible or silenced (Yu et al., 2019)?.

Taken together, research has identified a clear role for AR expression in genome-wide epigenetic status, along with the ability of the AR to recruit and drive the basic elements of the epigenetic machinery. Additionally, it is apparent that a refractory response to antiandrogenic treatment can occur irrespective of agent. With consideration to these findings, a potential feed-forward mechanism of AR overexpression, potentiated by androgens, may have significant mechanistic implications for the onset and progressive worsening of PFS with the "crash" after cessation of the medication, during which time the multi-systemic symptoms and physiological effects of the condition become apparent or intensify with significant interindividual variability in severity. This occurrence is most usually in a time frame of days or weeks, a timeframe correlating to the return and increase of endogenous DHT levels as the newly functional 5-alpha reductase enzyme is replenished. DHT has been demonstrated to alter the regulation of a number of AR coactivators gene-specifically depending on the level of the receptor, suggesting plausible involvement of coactivator regulation in a feedback loop potentiating increased AR signaling (Alfonso Urbanucci et al., 2008)?.

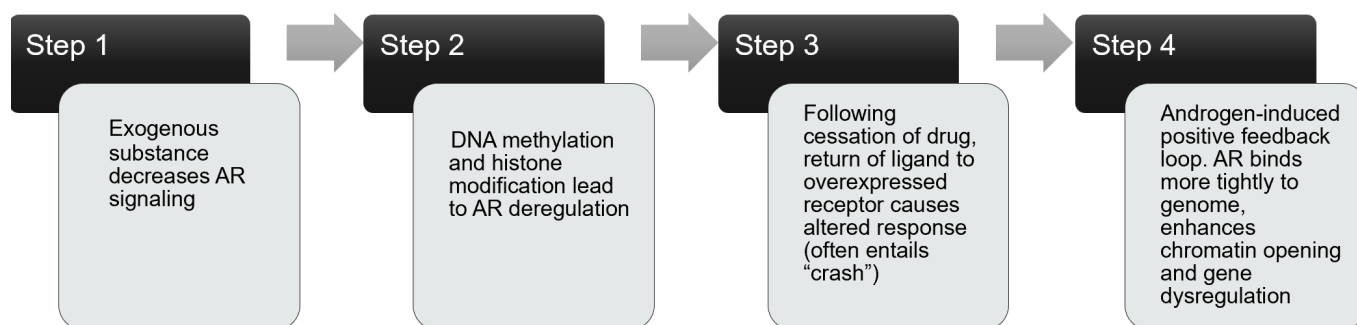


fig. A proposed basic mechanism in pathologically affected cells underlying the development of PFS.

As the transition to CRPC results from androgen deprivation or androgen-axis targeted treatment, an induction of AR deregulation could have relevance to the increased incidence of higher Gleason score prostate cancers in 5ari patients (Sarkar et al., 2019; Theoret et al., 2011; Traish et al., 2014; Van Rompay et al., 2018)?. It is of significance that, following three years of use and then cessation, finasteride has been demonstrated to accelerate the progression of male pattern hair loss significantly.

Using technologies unavailable at the time of finasteride's clinical approval, Van Neste recently reported the first evidence of what they describe as a "drug dependency" of terminal scalp hair follicles in AGA patients and a "post-finasteride rebound phenomenon" in patients who had stopped finasteride after 3 years of successful maintenance. During 3 years of finasteride use, 99%–100% terminal hair counts were recorded suggesting effective maintenance. However, while terminal hair was maintained on drug, within 30 months "off-drug" androgenic alopecia had significantly worsened, only 5.8% of terminal hair could be measured, with 94% having miniaturised and become unproductive. This is far in excess of the expected regression rates that were previously established in these patients and robustly predicted at 6% per year (Van Neste, 2019). It was previously reported that vertex dermal papilla cells in balding samples were 1.9 fold higher in AR expression than those from the occipital scalp (Kwon et al., 2004), and frontal follicles are 40% higher in AR expression in males compared with women (Sawaya & Price, 1997). Increased DNA methylation of the AR promoter in occipital follicles from men with AGA is suggestive of toxicity mediated by receptor levels (Cobb et al., 2011). In agreement, AGA models support an AR-mediated pathological process. Transgenic mice overexpressing human AR in the skin exhibit impaired hair regeneration when exposed to DHT, while hydroxyflutamide can abolish this effect (Crabtree et al., 2010). An adaptive increase in AR expression following androgen deprivation is therefore a plausible mechanistic explanation for an increase in hormone sensitivity causative of the dramatic finasteride-induced progression of male pattern hair loss observed by Van Neste. Similarly, epigenetic amplification in PFS could reflect the common reports of a significant acceleration in MPB following development of the condition.

Therapeutic responses to androgens and antiandrogens in PFS

There is no known therapeutic approach for PFS (Than et al., 2018) and no consistently safe or effective therapy has emerged from two decades of patient self-experimentation. Owing to the common PFS symptom profile ostensibly pointing towards decreased androgenic activity and low or hypogonadal levels of testosterone in some cases of PFS, many patients have undergone treatment with exogenous androgens. While this can be of benefit in some patients, it is very rare that this is complete or consistently effective even if a temporary improvement is observed in some symptoms. Remarkably, symptoms can be exacerbated by administration of androgens. This is well reported even in patients in whom PFS has caused a clinical hypogonadism. Patients receiving testosterone replacement therapy prior to PFS have reported a dramatic intolerance to exogenous androgens following the onset of the condition. Testosterone is ordinarily associated with a decrease in depression and improved verbal memory (Cherrier et al., 2014) as well as anxiolytic effect in men, women and animals (McHenry et al., 2014). The reverse has been well reported in PFS patients, even when hypogonadal. Beyond cognitive symptoms, sexual dysfunction and physical symptoms such as muscle wastage can be exacerbated. This is highly remarkable and paradoxical. A patient who since committed suicide reported further rapid penile shrinking upon local application of topical DHT gel at a dosage of 5g per day with therapeutic intent. This is striking and paradoxical with consideration as to the known effect of DHT in increasing penile size (Arteaga-Silva et al., 2008; Becker et al., 2016; Choi et al., 1993). Patients will often report feeling no response at all to high doses of testosterone. Interindividually variable "saturation" points with regard

to androgen response (Morgentaler & Traish, 2009; Zitzmann, 2009) have been hypothesised, and this may be of relevance to the therapeutic failure of testosterone in PFS. A threshold at which androgen-mediated toxicity reaches saturation has been observed with regard to the degree of symptoms seen in SBMA models (Chevalier-Larsen & Merry, 2011), in the toxic effect of DHT in SBMA motor neurons (Sheila et al., 2019), and in prostate cancer, in which testosterone therapy does not accelerate the disease progression despite androgen dependence (Morgentaler & Traish, 2009). In PFS, this reaction to exogenous ligand could plausibly be reflective of the degree of AR overexpression per site and per patient, and offers an explanation as to why more favourable partial responses to androgens are sometimes seen, while other patients can often rapidly worsen with raising androgen levels. Of note, it has been reported that an SBMA patient exhibited a notably similar reversible deterioration with androgen administration (Kinirons & Rouleau, 2008). Importantly, this would be in keeping with the observed responses of female transgenic mice overexpressing WT AR in skeletal muscle to exogenous testosterone equivalent to circulating male levels, which caused striking differences in deleterious physiological effects depending on the degree of AR overexpression (Monks et al., 2007).

Across the history of the propeciahelp forum, the most consequentially profound responses described entail significant modulation of symptoms by further exposure to substances that lower androgens through mechanisms including 5 alpha reductase inhibition, or substances that reduce concentrations of or inhibit AR. While rapid and severe worsening can occur, patients have equally often reported the dramatic return of function in the domains affected by PFS, usually temporarily. These are nearly always taken in the absence of the knowledge they are taking pharmaceuticals or natural extracts with antiandrogenic properties and are frequently sought out based upon their purported benefits in marketing and health editorials concerning relief of symptoms or through online reports from other patients. These have included zinc, quercetin, resveratrol, milk thistle, licorice root, turmeric/curcumin, sulforaphane, DIM, sodium butyrate, saw palmetto, tribulus terrestris, polyphenol rich products such as cacao nibs or pomegranate, and soy and soy isoflavones including genistein, all of which are notably antiandrogenic through various mechanisms (Agarwal et al., 2006; Boam, 2015; Cicero et al., 2019; De Amicis et al., 2019; Hiipakka et al., 2002; Jang et al., 2019; Kampa et al., 2017; Le et al., 2003; Sabbadin et al., 2019; Samykutty et al., 2013; Sandeep et al., 2015; Shiota et al., 2011; Xing, 2001). A remarkable overlap can be noted with nutraceuticals that are of increasing interest in the treatment of AR-mediated conditions and with substances or extracts causing patients to develop and present with PFS, as we have noted. Patients have independently described significant and remarkable multi-domain relief following use of AR antagonists including bicalutamide (Rice et al., 2019), and drugs with an antiandrogenic effects such as ibuprofen, paracetamol, dexamethasone, omeprazole, leuprolide acetate and mifepristone (Hoda et al., 2016; Inder et al., 2009; Kortenkamp, 2020; Kristensen et al., 2010; Song et al., 2004; Sørensen et al., 2016), and even finasteride itself. Recently, truvada, an antiretroviral medication combining tenofovir disoproxil and emtricitabine, has been reported to improve some PFS patients significantly in multiple symptom domains. Marketed as PrEP, truvada is a reverse transcriptase inhibitor. RTI drugs have been considered for potential therapeutic efficacy in hormonally refractive prostate cancer due to in vitro results suggesting the capability of Nevirapine to induce extensive reprogramming of gene expression, resensitizing cells to stimulation by extracellular ligand and consequentially re-establishing the efficacy of antiandrogen treatment with bicalutamide (Landriscina et al., 2009).

These common reports are highly remarkable and of relevance to the potential of a pathologic link between PolyQ toxicity and deleterious consequences of site-specific overexpression of the wild type androgen receptor. This would appear to be in alignment with functional rescue in SBMA models targeting the androgen pathway (Cortes & La Spada, 2018; Katsuno et al., 2003; Minamiyama, 2004; Nedelsky et al., 2010; Rinaldi et al., 2015), and the molecular level responses to androgens and antiandrogens in AR overexpressing CRPC as discussed.

It is of the utmost importance to establish that antiandrogenic therapeutic strategies are dangerous for PFS patients. Patients can persistently exacerbate or develop further symptoms in multiple domains of the condition upon rechallenge or subsequent exposure to substances with antiandrogenic effect. This often occurs after a dramatic improvement of existing multisystemic symptoms. In 2018, a PFS patient who had taken supplementary resveratrol described a profound reversal of symptoms including insomnia, erectile dysfunction, libido loss and fatigue shortly before taking his own life. We note a key vulnerability of this cohort to what we believe to be an aberrant epigenetic response following exposure to antiandrogenic substances. This vulnerability appears significantly exacerbated following initial development of PFS, and even phenol or isoflavone-rich foods have resulted in clear reports of persistent worsening or the triggering of further symptoms. PFS patients most at risk of this are, in our experience, those who present with severe phenotypes after short use of finasteride or a causative antiandrogenic substance. Therefore, until more is known regarding the molecular mechanisms underlying the development of PFS, we strongly urge physicians dealing with PFS patients to be aware of this unique and peculiar vulnerability to therapeutic substances or medicines with antiandrogenic modality. This is of relevance to both prescribed therapies such as SRI antidepressants and to self-driven “natural” therapeutic attempts that can involve high dose phenolic compounds or vitamins marketed as health supplements. Owing to the sometimes profound endocrine sensitivity induced by PFS, safely managing the condition can be a significant burden for patients.

Page Bibliography

1. Agarwal, R., Agarwal, C., Ichikawa, H., Singh, R., & Aggarwal, B. (2006). Anticancer potential of silymarin: from bench to bed side. *Anticancer Research*, 26(6B), 4457–4498. <https://www.ncbi.nlm.nih.gov/pubmed/17201169>
2. Agoulnik, I. U., Vaid, A., Nakka, M., Alvarado, M., Bingman, W. E., III, Erdem, H., Frolov, A., Smith, C. L., Ayala, G. E., Ittmann, M. M., & Weigel, N. L. (2006). Androgens Modulate Expression of Transcription Intermediary Factor 2, an Androgen Receptor Coactivator whose Expression Level Correlates with Early Biochemical Recurrence in Prostate Cancer. *Cancer Research*, 10594–10602. <https://doi.org/10.1158/0008-5472.can-06-1023>
3. Amadi, C., Odum, E. P., & Aleme, B. M. (2018). Low endogenous testosterone level is associated

- with high-risk prostate-specific antigen level in men with prostate cancer disease. *International Surgery Journal*, 1617. <https://doi.org/10.18203/2349-2902.isj20181581>
4. Arteaga-Silva, M., Viguera-Villaseñor, R. M., Retana-Márquez, S., Hernández-González, M., Chihuahua-Serrano, C., Bonilla-Jaime, H., Contreras, J. L., & Morali, G. (2008). Testosterone, androstenedione, and 5 α -dihydrotestosterone on male sexual behavior and penile spines in the hamster. *Physiology & Behavior*, 412–421. <https://doi.org/10.1016/j.physbeh.2008.02.007>
 5. Asim, M., Hafeez, B. B., Siddiqui, I. A., Gerlach, C., Patz, M., Mukhtar, H., & Baniahmad, A. (2011). Ligand-dependent Corepressor Acts as a Novel Androgen Receptor Corepressor, Inhibits Prostate Cancer Growth, and Is Functionally Inactivated by the Src Protein Kinase. *Journal of Biological Chemistry*, 37108–37117. <https://doi.org/10.1074/jbc.m111.292771>
 6. Bannister, A. J., & Kouzarides, T. (2011). Regulation of chromatin by histone modifications. *Cell Research*, 381–395. <https://doi.org/10.1038/cr.2011.22>
 7. Barbieri, C. E., Baca, S. C., Lawrence, M. S., Demichelis, F., Blattner, M., Theurillat, J.-P., White, T. A., Stojanov, P., Van Allen, E., Stransky, N., Nickerson, E., Chae, S.-S., Boysen, G., Auclair, D., Onofrio, R. C., Park, K., Kitabayashi, N., MacDonald, T. Y., Sheikh, K., ... Garraway, L. A. (2012). Exome sequencing identifies recurrent SPOP, FOXA1 and MED12 mutations in prostate cancer. *Nature Genetics*, 685–689. <https://doi.org/10.1038/ng.2279>
 8. Becker, D., Wain, L. M., Chong, Y. H., Gosai, S. J., Henderson, N. K., Milburn, J., Stott, V., & Wheeler, B. J. (2016). Topical dihydrotestosterone to treat micropenis secondary to partial androgen insensitivity syndrome (PAIS) before, during, and after puberty – a case series. *Journal of Pediatric Endocrinology and Metabolism*. <https://doi.org/10.1515/jpem-2015-0175>
 9. Boam, T. (2015). Anti-androgenic effects of flavonols in prostate cancer. *Ecancermedicalscience*. <https://doi.org/10.3332/ecancer.2015.585>
 10. Braadland, P. R., & Urbanucci, A. (2019). Chromatin reprogramming as an adaptation mechanism in advanced prostate cancer. *Endocrine-Related Cancer*, R211–R235. <https://doi.org/10.1530/erc-18-0579>
 11. Cai, C., He, H. H., Chen, S., Coleman, I., Wang, H., Fang, Z., Chen, S., Nelson, P. S., Liu, X. S., Brown, M., & Balk, S. P. (2011). Androgen Receptor Gene Expression in Prostate Cancer Is Directly Suppressed by the Androgen Receptor Through Recruitment of Lysine-Specific Demethylase 1. *Cancer Cell*, 457–471. <https://doi.org/10.1016/j.ccr.2011.09.001>

12. Chen, C. D., Welsbie, D. S., Tran, C., Baek, S. H., Chen, R., Vessella, R., Rosenfeld, M. G., & Sawyers, C. L. (2003). Molecular determinants of resistance to antiandrogen therapy. *Nature Medicine*, 33–39. <https://doi.org/10.1038/nm972>
13. Cherrier, M. M., Anderson, K., Shofer, J., Millard, S., & Matsumoto, A. M. (2014). Testosterone Treatment of Men With Mild Cognitive Impairment and Low Testosterone Levels. *American Journal of Alzheimer's Disease & Other Dementias*, 421–430. <https://doi.org/10.1177/1533317514556874>
14. Chevalier-Larsen, E. S., & Merry, D. E. (2011). Testosterone treatment fails to accelerate disease in a transgenic mouse model of spinal and bulbar muscular atrophy. *Disease Models & Mechanisms*, 141–145. <https://doi.org/10.1242/dmm.007849>
15. Choi, S. K., Han, S. W., Kim, D. H., & Lignieres, B. de. (1993). Transdermal Dihydrotestosterone Therapy and its Effects on Patients with Microphallus. *Journal of Urology*, 657–660. [https://doi.org/10.1016/s0022-5347\(17\)35576-3](https://doi.org/10.1016/s0022-5347(17)35576-3)
16. Christensen, B. R., Barata, P. C., Ledet, E. M., Layton, J. L., Lewis, B. E., & Sartor, O. (2019). High-Dose Testosterone and Radium-223 Response in Metastatic Castration-Resistant Prostate Cancer. *Clinical Genitourinary Cancer*, 476–479. <https://doi.org/10.1016/j.clgc.2019.07.022>
17. Cicero, A. F. G., Allkanjari, O., Busetto, G. M., Cai, T., Larganà, G., Magri, V., Perletti, G., Robustelli Della Cuna, F. S., Russo, G. I., Stamatiou, K., Trinchieri, A., & Vitalone, A. (2019). Nutraceutical treatment and prevention of benign prostatic hyperplasia and prostate cancer. *Archivio Italiano Di Urologia e Andrologia*. <https://doi.org/10.4081/aiua.2019.3.139>
18. Cobb, J. E., Wong, N. C., Yip, L. W., Martinick, J., Bosnich, R., Sinclair, R. D., Craig, J. M., Saffery, R., Harrap, S. B., & Ellis, J. A. (2011). Evidence of increased DNA methylation of the androgen receptor gene in occipital hair follicles from men with androgenetic alopecia. *British Journal of Dermatology*, 210–213. <https://doi.org/10.1111/j.1365-2133.2011.10335.x>
19. Comuzzi, B., Nemes, C., Schmidt, S., Jasarevic, Z., Lodde, M., Pycha, A., Bartsch, G., Offner, F., Culig, Z., & Hobisch, A. (2004). The androgen receptor co-activator CBP is up-regulated following androgen withdrawal and is highly expressed in advanced prostate cancer. *The Journal of Pathology*, 159–166. <https://doi.org/10.1002/path.1609>

20. Corradi, L. S., Góes, R. M., Vilamaior, P. S. L., & Taboga, S. R. (2009). Increased androgen receptor and remodeling in the prostatic stroma after the inhibition of 5-alpha reductase and aromatase in gerbil ventral prostate. *Microscopy Research and Technique*, 939–950. <https://doi.org/10.1002/jemt.20740>

21. Cortes, C. J., & La Spada, A. R. (2018). X-Linked Spinal and Bulbar Muscular Atrophy: From Clinical Genetic Features and Molecular Pathology to Mechanisms Underlying Disease Toxicity. In *Polyglutamine Disorders* (pp. 103–133). Springer International Publishing. https://doi.org/10.1007/978-3-319-71779-1_5

22. Coskuner, E. R., Ozkan, B., & Culha, M. G. (2019). Sexual Problems of Men With Androgenic Alopecia Treated With 5-Alpha Reductase Inhibitors. *Sexual Medicine Reviews*, 277–282. <https://doi.org/10.1016/j.sxmr.2018.07.003>

23. Coutinho, I., Day, T. K., Tilley, W. D., & Selth, L. A. (2016). Androgen receptor signaling in castration-resistant prostate cancer: a lesson in persistence. *Endocrine-Related Cancer*, T179–T197. <https://doi.org/10.1530/erc-16-0422>

24. Crabtree, J. S., Kilbourne, E. J., Peano, B. J., Chippari, S., Kenney, T., McNally, C., Wang, W., Harris, H. A., Winneker, R. C., Nagpal, S., & Thompson, C. C. (2010). A Mouse Model of Androgenetic Alopecia. *Endocrinology*, 2373–2380. <https://doi.org/10.1210/en.2009-1474>

25. De Amicis, F., Chimento, A., Montalto, F., Casaburi, I., Sirianni, R., & Pezzi, V. (2019). Steroid Receptor Signallings as Targets for Resveratrol Actions in Breast and Prostate Cancer. *International Journal of Molecular Sciences*, 1087. <https://doi.org/10.3390/ijms20051087>

26. Di Loreto, C., La Marra, F., Mazzon, G., Belgrano, E., Trombetta, C., & Cauci, S. (2014). Immunohistochemical Evaluation of Androgen Receptor and Nerve Structure Density in Human Prepuce from Patients with Persistent Sexual Side Effects after Finasteride Use for Androgenetic Alopecia. *PLoS ONE*, e100237. <https://doi.org/10.1371/journal.pone.0100237>

27. Duan, L., Chen, Z., Lu, J., Liang, Y., Wang, M., Roggero, C. M., Zhang, Q.-J., Gao, J., Fang, Y., Cao, J., Lu, J., Zhao, H., Dang, A., Pong, R.-C., Hernandez, E., Chang, C.-M., Hoang, D. T., Ahn, J.-M., Xiao, G., ... Liu, Z.-P. (2019). Histone lysine demethylase KDM4B regulates the alternative splicing of the androgen receptor in response to androgen deprivation. *Nucleic Acids Research*. <https://doi.org/10.1093/nar/gkz1004>

28. Enatsu, N., Chiba, K., Sumii, K., Fukuda, T., Okada, K., Matsushita, K., & Fujisawa, M. (2016).

Dutasteride-mediated morphological changes in the genitourinary tract associated with altered expression patterns of the androgen and estrogen receptors in male rats. *Andrology*, 347–353.

<https://doi.org/10.1111/andr.12297>

29. Friedlander, T. W., Roy, R., Tomlins, S. A., Ngo, V. T., Kobayashi, Y., Azameera, A., Rubin, M. A., Pienta, K. J., Chinnaiyan, A., Ittmann, M. M., Ryan, C. J., & Paris, P. L. (2011). Common Structural and Epigenetic Changes in the Genome of Castration-Resistant Prostate Cancer. *Cancer Research*, 616–625. <https://doi.org/10.1158/0008-5472.can-11-2079>
30. Grasso, C. S., Wu, Y.-M., Robinson, D. R., Cao, X., Dhanasekaran, S. M., Khan, A. P., Quist, M. J., Jing, X., Lonigro, R. J., Brenner, J. C., Asangani, I. A., Ateeq, B., Chun, S. Y., Siddiqui, J., Sam, L., Anstett, M., Mehra, R., Prensner, J. R., Palanisamy, N., ... Tomlins, S. A. (2012). The mutational landscape of lethal castration-resistant prostate cancer. *Nature*, 239–243. <https://doi.org/10.1038/nature11125>
31. Gravina, G. L., Marampon, F., Piccolella, M., Motta, M., Ventura, L., Pomante, R., Popov, V. M., Zani, B. M., Pestell, R. G., Tombolini, V., Jannini, E. A., & Festuccia, C. (2011). Hormonal Therapy Promotes Hormone-Resistant Phenotype by Increasing DNMT Activity and Expression in Prostate Cancer Models. *Endocrinology*, 4550–4561. <https://doi.org/10.1210/en.2011-1056>
32. Halievski, K., Nath, S., Katsuno, M., Adachi, H., Sobue, G., Breedlove, S., Lieberman, A., & Jordan, C. (2019). Disease Affects Bdnf Expression in Synaptic and Extrasynaptic Regions of Skeletal Muscle of Three SBMA Mouse Models. *International Journal of Molecular Sciences*, 1314. <https://doi.org/10.3390/ijms20061314>
33. Heemers, H. V., Sebo, T. J., Debes, J. D., Regan, K. M., Raclaw, K. A., Murphy, L. M., Hobisch, A., Culig, Z., & Tindall, D. J. (2007). Androgen Deprivation Increases p300 Expression in Prostate Cancer Cells. *Cancer Research*, 3422–3430. <https://doi.org/10.1158/0008-5472.can-06-2836>
34. Hiipakka, R. A., Zhang, H.-Z., Dai, W., Dai, Q., & Liao, S. (2002). Structure–activity relationships for inhibition of human 5 α -reductases by polyphenols. *Biochemical Pharmacology*, 1165–1176. [https://doi.org/10.1016/s0006-2952\(02\)00848-1](https://doi.org/10.1016/s0006-2952(02)00848-1)
35. Hoda, M. R., Kramer, M. W., Merseburger, A. S., & Cronauer, M. V. (2016). Androgen deprivation therapy with Leuprolide acetate for treatment of advanced prostate cancer. *Expert Opinion on Pharmacotherapy*, 105–113. <https://doi.org/10.1080/14656566.2016.1258058>

36. Hsieh, J.-T., Chen, S.-C., Yu, H.-J., & Chang, H.-C. (2011). Finasteride upregulates expression of androgen receptor in hyperplastic prostate and LNCaP cells: Implications for chemoprevention of prostate cancer. *The Prostate*, 1115–1121. <https://doi.org/10.1002/pros.21325>
37. Inder, W. J., Jang, C., Obeyesekere, V. R., & Alford, F. P. (2009). Dexamethasone administration inhibits skeletal muscle expression of the androgen receptor and IGF-1 - implications for steroid-induced myopathy. *Clinical Endocrinology*. <https://doi.org/10.1111/j.1365-2265.2009.03683.x>
38. Irwig, M. S. (2014). Androgen Levels and Semen Parameters Among Former Users of Finasteride With Persistent Sexual Adverse Effects. *JAMA Dermatology*, 1361. <https://doi.org/10.1001/jamadermatol.2014.1830>
39. Jang, Y.-G., Go, R.-E., Hwang, K.-A., & Choi, K.-C. (2019). Resveratrol inhibits DHT-induced progression of prostate cancer cell line through interfering with the AR and CXCR4 pathway. *The Journal of Steroid Biochemistry and Molecular Biology*, 105406. <https://doi.org/10.1016/j.jsbmb.2019.105406>
40. Kampa, M., Notas, G., & Castanas, E. (2017). Natural extranuclear androgen receptor ligands as endocrine disruptors of cancer cell growth. *Molecular and Cellular Endocrinology*, 43–48. <https://doi.org/10.1016/j.mce.2017.02.021>
41. Kanherkar, R. R., Getachew, B., Ben-Sheetrit, J., Varma, S., Heinbockel, T., Tizabi, Y., & Csoka, A. B. (2018). The Effect of Citalopram on Genome-Wide DNA Methylation of Human Cells. *International Journal of Genomics*, 1–12. <https://doi.org/10.1155/2018/8929057>
42. Katsuno, M., Adachi, H., Doyu, M., Minamiyama, M., Sang, C., Kobayashi, Y., Inukai, A., & Sobue, G. (2003). Leuprorelin rescues polyglutamine-dependent phenotypes in a transgenic mouse model of spinal and bulbar muscular atrophy. *Nature Medicine*, 768–773. <https://doi.org/10.1038/nm878>
43. Kaufman, J. M., & Vermeulen, A. (2005). The Decline of Androgen Levels in Elderly Men and Its Clinical and Therapeutic Implications. *Endocrine Reviews*, 833–876. <https://doi.org/10.1210/er.2004-0013>
44. Kim, W., & Ryan, C. J. (2012). Androgen Receptor Directed Therapies in Castration-Resistant Metastatic Prostate Cancer. *Current Treatment Options in Oncology*, 189–200. <https://doi.org/10.1007/s11864-012-0188-2>

45. Kinirons, P., & Rouleau, G. A. (2008). Administration of testosterone results in reversible deterioration in Kennedy's disease. *Journal of Neurology, Neurosurgery & Psychiatry*, 106–107. <https://doi.org/10.1136/jnmp.2006.101899>
46. Kortenkamp, A. (2020). Which chemicals should be grouped together for mixture risk assessments of male reproductive disorders? *Molecular and Cellular Endocrinology*, 110581. <https://doi.org/10.1016/j.mce.2019.110581>
47. Kristensen, D. M., Hass, U., Lesné, L., Lottrup, G., Jacobsen, P. R., Desdoits-Lethimonier, C., Boberg, J., Petersen, J. H., Toppari, J., Jensen, T. K., Brunak, S., Skakkebæk, N. E., Nellemann, C., Main, K. M., Jégou, B., & Leffers, H. (2010). Intrauterine exposure to mild analgesics is a risk factor for development of male reproductive disorders in human and rat. *Human Reproduction*, 235–244. <https://doi.org/10.1093/humrep/deq323>
48. Kwon, O. S., Han, J. H., Yoo, H. G., Lee, S. R., Kim, K. H., Eun, H. C., Sim, Y. C., & Cho, K. H. (2004). Expression of androgen receptor, estrogen receptor α and β in the dermal papilla of human hair follicles in vivo. *Journal of Dermatological Science*, 176–179. <https://doi.org/10.1016/j.jdermsci.2004.09.004>
49. Landriscina, M., Bagalà, C., Piscazzi, A., Schinzari, G., Quirino, M., Fabiano, A., Bianchetti, S., Cassano, A., Sica, G., & Barone, C. (2009). Nevirapine restores androgen signaling in hormone-refractory human prostate carcinoma cells both in vitro and in vivo. *The Prostate*, 744–754. <https://doi.org/10.1002/pros.20923>
50. Le, H. T., Schaldach, C. M., Firestone, G. L., & Bjeldanes, L. F. (2003). Plant-derived 3,3'-Diindolylmethane Is a Strong Androgen Antagonist in Human Prostate Cancer Cells. *Journal of Biological Chemistry*, 21136–21145. <https://doi.org/10.1074/jbc.m300588200>
51. MacArthur, B. D., Ma'ayan, A., & Lemischka, I. R. (2009). Systems biology of stem cell fate and cellular reprogramming. *Nature Reviews Molecular Cell Biology*, 672–681. <https://doi.org/10.1038/nrm2766>
52. McCarthy, M. M. (2019). Is sexual differentiation of brain and behavior epigenetic? *Current Opinion in Behavioral Sciences*, 83–88. <https://doi.org/10.1016/j.cobeha.2018.10.005>
53. McHenry, J., Carrier, N., Hull, E., & Kabbaj, M. (2014). Sex differences in anxiety and depression: Role of testosterone. *Frontiers in Neuroendocrinology*, 42–57.

<https://doi.org/10.1016/j.yfrne.2013.09.001>

54. Melcangi, R. C., Santi, D., Spezzano, R., Grimoldi, M., Tabacchi, T., Fusco, M. L., Diviccaro, S., Giatti, S., Carrà, G., Caruso, D., Simoni, M., & Cavaletti, G. (2017). Neuroactive steroid levels and psychiatric and andrological features in post-finasteride patients. *The Journal of Steroid Biochemistry and Molecular Biology*, 229–235. <https://doi.org/10.1016/j.jsbmb.2017.04.003>
55. Metzger, E., Wissmann, M., Yin, N., Müller, J. M., Schneider, R., Peters, A. H. F. M., Günther, T., Buettner, R., & Schüle, R. (2005). LSD1 demethylates repressive histone marks to promote androgen-receptor-dependent transcription. *Nature*, 436–439. <https://doi.org/10.1038/nature04020>
56. Metzger, E., Yin, N., Wissmann, M., Kunowska, N., Fischer, K., Friedrichs, N., Patnaik, D., Higgins, J. M. G., Potier, N., Scheidtmann, K.-H., Buettner, R., & Schüle, R. (2007). Phosphorylation of histone H3 at threonine 11 establishes a novel chromatin mark for transcriptional regulation. *Nature Cell Biology*, 53–60. <https://doi.org/10.1038/ncb1668>
57. Minamiyama, M. (2004). Sodium butyrate ameliorates phenotypic expression in a transgenic mouse model of spinal and bulbar muscular atrophy. *Human Molecular Genetics*, 1183–1192. <https://doi.org/10.1093/hmg/ddh131>
58. Mo, K., Razak, Z., Rao, P., Yu, Z., Adachi, H., Katsuno, M., Sobue, G., Lieberman, A. P., Westwood, J. T., & Monks, D. A. (2010). Microarray Analysis of Gene Expression by Skeletal Muscle of Three Mouse Models of Kennedy Disease/Spinal Bulbar Muscular Atrophy. *PLoS ONE*, e12922. <https://doi.org/10.1371/journal.pone.0012922>
59. Monks, D. A., Johansen, J. A., Mo, K., Rao, P., Eagleson, B., Yu, Z., Lieberman, A. P., Breedlove, S. M., & Jordan, C. L. (2007). Overexpression of wild-type androgen receptor in muscle recapitulates polyglutamine disease. *Proceedings of the National Academy of Sciences*, 18259–18264. <https://doi.org/10.1073/pnas.0705501104>
60. Monks, D. A., & Swift-Gallant, A. (2018). Non-neural androgen receptors affect sexual differentiation of brain and behaviour. *Journal of Neuroendocrinology*, e12493. <https://doi.org/10.1111/jne.12493>
61. Morgentaler, A., & Traish, A. M. (2009). Shifting the Paradigm of Testosterone and Prostate Cancer: The Saturation Model and the Limits of Androgen-Dependent Growth. *European Urology*, 310–321. <https://doi.org/10.1016/j.eururo.2008.09.024>

62. Nedelsky, N. B., Pennuto, M., Smith, R. B., Palazzolo, I., Moore, J., Nie, Z., Neale, G., & Taylor, J. P. (2010). Native Functions of the Androgen Receptor Are Essential to Pathogenesis in a *Drosophila* Model of Spinobulbar Muscular Atrophy. *Neuron*, 936–952.
<https://doi.org/10.1016/j.neuron.2010.08.034>
63. Nilsson, E. M., Laursen, K. B., Whitchurch, J., McWilliam, A., Ødum, N., Persson, J. L., Heery, D. M., Gudas, L. J., & Mongan, N. P. (2015). *MiR137* is an androgen regulated repressor of an extended network of transcriptional coregulators. *Oncotarget*.
<https://doi.org/10.18632/oncotarget.5958>
64. Ohshima, K., Hatakeyama, K., Nagashima, T., Watanabe, Y., Kanto, K., Doi, Y., Ide, T., Shimoda, Y., Tanabe, T., Ohnami, S., Ohnami, S., Serizawa, M., Maruyama, K., Akiyama, Y., Urakami, K., Kusahara, M., Mochizuki, T., & Yamaguchi, K. (2017). Integrated analysis of gene expression and copy number identified potential cancer driver genes with amplification-dependent overexpression in 1,454 solid tumors. *Scientific Reports*.
<https://doi.org/10.1038/s41598-017-00219-3>
65. Paulson, H. L., Shakkottai, V. G., Clark, H. B., & Orr, H. T. (2017). Polyglutamine spinocerebellar ataxias — from genes to potential treatments. *Nature Reviews Neuroscience*, 613–626. <https://doi.org/10.1038/nrn.2017.92>
66. Perner, S., Cronauer, M. V., Schrader, A. J., Klocker, H., Culig, Z., & Baniahmad, A. (2015). Adaptive responses of androgen receptor signaling in castration-resistant prostate cancer. *Oncotarget*. <https://doi.org/10.18632/oncotarget.4689>
67. Pihlajamaa, P., Sahu, B., & Jänne, O. A. (2015). Determinants of Receptor- and Tissue-Specific Actions in Androgen Signaling. *Endocrine Reviews*, 357–384.
<https://doi.org/10.1210/er.2015-1034>
68. Pomerantz, M. M., Li, F., Takeda, D. Y., Lenci, R., Chonkar, A., Chabot, M., Cejas, P., Vazquez, F., Cook, J., Shivdasani, R. A., Bowden, M., Lis, R., Hahn, W. C., Kantoff, P. W., Brown, M., Loda, M., Long, H. W., & Freedman, M. L. (2015). The androgen receptor cistrome is extensively reprogrammed in human prostate tumorigenesis. *Nature Genetics*, 1346–1351.
<https://doi.org/10.1038/ng.3419>
69. Prelich, G. (2012). Gene Overexpression: Uses, Mechanisms, and Interpretation. *Genetics*, 841–854. <https://doi.org/10.1534/genetics.111.136911>

70. Querin, G., Sorarù, G., & Pradat, P.-F. (2017). Kennedy disease (X-linked recessive bulbospinal neuronopathy): A comprehensive review from pathophysiology to therapy. *Revue Neurologique*, 326–337. <https://doi.org/10.1016/j.neurol.2017.03.019>
71. Rice, M. A., Malhotra, S. V., & Stoyanova, T. (2019). Second-Generation Antiandrogens: From Discovery to Standard of Care in Castration Resistant Prostate Cancer. *Frontiers in Oncology*. <https://doi.org/10.3389/fonc.2019.00801>
72. Rinaldi, C., Malik, B., & Greensmith, L. (2015). Targeted Molecular Therapies for SBMA. *Journal of Molecular Neuroscience*, 335–342. <https://doi.org/10.1007/s12031-015-0676-5>
73. Robinson, D., Van Allen, E. M., Wu, Y.-M., Schultz, N., Lonigro, R. J., Mosquera, J.-M., Montgomery, B., Taplin, M.-E., Pritchard, C. C., Attard, G., Beltran, H., Abida, W., Bradley, R. K., Vinson, J., Cao, X., Vats, P., Kunju, L. P., Hussain, M., Feng, F. Y., ... Chinnaiyan, A. M. (2015). Integrative Clinical Genomics of Advanced Prostate Cancer. *Cell*, 1215–1228. <https://doi.org/10.1016/j.cell.2015.05.001>
74. Ruggero, K., Farran-Matas, S., Martinez-Tebar, A., & Aytes, A. (2018). Epigenetic Regulation in Prostate Cancer Progression. *Current Molecular Biology Reports*, 101–115. <https://doi.org/10.1007/s40610-018-0095-9>
75. Ryan, C. J. (2018). *The Virility Paradox: The Vast Influence of Testosterone on Our Bodies, Minds, and the World We Live In*. BenBella Books.
76. Sabbadin, C., Bordin, L., Donà, G., Manso, J., Avruscio, G., & Armanini, D. (2019). Licorice: From Pseudohyperaldosteronism to Therapeutic Uses. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2019.00484>
77. Samykutty, A., Shetty, A. V., Dakshinamoorthy, G., Kalyanasundaram, R., Zheng, G., Chen, A., Bosland, M. C., Kajdacsy-Balla, A., & Gnanasekar, M. (2013). Vitamin K2, a Naturally Occurring Menaquinone, Exerts Therapeutic Effects on Both Hormone-Dependent and Hormone-Independent Prostate Cancer Cells. *Evidence-Based Complementary and Alternative Medicine*, 1–15. <https://doi.org/10.1155/2013/287358>
78. San Francisco, I. F., Rojas, P. A., DeWolf, W. C., & Morgentaler, A. (2014). Low free testosterone levels predict disease reclassification in men with prostate cancer undergoing active surveillance. *BJU International*, 229–235. <https://doi.org/10.1111/bju.12682>

79. Sandeep, P. M., Bovee, T. F. H., & Sreejith, K. (2015). Anti-Androgenic Activity of *Nardostachys jatamansi* DC and *Tribulus terrestris* L. and Their Beneficial Effects on Polycystic Ovary Syndrome–Induced Rat Models. *Metabolic Syndrome and Related Disorders*, 248–254. <https://doi.org/10.1089/met.2014.0136>
80. Sarkar, R. R., Parsons, J. K., Bryant, A. K., Ryan, S. T., Kader, A. K., McKay, R. R., D'Amico, A. V., Nguyen, P. L., Hulley, B. J., Einck, J. P., Mundt, A. J., Kane, C. J., Murphy, J. D., & Rose, B. S. (2019). Association of Treatment With 5 α -Reductase Inhibitors With Time to Diagnosis and Mortality in Prostate Cancer. *JAMA Internal Medicine*, 812. <https://doi.org/10.1001/jamainternmed.2019.0280>
81. Sawaya, M. E., & Price, V. H. (1997). Different Levels of 5 α -Reductase Type I and II, Aromatase, and Androgen Receptor in Hair Follicles of Women and Men with Androgenetic Alopecia. *Journal of Investigative Dermatology*, 296–300. <https://doi.org/10.1111/1523-1747.ep12335779>
82. Schuppe, E. R., Miles, M. C., & Fuxjager, M. J. (2020). Evolution of the androgen receptor: Perspectives from human health to dancing birds. *Molecular and Cellular Endocrinology*, 110577. <https://doi.org/10.1016/j.mce.2019.110577>
83. Seftel, A. (2005). Male hypogonadism. Part II: etiology, pathophysiology, and diagnosis. *International Journal of Impotence Research*, 223–228. <https://doi.org/10.1038/sj.ijir.3901365>
84. Sharma, N. L., Massie, C. E., Ramos-Montoya, A., Zecchini, V., Scott, H. E., Lamb, A. D., MacArthur, S., Stark, R., Warren, A. Y., Mills, I. G., & Neal, D. E. (2013). The Androgen Receptor Induces a Distinct Transcriptional Program in Castration-Resistant Prostate Cancer in Man. *Cancer Cell*, 35–47. <https://doi.org/10.1016/j.ccr.2012.11.010>
85. Shastry, B. S. (1995). Overexpression of genes in health and sickness. A bird's eye view. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 1–13. [https://doi.org/10.1016/0305-0491\(95\)00055-d](https://doi.org/10.1016/0305-0491(95)00055-d)
86. Sheila, M., Narayanan, G., Ma, S., Tam, W. L., Chai, J., & Stanton, L. W. (2019). Phenotypic and molecular features underlying neurodegeneration of motor neurons derived from spinal and bulbar muscular atrophy patients. *Neurobiology of Disease*, 1–13. <https://doi.org/10.1016/j.nbd.2018.10.019>

87. Shi, Y., Lan, F., Matson, C., Mulligan, P., Whetstone, J. R., Cole, P. A., Casero, R. A., & Shi, Y. (2004). Histone Demethylation Mediated by the Nuclear Amine Oxidase Homolog LSD1. *Cell*, 941–953. <https://doi.org/10.1016/j.cell.2004.12.012>
88. Shiota, M., Yokomizo, A., & Naito, S. (2011). Increased androgen receptor transcription: a cause of castration-resistant prostate cancer and a possible therapeutic target. *Journal of Molecular Endocrinology*, R25–R41. <https://doi.org/10.1530/jme-11-0018>
89. Siu, M. K., Chen, W.-Y., Tsai, H.-Y., Yeh, H.-L., Yin, J. J., Liu, S.-Y., & Liu, Y.-N. (2016). Androgen receptor regulates SRC expression through microRNA-203. *Oncotarget*. <https://doi.org/10.18632/oncotarget.8366>
90. Slack, J. M. W. (2002). Conrad Hal Waddington: the last Renaissance biologist? *Nature Reviews Genetics*, 889–895. <https://doi.org/10.1038/nrg933>
91. Song, L.-N., Coghlan, M., & Gelmann, E. P. (2004). Antiandrogen Effects of Mifepristone on Coactivator and Corepressor Interactions with the Androgen Receptor. *Molecular Endocrinology*, 70–85. <https://doi.org/10.1210/me.2003-0189>
92. Sørensen, A. M., Hansen, C. H., Bonomo, S., Olsen, L., Jørgensen, F. S., Weisser, J. J., Kretschmann, A. C., & Styrisshave, B. (2016). Enantioselective endocrine disrupting effects of omeprazole studied in the H295R cell assay and by molecular modeling. *Toxicology in Vitro*, 71–80. <https://doi.org/10.1016/j.tiv.2016.03.007>
93. Stelloo, S., Nevedomskaya, E., Kim, Y., Hoekman, L., Bleijerveld, O. B., Mirza, T., Wessels, L. F. A., van Weerden, W. M., Altelaar, A. F. M., Bergman, A. M., & Zwart, W. (2017). Endogenous androgen receptor proteomic profiling reveals genomic subcomplex involved in prostate tumorigenesis. *Oncogene*, 313–322. <https://doi.org/10.1038/onc.2017.330>
94. Swerdloff, R. S., Dudley, R. E., Page, S. T., Wang, C., & Salameh, W. A. (2017). Dihydrotestosterone: Biochemistry, Physiology, and Clinical Implications of Elevated Blood Levels. *Endocrine Reviews*, 220–254. <https://doi.org/10.1210/er.2016-1067>
95. Swift-Gallant, A., Coome, L. A., Ramzan, F., & Monks, D. A. (2016). Nonneural Androgen Receptors Affect Sexual Differentiation of Brain and Behavior. *Endocrinology*, 788–798. <https://doi.org/10.1210/en.2015-1355>

96. Takeda, D. Y., Spisák, S., Seo, J.-H., Bell, C., O'Connor, E., Korthauer, K., Ribli, D., Csabai, I., Solymosi, N., Szállási, Z., Stillman, D. R., Cejas, P., Qiu, X., Long, H. W., Tisza, V., Nuzzo, P. V., Rohanizadegan, M., Pomerantz, M. M., Hahn, W. C., & Freedman, M. L. (2018). A Somatic Acquired Enhancer of the Androgen Receptor Is a Noncoding Driver in Advanced Prostate Cancer. *Cell*, 422-432.e13. <https://doi.org/10.1016/j.cell.2018.05.037>
97. Taylor, B. S., Schultz, N., Hieronymus, H., Gopalan, A., Xiao, Y., Carver, B. S., Arora, V. K., Kaushik, P., Cerami, E., Reva, B., Antipin, Y., Mitsiades, N., Landers, T., Dolgalev, I., Major, J. E., Wilson, M., Socci, N. D., Lash, A. E., Heguy, A., ... Gerald, W. L. (2010). Integrative Genomic Profiling of Human Prostate Cancer. *Cancer Cell*, 11–22. <https://doi.org/10.1016/j.ccr.2010.05.026>
98. Teply, B. A., Wang, H., Lubber, B., Sullivan, R., Rifkind, I., Bruns, A., Spitz, A., DeCarli, M., Sinibaldi, V., Pratz, C. F., Lu, C., Silberstein, J. L., Luo, J., Schweizer, M. T., Drake, C. G., Carducci, M. A., Paller, C. J., Antonarakis, E. S., Eisenberger, M. A., & Denmeade, S. R. (2018). Bipolar androgen therapy in men with metastatic castration-resistant prostate cancer after progression on enzalutamide: an open-label, phase 2, multicohort study. *The Lancet Oncology*, 76–86. [https://doi.org/10.1016/s1470-2045\(17\)30906-3](https://doi.org/10.1016/s1470-2045(17)30906-3)
99. Tewari, A. K., Yardimci, G., Shibata, Y., Sheffield, N. C., Song, L., Taylor, B. S., Georgiev, S. G., Coetzee, G. A., Ohler, U., Furey, T. S., Crawford, G. E., & Febbo, P. G. (2012). Chromatin accessibility reveals insights into androgen receptor activation and transcriptional specificity. *Genome Biology*, R88. <https://doi.org/10.1186/gb-2012-13-10-r88>
100. Than, J. K., Rodriguez, K., & Khera, M. (2018). Post-finasteride Syndrome: A Review of Current Literature. *Current Sexual Health Reports*, 152–157. <https://doi.org/10.1007/s11930-018-0163-4>
101. Theoret, M. R., Ning, Y.-M., Zhang, J. J., Justice, R., Keegan, P., & Pazdur, R. (2011). The Risks and Benefits of 5 α -Reductase Inhibitors for Prostate-Cancer Prevention. *New England Journal of Medicine*, 97–99. <https://doi.org/10.1056/nejmp1106783>
102. Todd, T. W., & Lim, J. (2013). Aggregation formation in the polyglutamine diseases: Protection at a cost? *Molecules and Cells*, 185–194. <https://doi.org/10.1007/s10059-013-0167-x>
103. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>

104. Traish, A. M., Mulgaonkar, A., & Giordano, N. (2014). The Dark Side of 5 α -Reductase Inhibitors' Therapy: Sexual Dysfunction, High Gleason Grade Prostate Cancer and Depression. *Korean Journal of Urology*, 367. <https://doi.org/10.4111/kju.2014.55.6.367>
105. Urbanucci, A, Sahu, B., Seppälä, J., Larjo, A., Latonen, L. M., Waltering, K. K., Tammela, T. L. J., Vessella, R. L., Lähdesmäki, H., Jänne, O. A., & Visakorpi, T. (2011). Overexpression of androgen receptor enhances the binding of the receptor to the chromatin in prostate cancer. *Oncogene*, 2153–2163. <https://doi.org/10.1038/onc.2011.401>
106. Urbanucci, Alfonso, Barfeld, S. J., Kytölä, V., Ikonen, H. M., Coleman, I. M., Vodák, D., Sjöblom, L., Sheng, X., Tolonen, T., Minner, S., Burdelski, C., Kivinummi, K. K., Kohvakka, A., Kregel, S., Takhar, M., Alshalalfa, M., Davicioni, E., Erho, N., Lloyd, P., ... Mills, I. G. (2017). Androgen Receptor Deregulation Drives Bromodomain-Mediated Chromatin Alterations in Prostate Cancer. *Cell Reports*, 2045–2059. <https://doi.org/10.1016/j.celrep.2017.05.049>
107. Urbanucci, Alfonso, Marttila, S., Jänne, O. A., & Visakorpi, T. (2011). Androgen receptor overexpression alters binding dynamics of the receptor to chromatin and chromatin structure. *The Prostate*, 1223–1232. <https://doi.org/10.1002/pros.22473>
108. Urbanucci, Alfonso, Waltering, K. K., Suikki, H. E., Helenius, M. A., & Visakorpi, T. (2008). Androgen regulation of the androgen receptor coregulators. *BMC Cancer*. <https://doi.org/10.1186/1471-2407-8-219>
109. Van Neste, D. (2019). Maintenance of optimised hair growth from viable terminal scalp hair follicles at baseline with oral finasteride in male pattern hair loss and first evidence of a “drug dependency” and a post-finasteride “rebound effect.” *Skin Research and Technology*, 712–719. <https://doi.org/10.1111/srt.12707>
110. Van Rompay, M. I., Curtis Nickel, J., Ranganathan, G., Kantoff, P. W., Solomon, K. R., Lund, J. L., & McKinlay, J. B. (2018). Impact of 5 α -reductase inhibitor and β -blocker therapy for benign prostatic hyperplasia on prostate cancer incidence and mortality. *BJU International*, 511–518. <https://doi.org/10.1111/bju.14534>
111. Visakorpi, T., Hyytinen, E., Koivisto, P., Tanner, M., Keinänen, R., Palmberg, C., Palotie, A., Tammela, T., Isola, J., & Kallioniemi, O.-P. (1995). In vivo amplification of the androgen receptor gene and progression of human prostate cancer. *Nature Genetics*, 401–406. <https://doi.org/10.1038/ng0495-401>

112. Waltering, K. K., Helenius, M. A., Sahu, B., Manni, V., Linja, M. J., Janne, O. A., & Visakorpi, T. (2009). Increased Expression of Androgen Receptor Sensitizes Prostate Cancer Cells to Low Levels of Androgens. *Cancer Research*, 8141–8149. <https://doi.org/10.1158/0008-5472.can-09-0919>
 113. Xing, N. (2001). Quercetin inhibits the expression and function of the androgen receptor in LNCaP prostate cancer cells. *Carcinogenesis*, 409–414. <https://doi.org/10.1093/carcin/22.3.409>
 114. Yu, J., Gritsina, G., & Gao, W.-Q. (2019). Transcriptional repression by androgen receptor: roles in castration-resistant prostate cancer. *Asian Journal of Andrology*, 215. https://doi.org/10.4103/aja.aja_19_19
 115. Zitzmann, M. (2009). Pharmacogenetics of testosterone replacement therapy. *Pharmacogenomics*, 1341–1349. <https://doi.org/10.2217/pgs.09.58>
-

The role of the AR in areas relevant to the sexual dysfunction in PFS

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/the-role-of-the-ar-in-areas-relevant-to-the-sexual-dysfunction-in-pfs/>

Libido, erectile function, and penile structural maintenance

Male libido and sexual desire is primarily androgen-mediated and strictly testosterone dependent. Considerable evidence supports libido loss as the clearest symptom of hypogonadism (Santi et al., 2018). Evidence regarding the role of other hormones is less clear (Corona et al., 2016). Male-typical behaviour requires AR signaling in adults, and AR inactivation in male mice causes a complete loss of male sexual behaviour alongside a significant reduction in aggression (Sato et al., 2004).

Phosphorylated endothelial nitric oxide synthase (eNOS) has a key facilitative role in physiological penile erection following initiation by neuronal nitric oxide synthase (nNOS) (Burnett, 2004). In human aortic endothelium cells, T rapidly induces eNOS activation and production of nitric oxide through AR-dependent induction of PI3-kinase/Akt signaling (Yu et al., 2010). Additionally, AR inactivation in mice demonstrates a dramatic reduction in nNOS expression in the hypothalamus, suggesting AR regulation of this neurotransmitter and its sexually relevant functions (Sato et al., 2004). A particularly common and important symptom of PFS is the loss of nocturnal and morning erections. This is a central mechanism of unconscious sexual arousability (Santi et al., 2018). Inactivation of the noradrenergic cells in the locus coeruleus in the brain stem, a site expressive of the AR, is a testosterone-dependant process (Bancroft, 2005) that results in nocturnal penile tumescence.

Androgens are crucial to maintain male reproductive physiology and erectile function and are critical to the integrity and maintenance of the tunica albuginea, cavernous endothelium, cavernosal smooth muscle, and nerve structure and function (A. Traish & Kim, 2005; A. M. Traish, 2008; Zhang et al., 2013). Tissue integrity and structure is vital to venoocclusive function, and structural alteration will result in dysfunction that is both difficult to diagnose and challenging to treat (A. M. Traish, 2008). Castration of adult male rats significantly decreases penile length, girth, smooth muscle content and endothelial nitric oxide synthase activity, and this is reversible with testosterone administration (Halmenschlager et al., 2017; Hofer et al., 2015; Huh et al., 2018; A. M. Traish, 2008). Immunohistochemical study of stromal and endothelial human corpus cavernosum cells biopsied from potent males aged between 19 and 63 revealed age-independent high expression of the AR (74.9%) and low expression of ERα (11%). Cultured endothelial cells exposed to T or DHT showed dose-dependent and significant increases in

cellular metabolic activity than control groups with or without growth medium, while similar concentrations of estradiol or progesterone had no respective effect compared with controls (Schultheiss et al., 2003). This comparably reflects the testosterone-stimulated increase in proliferation reported in fetal smooth muscle cells (Crescioli et al., 2003), suggesting that peripheral androgen receptor function is important for maintenance of physiology and function of the endothelium in the adult human male penis. The significant expression of AR in penile tissue suggests its vulnerability to a proposed loss of function and potential toxic gain of function conferred by site-specific AR overexpression. This is of particular relevance to the progressive and often rapid penile atrophy after cessation of the drug experienced by some PFS patients that can occur after only one dose (Garreton et al., 2016). It is particularly relevant that PFS patients reporting this can have normal serum androgen levels (Irwig, 2014), or frequently relatively high - or intraindividually increased - levels of T.

In addition to venous leakage, clinical findings of calcification and atherosclerosis upon penile ultrasound are anecdotally reported findings following urological evaluation of PFS patients with atrophic changes to the penis. AR signaling is increasingly appreciated as involved in calcification and atherosclerotic lesions, in line with the well-appreciated heightened risk of cardiovascular disease in males, as recently reviewed by Takov et al (Takov et al., 2018). Vascular smooth muscle cells (VSMCs) provide structural integrity of blood vessels and control diameter via regulation of contraction and vasodilation (Metz et al., 2011). Zhu et al. reported significant expression of the AR in VSMCs and the presence of AR in calcified aortic and femoral artery tissue. In vitro investigation revealed striking induction of pro-calcificatory effects by both testosterone and DHT, and the lack of aromatase expression in these cells indicated direct mediation by AR signaling (Zhu et al., 2016). Arterial calcification and atherosclerosis has been associated with long term anabolic steroid abuse, hyperandrogenemia in women with polycystic ovary syndrome and postmenopausal women administered testosterone (Christian et al., 2003; Hak et al., 2007; Santora et al., 2006). However, in addition to pro-calcificatory effects, investigations have revealed that androgen induction of AR-mediated processes are atheroprotective (Son et al., 2010; Yu et al., 2010), further suggesting appropriate AR-mediated signaling is necessary for vascular health.

Testosterone treatment of rats during urethral wound healing increases myofibroblast proliferation and collagen deposition, and Hofer et al. speculate this may contribute to spongiobrosis and stricture development (Hofer et al., 2015). Finasteride has recently been suggested as a potential therapy in myocardial infarction. Evidence of increased DHT and androgen-responsive gene expression in mouse models of myocardial infarction was reported, and treatment with finasteride markedly improved cardiac function and reduced fibroblast collagen secretion (Froese et al., 2018). Interestingly, prominent collagen deposition is reported in the corpus cavernosum of rats treated with either finasteride or dutasteride (Sahin Kilic et al., 2018), reflecting androgen deprivation and hypogonadism (El-Sakka, 2011; A. Traish & Kim, 2005), possibly indicative of a similar histopathological effect of both reduced or increased androgen signaling.

Hypospadias, a congenital penile deformation associated with prenatal endocrine disruption (Wolf et al.,

1999)? and decreased androgen signaling (Aschim et al., 2004)?, is associated with altered expression of the AR (Vottero et al., 2011)?. Loss of AR expression is not correlated to severity (Celayir, 2018)?. Interestingly, Qiao et al. reported that AR was significantly upregulated in the preputial skin of boys with severe hypospadias compared with boys without hypospadias or boys with mild hypospadias, the latter demonstrating a more moderate elevation in AR expression (Qiao et al., 2012)?.

Sperm count, motility, and semen consistency

AR dysregulation is a plausible causative factor for well-reported changes to sperm count, motility, semen consistency and ejaculate volume in PFS. AR action in the male reproductive system is functionally critical to sperm differentiation, maturation and survival. Targeted AR knockout in mice causes azoospermia and infertility (Krutskikh et al., 2011)?. The AR has been recently shown to be critical across the spermatogenesis and maturation processes. Androgen blockade inhibits differentiation to spermatocytes. In vitro cell culture and in vivo confirmations revealed that promyelocytic leukemia zinc-finger, an important gene in differentiation of spermatogonial stem cells. AR in Sertoli cells indirectly regulates β 1 integrin via GATA2 and WT-1, and β 1 integrin further binds to E-cadherin to regulate the fate of spermatogonial stem cells. DHT treatment of AR-overexpressing Sertoli cells demonstrated AR indirectly down-regulates WT-1, a key gene in spermatogenesis, via GATA2 (J. Wang et al., 2019)?. WT-1 is critical to spermatogenesis and deficiency is associated with male infertility (X. N. Wang et al., 2013)?. The human epididymis is a complex tubular structure in which spermatozoa functionally develop and reach maturity, serving as conduit to the vas deferens from the testis (Cornwall, 2008)?. AR is prominently expressed throughout the epididymis (SAR et al., 1990; Zhou et al., 2002)?, and the importance of the AR in this tissue is well established (Robaire & Hamzeh, 2011)?. The critical influence of the AR in human epididymal cells has been confirmed by next generation deep sequencing protocols (Browne et al., 2019)?. The AR has been identified to regulate a functional transcriptional network of about 200 genes in the human caput epididymis epithelium and is therefore critically implicated in sperm maturation and fertility maintenance in men (Yang et al., 2018)?. The vas deferens fluid microenvironment is crucial to sperm transport and survival in the organ. In rats, vas deferens lumen size, fluid volume and osmolality have been demonstrated to be under the regulation of the AR, as was the expression of aquaporin isoforms AQP-1, AQP-2 and AQP-9. Testosterone was shown to increase water secretion and osmolality in this organ through the AR and was interrupted by Finasteride or Flutamide (Ramli et al., 2018)?.

Post-Orgasm illness and increased refractory period

Both a significantly increased refractory period and a post-orgasm modulation of symptoms is widely reported in PFS. Male accessory sex organs are responsive to prolactin. Post-orgasm increases in

prolactin are implicated in sex organ maintenance and functionality, whereas constant levels would prove deleterious (Hernandez et al., 2006). Prolactin administration has been observed to induce a dose-related increase of AR expression levels beyond levels explainable by organ weight increases in the testes, prostate and epididymis in male rats (BARAÑAO et al., 1982).

In male rats, AR mRNA levels in the ventral prostate were determined after consecutive ejaculations by Hernandez et al. AR, with a concurrent steady increase in AR mRNA, was significantly increased after one ejaculation (100% increase; $p < 0.05$). Levels were further highly increased after two and three ejaculations (200% and 300% increases respectively) to a total of 800%, returning to precopulatory levels rapidly after the fourth ejaculation. Interestingly, a rapid and significant copulation-induced increase in androgen receptor protein precedes higher expression of mRNA or serum elevation of testosterone, suggesting rapid regulatory processes. Additionally, testosterone reaching its maximal increase did not arrest the continual increase of AR mRNA, suggesting the existence of a balance of both gene transcription and stabilization in regards to AR-mRNA levels (Hernandez et al., 2007).

The paracrine influence of oxytocin (H. Nicholson, 1996), which systemically increases at orgasm (Ivell et al., 1997; OGAWA et al., 1980; Thackare et al., 2006), may be influential in the commonly reported modulation of PFS symptoms following orgasm, which can include severe multi-symptom worsening usually lasting a number of days. Administration of oxytocin to rats has been shown to increase testicular and epididymal weight, with a significant increase in 5 α reductase activity in these organs ($P < 0.005$ and $P < 0.01$ respectively). In vitro homogenates incubated with oxytocin additionally showed significant increases in 5 α reductase activity at low concentrations (10 pg/0.3-mg protein) (H. D. Nicholson & Jenkin, 1994). Oxytocin at physiological levels positively regulates the activity of type I and II 5 α reductases in human prostate epithelial cells (S.J. Assinder, 2007) and in LNCAP cell lines (Stephen J. Assinder et al., 2015). These results occurred at the level of post-translational protein activity and do not appear to regulate gene expression. Oxytocin is considered to be a potent growth inducer in prostate cancer (Xu et al., 2017). We hypothesise the increase in 5 α -reductase activity and increased androgen receptor expression may explain a significant and widely anecdotally reported impact of orgasm on PFS symptoms.

Page Bibliography

1. Aschim, E. L., Nordenskjöld, A., Giwercman, A., Lundin, K. B., Ruhayel, Y., Haugen, T. B., Grotmol, T., & Giwercman, Y. L. (2004). Linkage between Cryptorchidism, Hypospadias, and GGN Repeat Length in the Androgen Receptor Gene. *The Journal of Clinical Endocrinology & Metabolism*, 5105–5109. <https://doi.org/10.1210/jc.2004-0293>
2. Assinder, S.J. (2007). Oxytocin increases 5 α -reductase activity of human prostate epithelial cells,

- but not stromal cells. *The Prostate*, 115–121. <https://doi.org/10.1002/pros.20671>
3. Assinder, Stephen J., Davies, K., Suriya, J., & Liu-Fu, F. (2015). Oxytocin differentially effects 3 β -hydroxysteroid dehydrogenase and 5 α -reductase activities in prostate cancer cell lines. *Peptides*, 149–155. <https://doi.org/10.1016/j.peptides.2015.07.015>
 4. Bancroft, J. (2005). The endocrinology of sexual arousal. *Journal of Endocrinology*, 411–427. <https://doi.org/10.1677/joe.1.06233>
 5. BARAÑAO, J. S., TESONE, M., OLIVEIRA-FILHO, R. M., CHIAUZZI, V. A., CALVO, J. C., CHARREAU, E. H., & CALANDRA, R. S. (1982). Effects of Prolactin on Prostate Androgen Receptors in Male Rats. *Journal of Andrology*, 281–288. <https://doi.org/10.1002/j.1939-4640.1982.tb00684.x>
 6. Browne, J. A., Leir, S., Yin, S., & Harris, A. (2019). Transcriptional networks in the human epididymis. *Andrology*. <https://doi.org/10.1111/andr.12629>
 7. Burnett, A. L. (2004). Novel nitric oxide signaling mechanisms regulate the erectile response. *International Journal of Impotence Research*, S15–S19. <https://doi.org/10.1038/sj.ijir.3901209>
 8. Celayir, A. (2018). Expression of androgen, estrogen and progesterone hormone receptors in penile tissues of children with different types of hypospadias. *Northern Clinics of Istanbul*. <https://doi.org/10.14744/nci.2018.47108>
 9. Christian, R. C., Dumesic, D. A., Behrenbeck, T., Oberg, A. L., Sheedy, P. F., II, & Fitzpatrick, L. A. (2003). Prevalence and Predictors of Coronary Artery Calcification in Women with Polycystic Ovary Syndrome. *The Journal of Clinical Endocrinology & Metabolism*, 2562–2568. <https://doi.org/10.1210/jc.2003-030334>
 10. Cornwall, G. A. (2008). New insights into epididymal biology and function. *Human Reproduction Update*, 213–227. <https://doi.org/10.1093/humupd/dmn055>
 11. Corona, G., Isidori, A. M., Aversa, A., Burnett, A. L., & Maggi, M. (2016). Endocrinologic Control of Men's Sexual Desire and Arousal/Erection. *The Journal of Sexual Medicine*, 317–337. <https://doi.org/10.1016/j.jsxm.2016.01.007>

12. Crescioli, C., Maggi, M., Vannelli, G. B., Ferruzzi, P., Granchi, S., Mancina, R., Muratori, M., Forti, G., Serio, M., & Luconi, M. (2003). Expression of Functional Estrogen Receptors in Human Fetal Male External Genitalia. *The Journal of Clinical Endocrinology & Metabolism*, 1815–1824. <https://doi.org/10.1210/jc.2002-021085>

13. El-Sakka, A. I. (2011). Reversion of penile fibrosis: Current information and a new horizon. *Arab Journal of Urology*, 49–55. <https://doi.org/10.1016/j.aju.2011.03.013>

14. Froese, N., Wang, H., Zwadlo, C., Wang, Y., Grund, A., Gigina, A., Hofmann, M., Kilian, K., Scharf, G., Korf-Klingebiel, M., Melchert, A., Signorini, M. E. R., Halloin, C., Zweigerdt, R., Martin, U., Gruh, I., Wollert, K. C., Geffers, R., Bauersachs, J., & Heineke, J. (2018). Anti-androgenic therapy with finasteride improves cardiac function, attenuates remodeling and reverts pathologic gene-expression after myocardial infarction in mice. *Journal of Molecular and Cellular Cardiology*, 114–124. <https://doi.org/10.1016/j.yjmcc.2018.08.011>

15. Garreton, A. S., Valzacchi, G. R., & Layus, O. (2016). Post-Finasteride Syndrome: About 2 Cases and Review of the Literature. *Andrology-Open Access*. <https://doi.org/10.4172/2472-1212.1000170>

16. Hak, A. E., Westendorp, I. C. D., Pols, H. A. P., Hofman, A., & Witteman, J. C. M. (2007). High-dose testosterone is associated with atherosclerosis in postmenopausal women. *Maturitas*, 153–160. <https://doi.org/10.1016/j.maturitas.2006.07.004>

17. Halmenschlager, G., Rhoden, E. L., Motta, G. A., Sagrillo Fagundes, L., Medeiros, J. L., Jr., Meurer, R., & Rhoden, C. R. (2017). Testosterone replacement maintains smooth muscle content in the corpus cavernosum of orchietomized rats. *Asian Journal of Urology*, 223–229. <https://doi.org/10.1016/j.ajur.2017.02.001>

18. Hernandez, M. E., Soto-Cid, A., Aranda-Abreu, G. E., Díaz, R., Rojas, F., Garcia, L. I., Toledo, R., & Manzo, J. (2007). A study of the prostate, androgens and sexual activity of male rats. *Reproductive Biology and Endocrinology*. <https://doi.org/10.1186/1477-7827-5-11>

19. Hernandez, M. E., Soto-Cid, A., Rojas, F., Pascual, L. I., Aranda-Abreu, G. E., Toledo, R., Garcia, L. I., Quintanar-Stephano, A., & Manzo, J. (2006). Prostate response to prolactin in sexually active male rats. *Reproductive Biology and Endocrinology*. <https://doi.org/10.1186/1477-7827-4-28>

20. Hofer, M. D., Cheng, E. Y., Bury, M. I., Xu, W., Hong, S. J., Kaplan, W. E., & Sharma, A. K.

- (2015). Androgen Supplementation in Rats Increases the Inflammatory Response and Prolongs Urethral Healing. *Urology*, 691–697. <https://doi.org/10.1016/j.urology.2014.11.025>
21. Huh, J. S., Chung, B. H., Hong, C. H., Ryu, J. K., Kim, J. H., Han, W. K., & Park, K. K. (2018). The effects of testosterone replacement on penile structure and erectile function after long-term castration in adult male rats. *International Journal of Impotence Research*, 122–128. <https://doi.org/10.1038/s41443-017-0010-6>
22. Irwig, M. S. (2014). Androgen Levels and Semen Parameters Among Former Users of Finasteride With Persistent Sexual Adverse Effects. *JAMA Dermatology*, 1361. <https://doi.org/10.1001/jamadermatol.2014.1830>
23. Ivell, R., Balvers, M., Rust, W., Bathgate, R., & Einspanier, A. (1997). Oxytocin and Male Reproductive Function. In *Advances in Experimental Medicine and Biology* (pp. 253–264). Springer US. https://doi.org/10.1007/978-1-4615-5913-9_47
24. Krutskikh, A., De Gendt, K., Sharp, V., Verhoeven, G., Poutanen, M., & Huhtaniemi, I. (2011). Targeted Inactivation of the Androgen Receptor Gene in Murine Proximal Epididymis Causes Epithelial Hypotrophy and Obstructive Azoospermia. *Endocrinology*, 689–696. <https://doi.org/10.1210/en.2010-0768>
25. Metz, R. P., Patterson, J. L., & Wilson, E. (2011). Vascular Smooth Muscle Cells: Isolation, Culture, and Characterization. In *Methods in Molecular Biology* (pp. 169–176). Humana Press. https://doi.org/10.1007/978-1-61779-523-7_16
26. Nicholson, H. (1996). Oxytocin: a paracrine regulator of prostatic function. *Reviews of Reproduction*, 69–72. <https://doi.org/10.1530/ror.0.0010069>
27. Nicholson, H. D., & Jenkin, L. (1994). 5 α -Reductase Activity Increased by Oxytocin in the Rat Testis. In *Function of Somatic Cells in the Testis* (pp. 278–285). Springer New York. https://doi.org/10.1007/978-1-4612-2638-3_18
28. OGAWA, S., KUDO, S., KITSUNAI, Y., & FUKUCHI, S. (1980). INCREASE IN OXYTOCIN SECRETION AT EJACULATION IN MALE. *Clinical Endocrinology*, 95–97. <https://doi.org/10.1111/j.1365-2265.1980.tb01027.x>
29. Qiao, L., Tasian, G. E., Zhang, H., Cao, M., Ferretti, M., Cunha, G. R., & Baskin, L. S. (2012).

Androgen receptor is overexpressed in boys with severe hypospadias, and ZEB1 regulates androgen receptor expression in human foreskin cells. *Pediatric Research*, 393–398.

<https://doi.org/10.1038/pr.2011.49>

30. Ramli, N. S. K., Giribabu, N., Karim, K., & Salleh, N. (2018). Hormonal control of vas deferens fluid volume and aquaporin expression in rats. *Journal of Molecular Histology*, 21–34. <https://doi.org/10.1007/s10735-018-9804-1>
31. Robaire, B., & Hamzeh, M. (2011). Androgen Action in the Epididymis. *Journal of Andrology*, 592–599. <https://doi.org/10.2164/jandrol.111.014266>
32. Sahin Kilic, Engin Kolukcu, Fikret Erdemir, Ismail Benli, & Akgul Arici. (2018). The Effects of Oral 5-alpha Reductase Inhibitors on Penile Intracavernosal Pressures and Penile Morphology in Rat Model. *Urology Journal*. <https://doi.org/10.22037/uj.v0i0.4164>
33. Santi, D., Spaggiari, G., Gilioli, L., Potì, F., Simoni, M., & Casarini, L. (2018). Molecular basis of androgen action on human sexual desire. *Molecular and Cellular Endocrinology*, 31–41. <https://doi.org/10.1016/j.mce.2017.09.007>
34. Santora, L. J., Marin, J., Vangrow, J., Minegar, C., Robinson, M., Mora, J., & Frieds, G. (2006). Coronary Calcification in Body Builders Using Anabolic Steroids. *Preventive Cardiology*, 198–201. <https://doi.org/10.1111/j.1559-4564.2006.05210.x>
35. SAR, M., LUBAHN, D. B., FRENCH, F. S., & WILSON, E. M. (1990). Immunohistochemical Localization of the Androgen Receptor in Rat and Human Tissues*. *Endocrinology*, 3180–3186. <https://doi.org/10.1210/endo-127-6-3180>
36. Sato, T., Matsumoto, T., Kawano, H., Watanabe, T., Uematsu, Y., Sekine, K., Fukuda, T., Aihara, K. -i., Krust, A., Yamada, T., Nakamichi, Y., Yamamoto, Y., Nakamura, T., Yoshimura, K., Yoshizawa, T., Metzger, D., Chambon, P., & Kato, S. (2004). Brain masculinization requires androgen receptor function. *Proceedings of the National Academy of Sciences*, 1673–1678. <https://doi.org/10.1073/pnas.0305303101>
37. Schultheiss, D., Badalyan, R., Pilatz, A., Gabouev, A. I., Schlote, N., Wefer, J., von Wasielewski, R., Mertsching, H., Sohn, M., Stief, C. G., & Jonas, U. (2003). Androgen and estrogen receptors in the human corpus cavernosum penis: immunohistochemical and cell culture results. *World Journal of Urology*, 320–324. <https://doi.org/10.1007/s00345-003-0371-y>

38. Son, B.-K., Akishita, M., Iijima, K., Ogawa, S., Maemura, K., Yu, J., Takeyama, K., Kato, S., Eto, M., & Ouchi, Y. (2010). Androgen Receptor-dependent Transactivation of Growth Arrest-specific Gene 6 Mediates Inhibitory Effects of Testosterone on Vascular Calcification. *Journal of Biological Chemistry*, 7537–7544. <https://doi.org/10.1074/jbc.m109.055087>
39. Takov, K., Wu, J., Denvir, M. A., Smith, L. B., & Hadoke, P. W. F. (2018). The role of androgen receptors in atherosclerosis. *Molecular and Cellular Endocrinology*, 82–91. <https://doi.org/10.1016/j.mce.2017.10.006>
40. Thackare, H., Nicholson, H. D., & Whittington, K. (2006). Oxytocin—its role in male reproduction and new potential therapeutic uses. *Human Reproduction Update*, 437–448. <https://doi.org/10.1093/humupd/dmk002>
41. Traish, A., & Kim, N. (2005). ORIGINAL RESEARCH—ENDOCRINOLOGY: The Physiological Role of Androgens in Penile Erection: Regulation of Corpus Cavernosum Structure and Function. *The Journal of Sexual Medicine*, 759–770. <https://doi.org/10.1111/j.1743-6109.2005.00094.x>
42. Traish, A. M. (2008). Androgens Play a Pivotal Role in Maintaining Penile Tissue Architecture and Erection: A Review. *Journal of Andrology*, 363–369. <https://doi.org/10.2164/jandrol.108.006007>
43. Vottero, A., Minari, R., Viani, I., Tassi, F., Bonatti, F., Neri, T. M., Bertolini, L., Bernasconi, S., & Ghizzoni, L. (2011). Evidence for Epigenetic Abnormalities of the Androgen Receptor Gene in Foreskin from Children with Hypospadias. *The Journal of Clinical Endocrinology & Metabolism*, E1953–E1962. <https://doi.org/10.1210/jc.2011-0511>
44. Wang, J., Li, J., Xu, W., Xia, Q., Gu, Y., Song, W., Zhang, X., Yang, Y., Wang, W., Li, H., & Zou, K. (2019). Androgen promotes differentiation of PLZF+ spermatogonia pool via indirect regulatory pattern. *Cell Communication and Signaling*. <https://doi.org/10.1186/s12964-019-0369-8>
45. Wang, X. N., Li, Z. S., Ren, Y., Jiang, T., Wang, Y. Q., Chen, M., Zhang, J., Hao, J. X., Wang, Y. B., Sha, R. N., Huang, Y., Liu, X., Hu, J. C., Sun, G. Q., Li, H. G., Xiong, C. L., Xie, J., Jiang, Z. M., Cai, Z. M., ... Gao, F. (2013). The Wilms Tumor Gene, Wt1, Is Critical for Mouse Spermatogenesis via Regulation of Sertoli Cell Polarity and Is Associated with Non-Obstructive Azoospermia in Humans. *PLoS Genetics*, e1003645. <https://doi.org/10.1371/journal.pgen.1003645>

46. Wolf, C., Lambright, C., Mann, P., Price, M., Cooper, R. L., Ostby, J., & Gray, L. E., Jr. (1999). Administration of potentially antiandrogenic pesticides (procymidone, linuron, iprodione, chlozolinate, p,p'-DDE, and ketoconazole) and toxic substances (dibutyl- and diethylhexyl phthalate, PCB 169, and ethane dimethane sulphonate) during sexual differentiation produces diverse profiles of reproductive malformations in the male rat. *Toxicology and Industrial Health*, 94–118. <https://doi.org/10.1177/074823379901500109>
47. Xu, H., Fu, S., Chen, Q., Gu, M., Zhou, J., Liu, C., Chen, Y., & Wang, Z. (2017). The function of oxytocin: a potential biomarker for prostate cancer diagnosis and promoter of prostate cancer. *Oncotarget*. <https://doi.org/10.18632/oncotarget.16107>
48. Yang, R., Browne, J. A., Eggener, S. E., Leir, S.-H., & Harris, A. (2018). A novel transcriptional network for the androgen receptor in human epididymis epithelial cells. *MHR: Basic Science of Reproductive Medicine*, 433–443. <https://doi.org/10.1093/molehr/gay029>
49. Yu, J., Akishita, M., Eto, M., Ogawa, S., Son, B.-K., Kato, S., Ouchi, Y., & Okabe, T. (2010). Androgen Receptor-Dependent Activation of Endothelial Nitric Oxide Synthase in Vascular Endothelial Cells: Role of Phosphatidylinositol 3-Kinase/Akt Pathway. *Endocrinology*, 1822–1828. <https://doi.org/10.1210/en.2009-1048>
50. Zhang, M.-G., Wang, X.-J., Shen, Z.-J., & Gao, P.-J. (2013). Long-term Oral Administration of 5 α -reductase Inhibitor Attenuates Erectile Function by Inhibiting Autophagy and Promoting Apoptosis of Smooth Muscle Cells in Corpus Caverosum of Aged Rats. *Urology*, 743.e9-743.e15. <https://doi.org/10.1016/j.urology.2013.02.045>
51. Zhou, Q., Nie, R., Prins, G., Saunders, P., Katzenellenbogen, B., & Hess, R. (2002). Localization of androgen and estrogen receptors in adult male mouse reproductive tract. *Journal of Andrology*, 23(6), 870–881. <https://www.ncbi.nlm.nih.gov/pubmed/12399534>
52. Zhu, D., Hadoke, P. W. F., Wu, J., Vesey, A. T., Lerman, Daniel. A., Dweck, M. R., Newby, D. E., Smith, L. B., & MacRae, V. E. (2016). Ablation of the androgen receptor from vascular smooth muscle cells demonstrates a role for testosterone in vascular calcification. *Scientific Reports*. <https://doi.org/10.1038/srep24807>
-

The role of the AR in areas relevant to the physiological symptoms of PFS

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/the-role-of-the-ar-in-areas-relevant-to-the-physiological-symptoms-of-pfs/>

Muscle atrophy and muscular dysfunction

Evidence regarding the ligand- and dose-dependent atrophic consequences of AR overexpression in muscle have been discussed in a previous section. Muscle AR is a major determinant of muscle mass and function. Owing to this, selective androgen receptor modulators are in development with a focus on therapeutic application to diseases including muscle wasting and cachexia (Narayanan et al., 2018; Srinath & Dobs, 2014). In mice with AR knockout in satellite cells, the precursor cells of skeletal muscle, limb maximal grip strength is decreased by 7% despite similar mass, with altered fiber-type distribution observed in soleus muscles. The weight of the perineal LABC muscle is markedly reduced, weighing 52 percent less than control animals (Dubois et al., 2014). Significant levator ani weight reduction occurs in inducible ARKO mice in adulthood independent of earlier AR expression (Wu et al., 2019). It is well appreciated that both the innervating lower motor neurons and the skeletal muscle of the LABC are exquisitely sensitive to androgens (Z. Yu, 2006), highly expressive of AR (D. Ashley Monks & Holmes, 2017; Narayanan et al., 2018) and androgen dependent for survival and function (N. Forger et al., 1993; N. G. Forger et al., 1992; Johansen et al., 2007; C. Jordan et al., 1997; C. L. Jordan et al., 1991; Douglas Ashley Monks et al., 2004; Schröder, 1980; J. Xu et al., 2001). In female ovariectomized mice with consequently diminished pelvic muscles, two SARMs restored pelvic floor muscles to sham operated control weights, with a nonsignificant trend towards an overall increase in lean body mass (Ponnusamy et al., 2017). As a key site for histological analysis of AR-mediated toxicity (Nath et al., 2018), the pelvic floor muscle area is a promising site for biopsy and gene expression assay in PFS patients, as well as for less invasive study including EMG evaluation.

Defects in excitation contraction coupling and intracellular calcium homeostasis of skeletal muscle result in a wide range of myopathies including weakness, myalgia, cramping, muscle wasting, joint stiffness and exercise intolerance (Dowling et al., 2014). The AR is an important regulator of genes involved in muscle contraction, function, structure and calcium dynamics (Chivet et al., 2019). Using a computational biology approach, Chivet et al. identified androgen response elements in the enhancers, promoters, and 5'-untranslated regions of excitation-contraction coupling-related genes found to be dysregulated in their transcriptome analysis of AR100Q, AR113Q mice and SBMA patients. Restoration was achievable partly with castration and fully with suppression of polyQ AR using antisense oligonucleotides, suggesting a reversibility of the disruption. Importantly, these genes were found to be similarly dysregulated in castrated wild-type mice, establishing the key genes involved in muscle

contraction as being under the regulation of androgen signaling (Chivet et al., 2019)?.

Myostatin is a growth factor that strongly inhibits muscle growth (McPherron et al., 1997)?, and is under the regulation of AR signaling. Dubois et al. reported a >6-fold decrease in myostatin expression in levator ani muscle of satellite ARKO mice, as well as significant downregulation in the gastrocnemius. Additionally, a reduction of myostatin mRNA levels in orchietomised mice could be fully reversed with testosterone or DHT administration, demonstrating that myostatin is androgen regulated (Dubois et al., 2014)?.

However, Mendler et al. reported a strong suppression of myostatin mRNA levels by androgens in the skeletal muscle of young male rats. Considering the presence of ARE on the myostatin gene and the induction of androgen receptor coregulators by myostatin, they speculate that a negative feedback-loop exists between myostatin and androgen pathways (Mendler et al., 2007)?.

Skeletal and dental problems

Bone-related complaints are frequent, and diagnosis of osteopenia and osteoporosis are reported by PFS patients. All aspects of body composition are determined by the actions of sex steroids including in the skeleton. Body composition is generally more robust in men, and the risk of osteoporosis is approximately half that of women (Vanderschueren et al., 2014)?.

Hypogonadal men have lowered bone mineral density that is normalised by exogenous testosterone treatment (Behre et al., 1997)?.

In addition to ER, appropriate AR signaling is independently required for adult bone health and maintenance (J.-F. Chen et al., 2019)?.

AR is ubiquitously expressed in human bone marrow in both sexes (Mantalaris et al., 2001)?.

Detailed tissue specific and global studies of ARKO in bone have revealed a critical regulatory role for androgens in bone health and maintenance on a compartmental basis (Vanderschueren et al., 2014)?.

Men with complete or partial androgen insensitivity syndrome have a reduced final height that is intermediate between ordinary males and females, as well as reduced lumbar spine density that cannot be compensated by estrogen replacement (Danilovic et al., 2006)?.

The dramatic reduction in lumbar bone density in androgen insensitivity syndrome patients is not seen in men with 5alpha reductase type II insufficiency syndrome (Sobel et al., 2006)?.

In Asian men with prostate cancer, 12 months of ADT with either combined GnRH agonist and bicalutamide therapy or GnRH monotherapy induces the same significant loss of bone mineral density (Joung et al., 2017)?.

Collectively, this illustrates a direct role of the AR in human bone maintenance. To account for the potential of the confounding influence on developmental influences in lifelong ARKO models, Wu et al. developed an inducible ARKO model, demonstrating appropriate AR expression in adulthood is crucial for bone maintenance in adult male mice. Both pre and post-pubertal AR inactivation resulted in significant decreases in the mid-diaphyseal cortical area and cortical thickness in the tibia, as well as trabecular bone volume fraction in the metaphyseal region (Wu et al., 2019)?.

The reduced cortical thickness was seen to be a "phenocopy" of previously reported models of lifelong AR inactivation (Almeida et al., 2017)?.

In a transgenic mouse model, Wiren et al. explored the consequences of targeted AR overexpression in differentiated osteoblasts, demonstrating that excess AR signaling results in a significantly negative consequences on bone matrix quality, biomechanical competence, fragility and strength, while reducing turnover and inhibiting osteoblastic formation (Wiren et al., 2008). In line with these findings, Aro et al. locally delivered a SARM via an implanted sustained-release matrix in a rat bone marrow ablation model. Contrary to the stated hypothesis of an anabolic effect on intramedullary osteogenesis, only the lowest dose had a negligible anabolic effect, while all higher doses resulted in a dose-dependent decrease in new bone formation around the implant and the bone/implant contact. This was noted to be reflective of overexpression models (Aro et al., 2015). These findings support the suggestion of Vanderschueren et al that neither too high nor too low AR activity is favourable for bone. Steffens et al. have demonstrated in rats that, as with low levels of testosterone following orchidectomy, supraphysiological doses also increase ligature-induced periodontal bone loss (Joao P. Steffens et al., 2012; Joao Paulo Steffens et al., 2015), plausibly reflecting the curvilinear dose relationship of AR signaling (Gibson et al., 2018).

Tooth loss and gum problems are frequent in PFS patients, with many reports of rapid degeneration of teeth, gum recession and the condition causing the need to undergo gingival grafts. Similarly, significantly affected male patients have reported progressive alterations to the jaw structure after cessation. This is notably reported by two brothers who developed PFS after only two weeks of use. It is therefore again highly significant that several lines of evidence suggest periodontal and gingival tissues, tissues responsible for teeth structure and gum health, are dependent on androgens and specifically AR signaling. AR inhibition has been demonstrated to significantly increase periodontal bone loss and impairs bone repair in female rats and is regulatory of inflammatory markers in gingival tissue (João Paulo Steffens et al., 2018, 2019). Minocycline can stimulate 5 α reductase in gingival tissue, and combinatory administration with finasteride has suggested that some of the anabolic response to minocycline in these tissues are attributed to the AR pathway (Soory & Viridi, 1998). Parkar et al. analysed numerous human periodontal ligament and gingival tissue samples as well as cultured cells for expression of AR. In contrast to ER which was not detected, AR was readily detected in a high proportion of tissue and all fibroblasts, suggesting a high and direct sensitivity to androgens in these tissues with implications for inflammation, connective tissue and bone repair processes (Parkar et al., 1996). AR is also highly expressed in human tooth pulp, with a greater expression in males than females, and is subject to hormonal manipulation in vitro. T was observed to significantly reduce AR content in tooth pulp, while E2 or androstenedione increased AR mRNA. This suggests, as with bone, this tissue is highly androgen responsive (Dale et al., 2002). Wang et al. systematically examined the mandibles of castrated rhesus macaques in prime and old age against those of control animals to determine the impact of low androgens on bone and teeth. A prevalence of periodontitis, significant alveolar bone recession and severe temporomandibular joint osteoarthritis was observed in the old castrates. Faces were indicated to be generally narrower by reduced distance between rami. Cortical bone of the mandibular body and rami was thinner, and molar teeth were slender in castrates. These findings collectively suggest the importance of androgens to development and maintenance of facial structure, skeletal and dental health in macaques (Q. Wang et al., 2015). In addition, androgens exert a significant nociceptive behavioural response and are protective against temporomandibular joint pain in castrated male and female rats, but not sham-operated males. This was demonstrated to be mediated by the AR and is independent of aromatisation to

estrogen or the organisational effects of androgens (Fanton et al., 2017)?.

Metabolic regulation

Androgens and the AR are increasingly appreciated as important regulators of metabolic function through actions across the body, and increasing evidence suggests an important influence on metabolic regulation through actions in neurons in hypothalamic and extra-hypothalamic sites in addition to peripheral tissues (Morford et al., 2018)?. As in broader evidence we have discussed, there appears to be a parabolic nature to androgen signaling in metabolic function, with high and low levels being detrimental in both sexes, although the parabola is shifted far to the right in males (Morford et al., 2018)?. Low androgens and androgen deprivation therapy for prostate cancer increase the risk of type 2 diabetes and obesity in men, and studies in humans and animal models have associated low androgens with hyperglycemia, decreased pancreatic β -cell function, impaired fasting glucose, glucose intolerance, altered lipid profiles and metabolic syndrome (Morford et al., 2018; G. Navarro et al., 2016; I.-C. Yu et al., 2014)?. Central AR knockout in males causes late-onset insulin resistance, glucose intolerance, lipid accumulation in the liver and visceral obesity (I.-C. Yu et al., 2012)?. ARKO also induces leptin resistance (Fan et al., 2008)?. AR CAG repeat length is positively correlated with higher body fat content, increased leptin and hyperinsulinemia in men (Zitzmann et al., 2003)? owing to weaker AR signaling. Interestingly, the risk of type 2 diabetes was recently shown to be 30% greater over 11 years in men receiving either finasteride or dutasteride for BPH, without a difference between the drugs (Wei et al., 2019)?. Excessive androgen signaling is also detrimental to optimum metabolic function in males. Male powerlifters using anabolic steroids have diminished glucose tolerance secondary to insulin resistance when compared with non-steroid using athletes and sedentary weight men (COHEN & HICKMAN, 1987)?. In castrated rats administered high doses of testosterone, insulin resistance was observed, as with the castrated group. Castrated rats administered testosterone at a dosage that restored physiological levels abolished the perturbation of insulin sensitivity induced by castration, suggesting an appropriate "window" of androgen signaling is required for metabolic homeostasis in males (HOLMÄNG & BJÖRNTORP, 1992)?.

Hyperandrogenaemia in women results in metabolic effects strikingly coincident with hypogonadism in men, including predisposition to type 2 diabetes (Escobar-Morreale et al., 2014)?. In female mice fed a representative "Western" diet, chronic DHT administration predisposed subjects to type 2 diabetes due to activation of AR in the hypothalamus, which promoted hepatic insulin resistance. In these mice, increased AR signaling in pancreatic β cells increased mitochondrial oxygen consumption and caused insulin hypersecretion, oxidative injury, and predisposed to secondary β cell failure (G. Navarro et al., 2018)?. RNA-seq has identified a fold change >2 in the expression of 214 genes in AR-deficient islets, and that a third of these are proteins associated with cellular stress and inflammation, indicating a response to injury and emphasising the importance of appropriate AR signaling to β cell health (W. Xu et al., 2017)?. Another study in adult female rats showed hyperinsulinemia due to elevated DHT occurs without alteration in the number or size of pancreatic islets or change in β -cell area. Even though DHT treated

females had higher insulin levels than controls, they exhibited glucose intolerance with elevated plasma glucose. Ins1 was shown to have an ARE-like sequence that bound to AR upon DHT treatment, suggesting functional regulation of insulin by the AR and androgen. Additionally, skeletal muscle Ir², the major utiliser of glucose, was downregulated in this model (Mishra et al., 2018). Independent of obesity, female mice eating a normal diet administered low-dose DHT exhibit impaired whole-body glucose metabolism consisting of glucose intolerance, hepatocyte AR-mediated insulin resistance, impaired gluconeogenic capacity and hyperinsulinemia. This was in addition to observations pertaining to reproductive dysfunction including acyclicity, decreased corpora lutea, and increased atretic follicles that were beyond the scope of the study (Andrisse et al., 2016). Reflective of evidence in animal models, 50-90% of women with PCOS, a condition characterised by pathological hyperandrogenemia, display insulin resistance and glucose intolerance (Morford et al., 2018; W. Xu et al., 2019). Testosterone levels robustly correlate with the degree of insulin resistance and β -cell dysfunction in PCOS (Sahin et al., 2014; W. Xu et al., 2019). The Glucagon-Like Peptide-1 (GLP-1) receptor is widely expressed and also an important contributor to insulin and glucose homeostasis and β -cell proliferation (Bullock et al., 1996). Zhu et al recently demonstrated that GLP-1R expression is under the regulation of androgen signaling, and that this regulation was mediated by the DHT AR complex binding to an AR motif in the Glp1r gene promoter region (Zhu et al., 2019).

Glucocorticoid steroids pleiotropically mediate a number of functions essential for life including stress-related and circadian functions, immune regulation, metabolic and energy regulation including gluconeogenesis, and control of glucose uptake (Kadmiel & Cidlowski, 2013). Spaanderman et al recently demonstrated that androgen receptor signaling strongly influences glucocorticoid receptor signaling in metabolic tissues. AR agonism was demonstrated to potentiate glucocorticoid signaling in white and brown adipocytes in vitro and in vivo, while AR antagonism attenuated GR in white adipose tissue and the liver. 11 β -hydroxysteroid dehydrogenase type 1, critical to glucocorticoid homeostasis, was shown to be AR regulated. They also demonstrated increased glucocorticoid signalling enhanced fat mass and significantly reduced lean mass without significantly altering weight and induced hyperlipidaemia which was attenuated with the antiandrogen enzalutamide (Spaanderman et al., 2019).

Androgens, and appropriate proteomic quantity and status of AR, are crucially important to metabolic function and determinant of many aspects of metabolic health. Therefore, a dysregulated androgen receptor is a plausible mechanistic factor in the metabolic disturbances observed in PFS. Additionally, as recent findings implicate insulin receptor and glucagon-like peptide 1 expression in dopaminergic function and mood disorders (Mansur et al., 2018, 2019), the increasing appreciation of the regulation of androgen signaling upon metabolic systems may have functional relevance to the psychological disturbances in PFS.

Digestive complaints, dysmotility, bile acid synthesis and microbiome

Digestive complaints are frequent in PFS with dysmotility, diarrhoea, constipation, and pale stools well reported. Well appreciated sex differences exist in digestive conditions such as IBS, suggesting an influence of sex hormones (Y. S. Kim & Kim, 2018), and women are generally considered to be more disposed to functional gastroenterological disorders (Houghton et al., 2016). Interestingly, testosterone has been reported to be higher in male IBS patients than controls (B. J. Kim et al., 2008). González-Montelongo et al. demonstrated that the digestive tract is a key target of functionally relevant androgen action owing to the AR-mediated regulatory influence of intestinal smooth muscle transit (María C. González-Montelongo et al., 2010). Calcium sensitization and potentiation of contractile activity in ileal and colonic muscles is rapidly and powerfully induced by androgens at physiological concentrations through a strictly androgen-receptor dependent mechanism (María C. González-Montelongo et al., 2006; María C. González-Montelongo et al., 2010) that induces non-genomic cellular signal cascades. These in turn increase ornithine decarboxylase and intracellular polyamines (María C. González-Montelongo et al., 2013), important modulators of intestinal peristalsis (Sánchez et al., 2017).

Dysregulation of bile acid metabolism can result in malabsorption and hyperbilirubinemia (Chiang, 2013) which is a frequent serum abnormality reported by PFS patients. Aldo-keto reductase family 1 member D1 (AKR1D1), a 4-3-oxosteroid 5-reductase, is required to synthesise bile acid from cholesterol (Chiang, 2013). Upregulation of Peroxisome Proliferator-activated Receptor (PPAR) has been demonstrated to markedly decrease AKR1D1 promoter transactivation and expression in vitro in HepG2 cells and in vivo, disrupting bile acid homeostasis (Valanejad et al., 2018). PPAR also induces glucuronidation of bile acids, making this an important regulator of metabolism (Barbier et al., 2003). PPAR has been demonstrated to be under direct regulation by androgens (Collett et al., 2000; Zhang et al., 2012), and this suggests androgen receptor dysregulation may have functional consequences on bile acid synthesis and metabolism due to crosstalk between these pathways.

Androgen dysregulation has been well demonstrated to induce changes in the microbiome composition, including mice models of hyperandrogenemia, castrated mice and PCa patients undergoing multiple different antiandrogen therapies (Guo et al., 2016; Harada et al., 2016; Sfanos et al., 2018; Sherman et al., 2018). The absence of species does not appear to affect the influence of androgens on composition (Torres et al., 2019). Additionally, the microbiome composition of Finasteride treated rats is shown to differ from control animals (Diviccaro et al., 2019).

Immune system and wound healing

For many patients PFS entails an alteration of immune responses, including intraindividual changes in the incidences of viral infection, fungal infections and the modulation of allergies. Various studies have

highlighted essential androgen regulation of the immune system (Lai, Lai, et al., 2012). Data indicates an extensive role for the AR in haematopoiesis (Mantalaris et al., 2001), and immune cell lines including neutrophils, mast cells, macrophages, B cells, T and Treg cells express AR (W. Chen et al., 2010; Ma et al., 2019; Mantalaris et al., 2001; Viselli et al., 1997; Walecki et al., 2015). Rodent studies have indicated that androgen signaling directly influences differentiation and function of T and B cells, central to the adaptive immune system, and possibly contributes to sex differences in autoimmune disorders (Gubbels Bupp & Jorgensen, 2018). Androgens and the AR have an increasingly appreciated role in thymopoiesis and T cell transcriptional function partly by modulation of thymic epithelial cells and affect thymic size and output (M. A. Brown & Su, 2019). Kadel and Kovats, reviewing the understanding of the regulation of sex hormones and viral immunity, suggest that receptor expression may underlie numbers of and functional regulation of innate immune cells in response to hormones (Kadel & Kovats, 2018). Further, sex differences in epigenetically imprinted regions of open or closed chromatin in hematopoietic stem cells may exist, and the sex-divergent epigenome may be responsive to the sex hormone environment (M. A. Brown & Su, 2019; Kadel & Kovats, 2018).

Neutrophils are significantly the most abundant granulocyte and form an vital part of the innate immune system, responding rapidly through chemotaxis to clear bacterial and fungal infections (Desai & Lionakis, 2018; Lai, Lai, et al., 2012). As well as phagocytic removal of cellular debris and pathogens, neutrophils secrete and scavenge a number of cytokines and chemokines that recruit and activate macrophages and monocytes in resolution of inflammation (Gordon & Taylor, 2005; Jones et al., 2016; Pham, 2006; Rittirsch et al., 2008). In men and women neutrophils strongly express AR at all stages of granulopoiesis from myeloblasts to mature neutrophils (Mantalaris et al., 2001). In humans, neutropenia can occur with antiandrogen treatment (Eaton & Blackmore, 2001; McDonnell & Livingston, 1994) but neutrophil counts decrease more moderately following castration (Chuang et al., 2009).

With both in vivo and in vitro studies, Chuang et al. demonstrated that the AR exerts a direct and profound effect upon which neutrophil homeostasis is critically dependent. AR knockout mice are significantly more susceptible to infection. A 90% reduction of neutrophils is observed in male AR knockout and Tfm mice compared with wild type, while castration results in a less significant neutrophil reduction in blood and bone marrow, reflecting human findings. Exogenous androgens restored neutrophil levels in castrated WT mice, but not Tfm or AR knockout mice. Female mice have normal neutrophil levels in the presence of ten-fold lower androgen levels than males, whereas female AR knockout mice are neutropenic, suggesting a direct importance of the AR rather than androgens. It was further demonstrated that loss of AR results in defects in terminal differentiation of neutrophils, and AR restoration in AR knockout granulocyte-macrophage progenitor cells rescued the neutrophil maturation process. AR was also shown to be significantly important to neutrophil production mechanistically by regulation of granulocyte-colony stimulating factor (G-CSF) signaling. Loss of AR in granulocytes leads to suppression of G-CSF resulting from an increase in protein inhibitor of activated STAT protein 3 (PIAS3) binding to STAT3, which is rescued by AR in a dose-dependent manner, apparently without dependence on androgens. Thus, AR is required for G-CSF induction of ERK activation and consequent proliferation of granulocytes (Chuang et al., 2009). Higher androgen levels have been demonstrated to

impair the bactericidal abilities of neutrophils and increase the expression of anti-inflammatory cytokines IL10 and TGF β 1 in a rat model of bacterial prostate inflammation, prolonging the inflammatory response (Scalerandi et al., 2018)?.

Slowed wound healing is very frequently reported in PFS. As with immune differences, sex differences exist in the speed of cutaneous wound healing, with males healing slower than females (Taylor et al., 2002)?. Higher androgen levels are observed to be inhibitory of cutaneous wound healing (Ashcroft & Mills, 2002; Fimmel & Zouboulis, 2005)?, and DHT is more potently inhibitory of upon re-epithelialization than testosterone (Gilliver et al., 2009)?. In line with findings in dermal wound healing, androgens were demonstrated to prolong healing in castrated rats administered testosterone following urethral surgery. Those administered testosterone had significantly increased neutrophils, higher macrophage counts, significantly higher immunomodulators such as TNF α , TGF β -1, VEGF α and IL-10, a more intense and longer inflammatory phase and an increase in myofibroblast proliferation and collagen tissue deposition in the delayed proliferative phase (Hofer et al., 2015)?. Following prostate resection, both castration (X.-J. Wang et al., 2017)? and finasteride (Ruizhe Zhao et al., 2017)? were seen to speed wound healing and induce re-epithelialization, while DHT enhanced macrophages TNF- α secretion through AR signaling. This extended the inflammatory phase, delaying and weakening the anti-inflammatory stage.

Mechanistic studies have revealed that the AR, and not androgens, are critical to the suppression of wound healing (Lai, Chang, et al., 2012)?. AR knockout males have markedly accelerated wound healing that is not reversed with DHT administration (Lai et al., 2009)?, demonstrating increased reepithelialisation, keratinocyte proliferation and matrix deposition. By contrast, AR knockout does not affect the wound healing rate in female mice (Yiwei Wang et al., 2016)?. In a model of autoimmune myocarditis, AR suppression with the AR degrader ASC-J9 promoted anti-inflammatory cytokines and M2 macrophage polarization via STAT3/SOCS3 regulation, suggesting ASC-J9s potential as a protective therapeutic in inflammatory cardiomyopathy (Ma et al., 2019)?. Local AR antagonists and degraders including ASC-J9 are reported to speed wound healing (Lai et al., 2009; Toraldo et al., 2012; Yiwei Wang et al., 2016)?. While AR has an upregulatory effect on TNF- α and CCR2 expression, suppressing cutaneous wound healing (Lai et al., 2009)?, TNF- α has been shown to increase in ARKO mice (Bourghardt et al., 2010)?. Androgens have been reported to be inhibitory of inflammatory cytokine production after haemorrhagic shock and burns (Lai, Lai, et al., 2012)?. In contrast to the discussed studies, testosterone has been shown to reduce TNF- α and IL-1 β in hypogonadal men (Kalinchenko et al., 2010; Malkin et al., 2004)?. Men and women with rheumatoid arthritis have significantly decreased androgen levels in synovial fluid of inflamed tissue (Cutolo, 2009)?.

Considering the increased inflammatory markers in hypogonadism and the anti-inflammatory influence of testosterone in hypogonadal men, Traish et al. suggest that androgens may be necessary in maintaining inflammatory homeostasis (Traish et al., 2018)?. This would be in agreement with a "bell curve" effect of androgen signaling on cellular homeostasis and consistent with Gibson's description of the new

appreciation of testosterone as a "goldilocks molecule" (Gibson et al., 2018)?.

Dry Eye

Dry eye problems are extremely well reported in anecdotes on our forum and from post-finasteride, Accutane and SSRI patients. Androgens play a direct role in the development of lacrimal gland inflammation and aqueous-deficient dry eye disease (Morthen et al., 2019)?. Androgen deficiency is a major cause of dry eye, and this is particularly prevalent in women following the menopausal decrease in androgen levels (K. Li et al., 2017)?. Androgen administration alleviates dry eye symptoms and increases tear flow in Sjogren syndrome patients, suppresses inflammation in mice models of dry eye, and completely resolves symptoms in dry eye dogs (Morthen et al., 2019)?. Complete androgen insensitivity syndrome causes dry eye, meibomian gland dysfunction, lipid tear film layer instability and decreased mucous levels in humans (Mantelli et al., 2006)?. Finasteride has been used to generate a rat model of androgen deficient dry eye, downregulating the AR, disrupting androgen-influenced inflammatory homeostasis, and significantly increasing levels of the inflammatory cytokines IL-1?, IL-4, IL-6, IL-10, MMP-8, FasL and TNF-? in the lacrimal glands as compared with control rats (K. Li et al., 2017; S. Singh et al., 2014)?.

The androgen receptor mRNA and protein have been identified in epithelial cell nuclei of the human meibomian glands, lacrimal glands, cornea and conjunctiva (Rocha, 2000; Wickham et al., 2000)?. DHT has been demonstrated to significantly regulate the expression of approximately 3,000 genes in immortalized human meibomian gland and conjunctival epithelial cells (Khandelwal et al., 2012)?, including many related to inflammation and mucus production.

The testosterone-induced regulation of numerous immune related gene expressions in the lacrimal tissue of Sjogren syndrome and diabetic mouse models differed considerably, with a significant inflammatory effect of androgens in the diabetic mice model as opposed to the anti-inflammatory response seen in the Sjogren's syndrome model. AR status was hypothesised as a possible mediating "on/off switch" for the microenvironment-dependent response (Morthen et al., 2019)?. Interestingly, hyperandrogenic PCOS patients experience dry eye, tear reduction and meibomian gland dysfunction (Baser et al., 2016; Bonini et al., 2007; Yuksel et al., 2015)?, lending further support to the suggestion that appropriate androgen signaling is required for inflammatory homeostasis. Local AR dysregulation in PFS could underlie the dry eyes and tear-related symptoms reported by patients.

Skin

Skin is an androgen-sensitive organ (Ashcroft & Mills, 2002) and a major target of androgen action. The AR is expressed in human skin fibroblasts, basal cells, sebocytes, pilosebaceous units, sweat gland secretory cells, dermal papilla, and keratinocytes (Alesci & Bornstein, 2000; Pelletier & Ren, 2004). The AR has been shown to have a profound and determinant effect on the collagen content of the skin of the adult mouse in both genders (Markova et al., 2004). Immunohistochemical staining has shown that AR staining intensity and immunoreactivity correlates strongly with the height of the apocrine sweat secretory epithelium (Beier et al., 2004), and as low epithelium is associated with inactivity, this would suggest AR signaling has a direct role in sweat secretion (Ceruti et al., 2018). Androgens have been understood to be a leading factor in acne pathogenesis for nearly a century (J. B. HAMILTON, 1941), and androgen signaling influences both the sebaceous gland activity and inflammation associated with acne (Lai, Chang, et al., 2012). Comprised of sebocytes, the sebaceous gland is an important in production of sebum, the lipids comprising which are important in skin barrier function, water resistance, sun damage and UV resistance, and establishment of the commensal bacterial flora of the skin (Szöllösi et al., 2017). The sebaceous gland is capable of synthesising pregnenolone from cholesterol via p450 side chain cleavage (Thiboutot et al., 2003) as well as metabolising androgens through enzymes including hydroxysteroid dehydrogenases and 5 alpha reductase type 1 (Szöllösi et al., 2017). The proliferative effects of androgens on sebocytes are dependent on the physiological site of localisation (Akamatsu et al., 1992). Recent in vitro investigation has demonstrated differentiation of immature sebocytes is under strong AR regulation, and lipid synthesis and storage is induced by androgens in an AR-dependent process. This was demonstrated to be independent of the presence of serum or other cofactors (Barrault et al., 2015).

Alteration in skin pigmentation and tanning response is very commonly reported in PFS patients and a case of PFS involving significant vitiligo was reported by Motofei et al. (Motofei et al., 2017). Early observations by Hamilton noted a poor tanning response to ultraviolet radiation in castrated men, and testosterone treatment would improve melanisation (J. HAMILTON, 1948). Androgens and the AR are involved in melanocyte biology and function, and melanocytes synthesise DHT (Slominski et al., 2004). Genital skin increases in pigmentation at puberty, and this increase in pigmentation is not seen in hypogonadal men (Köhn et al., 2000).

Cooper et al. reported three cases of myotonic dystrophy - a disease associated with low androgen levels - exhibiting androgen dependent diseases including acne, hidradenitis suppurativa, androgenetic alopecia and keratosis pilaris. They speculated a functional difference in AR may account for the frontal balding in myotonic dystrophy, and that in androgen-mediated conditions, the peripheral response to androgens differs between individuals, mediated by peripheral androgen receptors, with absolute levels of circulating androgens being of limited importance (Cooper et al., 2003).

Mitochondrial function

With broad physiological relevance, AR is an important regulator of overall mitochondrial function and is suggested to impact gene transcription through retrograde signaling (Bajpai et al., 2019). Testosterone had been hypothesised to regulate mitochondrial function owing to prior data including serum levels correlating with oxidative phosphorylation gene expression in skeletal muscle (Pitteloud et al., 2005). In a significant contribution to the understanding of the nonclassical role of the AR, Bajpai et al. demonstrated that the AR contains a mitochondrial localisation sequence and is imported into the mitochondria independent of association with ligand where it localises and regulates multiple processes via signaling cascades. Through a number of studies, they elucidated several roles for the AR in regulation of mitochondria. AR negatively regulates assembly factors of, and destabilises, oxidative phosphorylation supercomplexes. The AR is regulatory of the enzymatic activity of oxidative phosphorylation complexes and a large number of oxidative phosphorylation subunits. The AR regulates mitochondrial protein translation through control of the expression of nuclear ribosomal genes in the mitochondria. AR expression was shown to negatively correlate with mitochondrial DNA content and to TFAM (transcription factor A mitochondrial) protein content, which is regulatory of mitochondrial DNA. Mitochondrial stress was demonstrated to increase expression of the AR and its import into the mitochondria, suggesting an intricate link between both (Bajpai et al., 2019). Taken together, the well demonstrated impact on mitochondrial function would suggest aberrant AR signaling is capable of inducing significant mitochondrial dysfunction, which in turn could result in numerous detrimental effects at the cellular and consequently systemic level. This is of significance to the mechanistic overlap of wild-type gene amplification and polyglutamine expansion (D. A. Monks et al., 2007) with consideration as to the aforementioned implication of mitochondrial dysfunction in SBMA. Beyond an indispensable role in cellular energy production, metabolism, apoptosis and proliferation (van der Blik et al., 2017), mitochondria play a major role in aspects of health and disease (Chakrabarty et al., 2018; Ru?Zhou Zhao et al., 2019) including t-cell and macrophage immune response (Liu & Ho, 2018), neurodegeneration and neuroprotection (Darryll M.A. Oliver & P. Hemachandra Reddy, 2019; P. A. Li et al., 2017), sensorineural hearing loss (Kamogashira et al., 2015), cardiomyopathy (Lorenzo et al., 2013), atherosclerosis (Hulsmans et al., 2012), macular degeneration (E. E. Brown et al., 2018), periodontitis (Y. Chen et al., 2019), non-alcoholic fatty liver disease (Simões et al., 2018), cancer (Higuchi et al., 2005; K. K. Singh & Modica-Napolitano, 2017), and normal aging (Y. Wang & Hekimi, 2015).

LH/T Deregulation

PFS patients often report atypical hormonal profiles, and in cases who had profiles from before exposure to finasteride, a significantly altered hormonal milieu is frequently apparent. Curiously, PFS patients commonly report LH disproportionately low in relation to Testosterone levels, and this has been noted in a studied cohort (Di Loreto, 2011). A feedback loop of hypothalamic gonadotropin-releasing hormone

(GnRH) and subsequent LH release from the pituitary stimulate male testosterone synthesis, which in turn negatively regulates GnRH release by acting on steroid receptors in Kiss1/NKB/Dynorphin (KNDy) neurons (V. M. Navarro et al., 2011; Ruka et al., 2016; Smith et al., 2005)?. Neural ARKO male mice show elevated levels of T (Raskin et al., 2009)?, and evidence from ER α knockout additionally illustrates that the AR plays the primary role in negative-feedback regulation of hypothalamic LH release (Wersinger et al., 1999)?.

Page Bibliography

1. Akamatsu, Hirohiko., Zouboulis, C. C., & Orfanos, C. E. (1992). Control of Human Sebocyte Proliferation In Vitro by Testosterone and 5-Alpha-Dihydrotestosterone Is Dependent on the Localization of the Sebaceous Glands. *Journal of Investigative Dermatology*, 509–511. <https://doi.org/10.1111/1523-1747.ep12616181>
2. Alesci, S., & Bornstein, S. R. (2000). Neuroimmunoregulation of Androgens in the Adrenal Gland and the Skin. *Hormone Research in Paediatrics*, 281–286. <https://doi.org/10.1159/000053272>
3. Almeida, M., Laurent, M. R., Dubois, V., Claessens, F., O'Brien, C. A., Bouillon, R., Vanderschueren, D., & Manolagas, S. C. (2017). Estrogens and Androgens in Skeletal Physiology and Pathophysiology. *Physiological Reviews*, 135–187. <https://doi.org/10.1152/physrev.00033.2015>
4. Andrisse, S., Childress, S., Ma, Y., Billings, K., Chen, Y., Xue, P., Stewart, A., Sonko, M. L., Wolfe, A., & Wu, S. (2016). Low-Dose Dihydrotestosterone Drives Metabolic Dysfunction via Cytosolic and Nuclear Hepatic Androgen Receptor Mechanisms. *Endocrinology*, 531–544. <https://doi.org/10.1210/en.2016-1553>
5. Aro, H., Kulkova, J., Moritz, N., Kähkönen, E., & Mattila, R. (2015). Local delivery of a selective androgen receptor modulator failed as an anabolic agent in a rat bone marrow ablation model. *Acta Orthopaedica*, 86(6), 751–759. <https://doi.org/10.3109/17453674.2015.1074840>
6. Ashcroft, G. S., & Mills, S. J. (2002). Androgen receptor-mediated inhibition of cutaneous wound healing. *Journal of Clinical Investigation*, 615–624. <https://doi.org/10.1172/jci0215704>
7. Bajpai, P., Koc, E., Sonpavde, G., Singh, R., & Singh, K. K. (2019). Mitochondrial localization, import, and mitochondrial function of the androgen receptor. *Journal of Biological Chemistry*, 6621–6634. <https://doi.org/10.1074/jbc.ra118.006727>

8. Barbier, O., Duran-Sandoval, D., Pineda-Torra, I., Kosykh, V., Fruchart, J.-C., & Staels, B. (2003). Peroxisome Proliferator-activated Receptor γ Induces Hepatic Expression of the Human Bile Acid Glucuronidating UDP-glucuronosyltransferase 2B4 Enzyme. *Journal of Biological Chemistry*, 32852–32860. <https://doi.org/10.1074/jbc.m305361200>
9. Barrault, C., Garnier, J., Pedretti, N., Cordier-Dirikoc, S., Ratineau, E., Deguerce, A., & Bernard, F.-X. (2015). Androgens induce sebaceous differentiation in sebocyte cells expressing a stable functional androgen receptor. *The Journal of Steroid Biochemistry and Molecular Biology*, 34–44. <https://doi.org/10.1016/j.jsbmb.2015.04.005>
10. Baser, G., Yildiz, N., & Calan, M. (2016). Evaluation of Meibomian Gland Dysfunction in Polycystic Ovary Syndrome and Obesity. *Current Eye Research*, 661–665. <https://doi.org/10.1080/02713683.2016.1233985>
11. Behre, H. M., Kliesch, S., Leifke, E., Link, T. M., & Nieschlag, E. (1997). Long-Term Effect of Testosterone Therapy on Bone Mineral Density in Hypogonadal Men. *The Journal of Clinical Endocrinology & Metabolism*, 2386–2390. <https://doi.org/10.1210/jcem.82.8.4163>
12. Beier, K., Ginez, I., & Schaller, H. (2004). Localization of steroid hormone receptors in the apocrine sweat glands of the human axilla. *Histochemistry and Cell Biology*, 61–65. <https://doi.org/10.1007/s00418-004-0736-3>
13. Bonini, S., Mantelli, F., Moretti, C., Lambiase, A., Bonini, S., & Micera, A. (2007). Itchy-Dry Eye Associated with Polycystic Ovary Syndrome. *American Journal of Ophthalmology*, 763-771.e2. <https://doi.org/10.1016/j.ajo.2007.01.030>
14. Bourghardt, J., Wilhelmson, A. S. K., Alexanderson, C., De Gendt, K., Verhoeven, G., Krettek, A., Ohlsson, C., & Tivesten, Å. (2010). Androgen Receptor-Dependent and Independent Atheroprotection by Testosterone in Male Mice. *Endocrinology*, 5428–5437. <https://doi.org/10.1210/en.2010-0663>
15. Brown, E. E., Lewin, A. S., & Ash, J. D. (2018). Mitochondria: Potential Targets for Protection in Age-Related Macular Degeneration. In *Retinal Degenerative Diseases* (pp. 11–17). Springer International Publishing. https://doi.org/10.1007/978-3-319-75402-4_2
16. Brown, M. A., & Su, M. A. (2019). An Inconvenient Variable: Sex Hormones and Their Impact

- on T Cell Responses. *The Journal of Immunology*, 1927–1933.
<https://doi.org/10.4049/jimmunol.1801403>
17. Bullock, B. P., Heller, R. S., & Habener, J. F. (1996). Tissue distribution of messenger ribonucleic acid encoding the rat glucagon-like peptide-1 receptor. *Endocrinology*, 2968–2978.
<https://doi.org/10.1210/endo.137.7.8770921>
18. Ceruti, J. M., Leirós, G. J., & Balañá, M. E. (2018). Androgens and androgen receptor action in skin and hair follicles. *Molecular and Cellular Endocrinology*, 122–133.
<https://doi.org/10.1016/j.mce.2017.09.009>
19. Chakrabarty, S., Kabekkodu, S. P., Singh, R. P., Thangaraj, K., Singh, K. K., & Satyamoorthy, K. (2018). Mitochondria in health and disease. *Mitochondrion*, 25–29.
<https://doi.org/10.1016/j.mito.2018.06.006>
20. Chen, J.-F., Lin, P.-W., Tsai, Y.-R., Yang, Y.-C., & Kang, H.-Y. (2019). Androgens and Androgen Receptor Actions on Bone Health and Disease: From Androgen Deficiency to Androgen Therapy. *Cells*, 1318. <https://doi.org/10.3390/cells8111318>
21. Chen, W., Beck, I., Schober, W., Brockow, K., Effner, R., Buters, J. T. M., Behrendt, H., & Ring, J. (2010). Human mast cells express androgen receptors but treatment with testosterone exerts no influence on IgE-independent mast cell degranulation elicited by neuromuscular blocking agents. *Experimental Dermatology*, 302–304. <https://doi.org/10.1111/j.1600-0625.2009.00969.x>
22. Chen, Y., Ji, Y., Jin, X., Sun, X., Zhang, X., Chen, Y., Shi, L., Cheng, H., Mao, Y., Li, X., Hou, Y., Zhang, D., Zhao, S., Ma, J., & Huang, S. (2019). Mitochondrial abnormalities are involved in periodontal ligament fibroblast apoptosis induced by oxidative stress. *Biochemical and Biophysical Research Communications*, 483–490. <https://doi.org/10.1016/j.bbrc.2018.12.143>
23. Chiang, J. Y. L. (2013). Bile Acid Metabolism and Signaling. In *Comprehensive Physiology*. John Wiley & Sons, Inc. <https://doi.org/10.1002/cphy.c120023>
24. Chivet, M., Marchioretto, C., Pirazzini, M., Piol, D., Scaramuzzino, C., Polanco, J., Nath, S., Zuccaro, E., Nogara, L., Canato, M., Marcucci, L., Parodi, S., Romanello, V., Armani, A., D'Antonio, M., Sambataro, F., Dassi, E., Pegoraro, E., Sorarù, G., ... Pennuto, M. (2019). *Polyglutamine-expanded androgen receptor disrupts muscle triad, calcium dynamics and the excitation-contraction coupling gene expression program*. Cold Spring Harbor Laboratory. <https://doi.org/10.1101/618405>

25. Chuang, K.-H., Altuwajri, S., Li, G., Lai, J.-J., Chu, C.-Y., Lai, K.-P., Lin, H.-Y., Hsu, J.-W., Keng, P., Wu, M.-C., & Chang, C. (2009). Neutropenia with impaired host defense against microbial infection in mice lacking androgen receptor. *The Journal of Experimental Medicine*, 1181–1199. <https://doi.org/10.1084/jem.20082521>
26. COHEN, J. C., & HICKMAN, R. (1987). Insulin Resistance and Diminished Glucose Tolerance in Powerlifters Ingesting Anabolic Steroids*. *The Journal of Clinical Endocrinology & Metabolism*, 960–963. <https://doi.org/10.1210/jcem-64-5-960>
27. Collett, G., Betts, A., Johnson, M., Pulimood, A., Cook, S., Neal, D., & Robson, C. (2000). Peroxisome proliferator-activated receptor alpha is an androgen-responsive gene in human prostate and is highly expressed in prostatic adenocarcinoma. *Clinical Cancer Research : An Official Journal of the American Association for Cancer Research*, 6(8), 3241–3248. <https://www.ncbi.nlm.nih.gov/pubmed/10955810>
28. Cooper, S., Dawber, R., & Hilton-Jones, D. (2003). Three cases of androgen-dependent disease associated with myotonic dystrophy. *Journal of the European Academy of Dermatology and Venereology*, 56–58. <https://doi.org/10.1046/j.1468-3083.2003.00507.x>
29. Cutolo, M. (2009). Androgens in rheumatoid arthritis: when are they effectors? *Arthritis Research & Therapy*, 126. <https://doi.org/10.1186/ar2804>
30. Dale, J. B., Sarich, S. L., Bretz, T. M., Hatton, J. F., & Zachow, R. J. (2002). Hormonal Regulation of Androgen Receptor Messenger Ribonucleic Acid Expression in Human Tooth Pulp. *Journal of Dental Research*, 360–365. <https://doi.org/10.1177/154405910208100514>
31. Danilovic, D. L. S., Correa, P. H. S., Costa, E. M. F., Melo, K. F. S., Mendonca, B. B., & Arnhold, I. J. P. (2006). Height and bone mineral density in androgen insensitivity syndrome with mutations in the androgen receptor gene. *Osteoporosis International*, 369–374. <https://doi.org/10.1007/s00198-006-0243-6>
32. Darryll M.A. Oliver, & P. Hemachandra Reddy. (2019). Molecular Basis of Alzheimer's Disease: Focus on Mitochondria. *Journal of Alzheimer's Disease*, S95–S116. <https://doi.org/10.3233/JAD-190048>
33. Desai, J. V., & Lionakis, M. S. (2018). The Role of Neutrophils in Host Defense Against Invasive

- Fungal Infections. *Current Clinical Microbiology Reports*, 181–189.
<https://doi.org/10.1007/s40588-018-0098-6>
34. Di Loreto, C. (2011, September 16). *A semi quantitative evaluation of % cells positive for AR.* -, Trieste.
35. Diviccaro, S., Giatti, S., Borgo, F., Barcella, M., Borghi, E., Trejo, J. L., Garcia-Segura, L. M., & Melcangi, R. C. (2019). Treatment of male rats with finasteride, an inhibitor of 5 α -reductase enzyme, induces long-lasting effects on depressive-like behavior, hippocampal neurogenesis, neuroinflammation and gut microbiota composition. *Psychoneuroendocrinology*, 206–215.
<https://doi.org/10.1016/j.psyneuen.2018.09.021>
36. Dowling, J. J., Lawlor, M. W., & Dirksen, R. T. (2014). Triadopathies: An Emerging Class of Skeletal Muscle Diseases. *Neurotherapeutics*, 773–785.
<https://doi.org/10.1007/s13311-014-0300-3>
37. Dubois, V., Laurent, M. R., Sinnesael, M., Cielen, N., Helsen, C., Clinckemalie, L., Spans, L., Gayan?Ramirez, G., Deldicque, L., Hespel, P., Carmeliet, G., Vanderschueren, D., & Claessens, F. (2014). A satellite cell?specific knockout of the androgen receptor reveals myostatin as a direct androgen target in skeletal muscle. *The FASEB Journal*, 2979–2994.
<https://doi.org/10.1096/fj.14-249748>
38. Eaton, V. S., & Blackmore, T. K. (2001). Nilutamide-induced neutropenia. *BJU International*, 801–802. <https://doi.org/10.1046/j.1464-4096.2001.00905.x>
39. Escobar-Morreale, H. F., Alvarez-Blasco, F., Botella-Carretero, J. I., & Luque-Ramirez, M. (2014). The striking similarities in the metabolic associations of female androgen excess and male androgen deficiency. *Human Reproduction*, 2083–2091. <https://doi.org/10.1093/humrep/deu198>
40. Fan, W., Yanase, T., Nishi, Y., Chiba, S., Okabe, T., Nomura, M., Yoshimatsu, H., Kato, S., Takayanagi, R., & Nawata, H. (2008). Functional Potentiation of Leptin-Signal Transducer and Activator of Transcription 3 Signaling by the Androgen Receptor. *Endocrinology*, 6028–6036.
<https://doi.org/10.1210/en.2008-0431>
41. Fanton, L. E., Macedo, C. G., Torres-Chávez, K. E., Fischer, L., & Tambeli, C. H. (2017). Activational action of testosterone on androgen receptors protects males preventing temporomandibular joint pain. *Pharmacology Biochemistry and Behavior*, 30–35.
<https://doi.org/10.1016/j.pbb.2016.07.005>

42. Fimmel, S., & Zouboulis, C. C. (2005). Influence of physiological androgen levels on wound healing and immune status in men. *The Aging Male*, 166–174. <https://doi.org/10.1080/13685530500233847>
43. Forger, N. G., Hodges, L. L., Roberts, S. L., & Breedlove, S. M. (1992). Regulation of motoneuron death in the spinal nucleus of the bulbocavernosus. *Journal of Neurobiology*, 1192–1203. <https://doi.org/10.1002/neu.480230910>
44. Forger, N., Roberts, S., Wong, V., & Breedlove, S. (1993). Ciliary neurotrophic factor maintains motoneurons and their target muscles in developing rats. *The Journal of Neuroscience*, 4720–4726. <https://doi.org/10.1523/jneurosci.13-11-04720.1993>
45. Gibson, D. A., Saunders, P. T. K., & McEwan, I. J. (2018). Androgens and androgen receptor: Above and beyond. *Molecular and Cellular Endocrinology*, 1–3. <https://doi.org/10.1016/j.mce.2018.02.013>
46. Gilliver, S., Ruckshanthi, J., Hardman, M., Zeef, L., & Ashcroft, G. (2009). 5 α -Dihydrotestosterone (DHT) retards wound closure by inhibiting re-epithelialization. *The Journal of Pathology*, 73–82. <https://doi.org/10.1002/path.2444>
47. González-Montelongo, María C., Marín, R., Gómez, T., & Díaz, M. (2006). Androgens Differentially Potentiate Mouse Intestinal Smooth Muscle by Nongenomic Activation of Polyamine Synthesis and Rho Kinase Activation. *Endocrinology*, 5715–5729. <https://doi.org/10.1210/en.2006-0780>
48. González-Montelongo, María C., Marín, R., Gómez, T., Marrero-Alonso, J., & Díaz, M. (2010). Androgens Induce Nongenomic Stimulation of Colonic Contractile Activity through Induction of Calcium Sensitization and Phosphorylation of LC20 and CPI-17. *Molecular Endocrinology*, 1007–1023. <https://doi.org/10.1210/me.2009-0472>
49. González-Montelongo, María C., Marín, R., Pérez, J. A., Gómez, T., & Díaz, M. (2013). Polyamines Transduce the Nongenomic, Androgen-Induced Calcium Sensitization in Intestinal Smooth Muscle. *Molecular Endocrinology*, 1603–1616. <https://doi.org/10.1210/me.2013-1201>
50. Gordon, S., & Taylor, P. R. (2005). Monocyte and macrophage heterogeneity. *Nature Reviews Immunology*, 953–964. <https://doi.org/10.1038/nri1733>

51. Gubbels Bupp, M. R., & Jorgensen, T. N. (2018). Androgen-Induced Immunosuppression. *Frontiers in Immunology*. <https://doi.org/10.3389/fimmu.2018.00794>
52. Guo, Y., Qi, Y., Yang, X., Zhao, L., Wen, S., Liu, Y., & Tang, L. (2016). Association between Polycystic Ovary Syndrome and Gut Microbiota. *PLOS ONE*, e0153196. <https://doi.org/10.1371/journal.pone.0153196>
53. HAMILTON, J. (1948). Influence of the endocrine status upon pigmentation in man and in mammals. *Annals of the New York Academy of Sciences*, 4, 341–357. <https://www.ncbi.nlm.nih.gov/pubmed/18862177>
54. HAMILTON, J. B. (1941). MALE HORMONE SUBSTANCE: A PRIME FACTOR IN ACNE1. *The Journal of Clinical Endocrinology & Metabolism*, 570–592. <https://doi.org/10.1210/jcem-1-7-570>
55. Harada, N., Hanaoka, R., Horiuchi, H., Kitakaze, T., Mitani, T., Inui, H., & Yamaji, R. (2016). Castration influences intestinal microflora and induces abdominal obesity in high-fat diet-fed mice. *Scientific Reports*. <https://doi.org/10.1038/srep23001>
56. Higuchi, M., Kudo, T., Suzuki, S., Evans, T. T., Sasaki, R., Wada, Y., Shirakawa, T., Sawyer, J. R., & Gotoh, A. (2005). Mitochondrial DNA determines androgen dependence in prostate cancer cell lines. *Oncogene*, 1437–1445. <https://doi.org/10.1038/sj.onc.1209190>
57. Hofer, M. D., Cheng, E. Y., Bury, M. I., Xu, W., Hong, S. J., Kaplan, W. E., & Sharma, A. K. (2015). Androgen Supplementation in Rats Increases the Inflammatory Response and Prolongs Urethral Healing. *Urology*, 691–697. <https://doi.org/10.1016/j.urology.2014.11.025>
58. HOLMÄNG, A., & BJÖRNTORP, P. (1992). The effects of testosterone on insulin sensitivity in male rats. *Acta Physiologica Scandinavica*, 505–510. <https://doi.org/10.1111/j.1748-1716.1992.tb09452.x>
59. Houghton, L. A., Heitkemper, M., Crowell, M. D., Emmanuel, A., Halpert, A., McRoberts, J. A., & Toner, B. (2016). Age, Gender, and Women’s Health and the Patient. *Gastroenterology*, 1332-1343.e4. <https://doi.org/10.1053/j.gastro.2016.02.017>

60. Hulsmans, M., Van Dooren, E., & Holvoet, P. (2012). Mitochondrial Reactive Oxygen Species and Risk of Atherosclerosis. *Current Atherosclerosis Reports*, 264–276.
<https://doi.org/10.1007/s11883-012-0237-0>
61. Johansen, J. A., Breedlove, S. M., & Jordan, C. L. (2007). Androgen Receptor Expression in the Levator Ani Muscle of Male Mice. *Journal of Neuroendocrinology*, 823–826.
<https://doi.org/10.1111/j.1365-2826.2007.01592.x>
62. Jones, H. R., Robb, C. T., Perretti, M., & Rossi, A. G. (2016). The role of neutrophils in inflammation resolution. *Seminars in Immunology*, 137–145.
<https://doi.org/10.1016/j.smim.2016.03.007>
63. Jordan, C. L., Breedlove, S. M., & Arnold, A. P. (1991). Ontogeny of steroid accumulation in spinal lumbar motoneurons of the rat: Implications for androgen's site of action during synapse elimination. *The Journal of Comparative Neurology*, 441–448.
<https://doi.org/10.1002/cne.903130304>
64. Jordan, C., Padgett, B., Hershey, J., Prins, G., & Arnold, A. (1997). Ontogeny of androgen receptor immunoreactivity in lumbar motoneurons and in the sexually dimorphic levator ani muscle of male rats. *The Journal of Comparative Neurology*, 379(1), 88–98.
<https://www.ncbi.nlm.nih.gov/pubmed/9057114>
65. Joung, J. Y., Kim, S. H., Kim, S., Rha, K. H., Kim, H. G., Kwak, C., Lee, J. Y., Jeon, S. S., Hong, S. K., Jeong, H., Jo, M. K., You, D., Jeong, I. G., Hong, J. H., & Kim, C.-S. (2017). Comparison of bone mineral loss by combined androgen block agonist versus GnRH in patients with prostate cancer: A 12 month-prospective observational study. *Scientific Reports*.
<https://doi.org/10.1038/srep39562>
66. Kadel, S., & Kovats, S. (2018). Sex Hormones Regulate Innate Immune Cells and Promote Sex Differences in Respiratory Virus Infection. *Frontiers in Immunology*.
<https://doi.org/10.3389/fimmu.2018.01653>
67. Kadmiel, M., & Cidlowski, J. A. (2013). Glucocorticoid receptor signaling in health and disease. *Trends in Pharmacological Sciences*, 518–530. <https://doi.org/10.1016/j.tips.2013.07.003>
68. Kalinchenko, S. Y., Tishova, Y. A., Mskhalaya, G. J., Gooren, L. J. G., Giltay, E. J., & Saad, F. (2010). Effects of testosterone supplementation on markers of the metabolic syndrome and inflammation in hypogonadal men with the metabolic syndrome: the double-blinded placebo-

controlled Moscow study. *Clinical Endocrinology*, 602–612.

<https://doi.org/10.1111/j.1365-2265.2010.03845.x>

69. Kamogashira, T., Fujimoto, C., & Yamasoba, T. (2015). Reactive Oxygen Species, Apoptosis, and Mitochondrial Dysfunction in Hearing Loss. *BioMed Research International*, 1–7. <https://doi.org/10.1155/2015/617207>
70. Khandelwal, P., Liu, S., & Sullivan, D. (2012). Androgen regulation of gene expression in human meibomian gland and conjunctival epithelial cells. *Molecular Vision*, 18, 1055–1067. <https://www.ncbi.nlm.nih.gov/pubmed/22605918>
71. Kim, B. J., Rhee, P.-L., Park, J. H., Chang, D. K., Kim, Y.-H., Son, H. J., Kim, J. J., Rhee, J. C., & Lee, H. (2008). Male Sex Hormones May Influence the Symptoms of Irritable Bowel Syndrome in Young Men. *Digestion*, 88–92. <https://doi.org/10.1159/000166600>
72. Kim, Y. S., & Kim, N. (2018). Sex-Gender Differences in Irritable Bowel Syndrome. *Journal of Neurogastroenterology and Motility*, 544–558. <https://doi.org/10.5056/jnm18082>
73. Köhn, F.-M., Ring, J., & Schill, W.-B. (2000). *Der Hautarzt*, 223. <https://doi.org/10.1007/s001050051109>
74. Lai, J.-J., Chang, P., Lai, K.-P., Chen, L., & Chang, C. (2012). The role of androgen and androgen receptor in skin-related disorders. *Archives of Dermatological Research*, 499–510. <https://doi.org/10.1007/s00403-012-1265-x>
75. Lai, J.-J., Lai, K.-P., Chuang, K.-H., Chang, P., Yu, I.-C., Lin, W.-J., & Chang, C. (2009). Monocyte/macrophage androgen receptor suppresses cutaneous wound healing in mice by enhancing local TNF- α expression. *Journal of Clinical Investigation*, 3739–3751. <https://doi.org/10.1172/jci39335>
76. Lai, J.-J., Lai, K.-P., Zeng, W., Chuang, K.-H., Altuwaijri, S., & Chang, C. (2012). Androgen Receptor Influences on Body Defense System via Modulation of Innate and Adaptive Immune Systems. *The American Journal of Pathology*, 1504–1512. <https://doi.org/10.1016/j.ajpath.2012.07.008>
77. Li, K., Zhang, C., Yang, Z., Wang, Y., & Si, H. (2017). Evaluation of a novel dry eye model induced by oral administration of finasteride. *Molecular Medicine Reports*, 8763–8770.

<https://doi.org/10.3892/mmr.2017.7754>

78. Li, P. A., Hou, X., & Hao, S. (2017). Mitochondrial biogenesis in neurodegeneration. *Journal of Neuroscience Research*, 2025–2029. <https://doi.org/10.1002/jnr.24042>
79. Liu, P.-S., & Ho, P.-C. (2018). Mitochondria: A master regulator in macrophage and T cell immunity. *Mitochondrion*, 45–50. <https://doi.org/10.1016/j.mito.2017.11.002>
80. Lorenzo, O., Ramírez, E., Picatoste, B., Egado, J., & Tuñón, J. (2013). Alteration of Energy Substrates and ROS Production in Diabetic Cardiomyopathy. *Mediators of Inflammation*, 1–11. <https://doi.org/10.1155/2013/461967>
81. Ma, W., Zhang, J., Guo, L., Wang, Y., Lu, S., Wang, Z., Lu, Q., & Wei, F. (2019). Suppressed androgen receptor expression promotes M2 macrophage reprogramming through the STAT3/SOCS3 pathway. *EXCLI Journal*, 18, 21–29. <https://www.ncbi.nlm.nih.gov/pubmed/30956636>
82. Malkin, C. J., Pugh, P. J., Jones, R. D., Kapoor, D., Channer, K. S., & Jones, T. H. (2004). The Effect of Testosterone Replacement on Endogenous Inflammatory Cytokines and Lipid Profiles in Hypogonadal Men. *The Journal of Clinical Endocrinology & Metabolism*, 3313–3318. <https://doi.org/10.1210/jc.2003-031069>
83. Mantalaris, A., Panoskaltsis, N., Sakai, Y., Bourne, P., Chang, C., Messing, E. M., & David Wu, J. H. (2001). Localization of androgen receptor expression in human bone marrow. *The Journal of Pathology*, 361–366. [https://doi.org/10.1002/1096-9896\(0000\)9999:9999<::aid-path803>3.0.co;2-w](https://doi.org/10.1002/1096-9896(0000)9999:9999<::aid-path803>3.0.co;2-w)
84. Mantelli, F., Moretti, C., Micera, A., & Bonini, S. (2006). Conjunctival mucin deficiency in complete androgen insensitivity syndrome (CAIS). *Graefe's Archive for Clinical and Experimental Ophthalmology*, 899–902. <https://doi.org/10.1007/s00417-006-0452-x>
85. Markova, M. S., Zeskand, J., McEntee, B., Rothstein, J., Jimenez, S. A., & Siracusa, L. D. (2004). A Role for the Androgen Receptor in Collagen Content of the Skin. *Journal of Investigative Dermatology*, 1052–1056. <https://doi.org/10.1111/j.0022-202x.2004.23494.x>
86. McDonnell, N. D., & Livingston, R. B. (1994). Severe Reversible Neutropenia Following Treatment of Prostate Cancer with Flutamide. *Journal of Urology*, 1353–1354.

[https://doi.org/10.1016/s0022-5347\(17\)35251-5](https://doi.org/10.1016/s0022-5347(17)35251-5)

87. McPherron, A. C., Lawler, A. M., & Lee, S.-J. (1997). Regulation of skeletal muscle mass in mice by a new TGF- β superfamily member. *Nature*, 83–90. <https://doi.org/10.1038/387083a0>
88. Mendler, L., Baka, Z., Kovács-Simon, A., & Dux, L. (2007). Androgens negatively regulate myostatin expression in an androgen-dependent skeletal muscle. *Biochemical and Biophysical Research Communications*, 237–242. <https://doi.org/10.1016/j.bbrc.2007.07.023>
89. Mishra, J. S., More, A. S., & Kumar, S. (2018). Elevated androgen levels induce hyperinsulinemia through increase in Ins1 transcription in pancreatic beta cells in female rats†. *Biology of Reproduction*, 520–531. <https://doi.org/10.1093/biolre/iy017>
90. Monks, D. A., Johansen, J. A., Mo, K., Rao, P., Eagleson, B., Yu, Z., Lieberman, A. P., Breedlove, S. M., & Jordan, C. L. (2007). Overexpression of wild-type androgen receptor in muscle recapitulates polyglutamine disease. *Proceedings of the National Academy of Sciences*, 18259–18264. <https://doi.org/10.1073/pnas.0705501104>
91. Monks, D. Ashley, & Holmes, M. M. (2017). Androgen receptors and muscle: a key mechanism underlying life history trade-offs. *Journal of Comparative Physiology A*, 51–60. <https://doi.org/10.1007/s00359-017-1222-4>
92. Monks, Douglas Ashley, O'Bryant, E. L., & Jordan, C. L. (2004). Androgen receptor immunoreactivity in skeletal muscle: Enrichment at the neuromuscular junction. *The Journal of Comparative Neurology*, 59–72. <https://doi.org/10.1002/cne.20088>
93. Morford, J. J., Wu, S., & Mauvais-Jarvis, F. (2018). The impact of androgen actions in neurons on metabolic health and disease. *Molecular and Cellular Endocrinology*, 92–102. <https://doi.org/10.1016/j.mce.2017.09.001>
94. Morthen, M. K., Tellefsen, S., Richards, S. M., Lieberman, S. M., Rahimi Darabad, R., Kam, W. R., & Sullivan, D. A. (2019). Testosterone Influence on Gene Expression in Lacrimal Glands of Mouse Models of Sjögren Syndrome. *Investigative Ophthalmology & Visual Science*, 2181. <https://doi.org/10.1167/iovs.19-26815>
95. Motofei, I. G., Rowland, D. L., Georgescu, S. R., Tampa, M., Paunica, S., Constantin, V. D., Balalau, C., Manea, M., Baleanu, B. C., & Sinescu, I. (2017). Post-Finasteride Adverse Effects in

Male Androgenic Alopecia: A Case Report of Vitiligo. *Skin Pharmacology and Physiology*, 42–45. <https://doi.org/10.1159/000455972>

96. Narayanan, R., Coss, C. C., & Dalton, J. T. (2018). Development of selective androgen receptor modulators (SARMs). *Molecular and Cellular Endocrinology*, 134–142. <https://doi.org/10.1016/j.mce.2017.06.013>
97. Nath, S. R., Yu, Z., Gipson, T. A., Marsh, G. B., Yoshidome, E., Robins, D. M., Todi, S. V., Housman, D. E., & Lieberman, A. P. (2018). Androgen receptor polyglutamine expansion drives age-dependent quality control defects and muscle dysfunction. *Journal of Clinical Investigation*, 3630–3641. <https://doi.org/10.1172/jci99042>
98. Navarro, G., Allard, C., Morford, J. J., Xu, W., Liu, S., Molinas, A. J. R., Butcher, S. M., Fine, N. H. F., Blandino-Rosano, M., Sure, V. N., Yu, S., Zhang, R., Münzberg, H., Jacobson, D. A., Katakam, P. V., Hodson, D. J., Bernal-Mizrachi, E., Zsombok, A., & Mauvais-Jarvis, F. (2018). Androgen excess in pancreatic β cells and neurons predisposes female mice to type 2 diabetes. *JCI Insight*. <https://doi.org/10.1172/jci.insight.98607>
99. Navarro, G., Xu, W., Jacobson, D. A., Wicksteed, B., Allard, C., Zhang, G., De Gendt, K., Kim, S. H., Wu, H., Zhang, H., Verhoeven, G., Katzenellenbogen, J. A., & Mauvais-Jarvis, F. (2016). Extranuclear Actions of the Androgen Receptor Enhance Glucose-Stimulated Insulin Secretion in the Male. *Cell Metabolism*, 837–851. <https://doi.org/10.1016/j.cmet.2016.03.015>
100. Navarro, V. M., Gottsch, M. L., Wu, M., García-Galiano, D., Hobbs, S. J., Bosch, M. A., Pinilla, L., Clifton, D. K., Dearth, A., Ronnekleiv, O. K., Braun, R. E., Palmiter, R. D., Tena-Sempere, M., Alreja, M., & Steiner, R. A. (2011). Regulation of NKB Pathways and Their Roles in the Control of Kiss1 Neurons in the Arcuate Nucleus of the Male Mouse. *Endocrinology*, 4265–4275. <https://doi.org/10.1210/en.2011-1143>
101. Parkar, M. H., Newman, H. N., & Olsen, I. (1996). Polymerase chain reaction analysis of oestrogen and androgen receptor expression in human gingival and periodontal tissue. *Archives of Oral Biology*, 979–983. [https://doi.org/10.1016/s0003-9969\(96\)00053-2](https://doi.org/10.1016/s0003-9969(96)00053-2)
102. Pelletier, G., & Ren, L. (2004). Localization of sex steroid receptors in human skin. *Histology and Histopathology*, 19(2), 629–636. <https://doi.org/10.14670/HH-19.629>
103. Pham, C. T. N. (2006). Neutrophil serine proteases: specific regulators of inflammation. *Nature Reviews Immunology*, 541–550. <https://doi.org/10.1038/nri1841>

104. Pitteloud, N., Mootha, V. K., Dwyer, A. A., Hardin, M., Lee, H., Eriksson, K.-F., Tripathy, D., Yialamas, M., Groop, L., Elahi, D., & Hayes, F. J. (2005). Relationship Between Testosterone Levels, Insulin Sensitivity, and Mitochondrial Function in Men. *Diabetes Care*, 1636–1642. <https://doi.org/10.2337/diacare.28.7.1636>
105. Ponnusamy, S., Coss, C. C., Thiyagarajan, T., Watts, K., Hwang, D.-J., He, Y., Selth, L. A., McEwan, I. J., Duke, C. B., Pagadala, J., Singh, G., Wake, R. W., Ledbetter, C., Tilley, W. D., Moldoveanu, T., Dalton, J. T., Miller, D. D., & Narayanan, R. (2017). Novel Selective Agents for the Degradation of Androgen Receptor Variants to Treat Castration-Resistant Prostate Cancer. *Cancer Research*, 6282–6298. <https://doi.org/10.1158/0008-5472.can-17-0976>
106. Raskin, K., de Gendt, K., Duittoz, A., Liere, P., Verhoeven, G., Tronche, F., & Mhaouty-Kodja, S. (2009). Conditional Inactivation of Androgen Receptor Gene in the Nervous System: Effects on Male Behavioral and Neuroendocrine Responses. *Journal of Neuroscience*, 4461–4470. <https://doi.org/10.1523/jneurosci.0296-09.2009>
107. Rittirsch, D., Flierl, M. A., & Ward, P. A. (2008). Harmful molecular mechanisms in sepsis. *Nature Reviews Immunology*, 776–787. <https://doi.org/10.1038/nri2402>
108. Rocha, E. M. (2000). Identification of androgen receptor protein and 5alpha -reductase mRNA in human ocular tissues. *British Journal of Ophthalmology*, 76–84. <https://doi.org/10.1136/bjo.84.1.76>
109. Ruka, K. A., Burger, L. L., & Moenter, S. M. (2016). Both Estrogen and Androgen Modify the Response to Activation of Neurokinin-3 and μ -Opioid Receptors in Arcuate Kisspeptin Neurons From Male Mice. *Endocrinology*, 752–763. <https://doi.org/10.1210/en.2015-1688>
110. Sahin, S., Eroglu, M., Selcuk, S., Turkgeldi, L., Kozali, S., Davutoglu, S., & Muhcu, M. (2014). Intrinsic factors rather than vitamin D deficiency are related to insulin resistance in lean women with polycystic ovary syndrome. *European Review for Medical and Pharmacological Sciences*, 18(19), 2851–2856. <https://www.ncbi.nlm.nih.gov/pubmed/25339479>
111. Sánchez, M., Suárez, L., Andrés, M. T., Flórez, B. H., Bordallo, J., Riestra, S., & Cantabrana, B. (2017). Modulatory effect of intestinal polyamines and trace amines on the spontaneous phasic contractions of the isolated ileum and colon rings of mice. *Food & Nutrition Research*, 1321948. <https://doi.org/10.1080/16546628.2017.1321948>

112. Scalerandi, M. V., Peinetti, N., Leimgruber, C., Cuello Rubio, M. M., Nicola, J. P., Menezes, G. B., Maldonado, C. A., & Quintar, A. A. (2018). Inefficient N2-Like Neutrophils Are Promoted by Androgens During Infection. *Frontiers in Immunology*. <https://doi.org/10.3389/fimmu.2018.01980>
113. Schröder, H. D. (1980). Organization of the motoneurons innervating the pelvic muscles of the male rat. *Journal of Comparative Neurology*, 567–587. <https://doi.org/10.1002/cne.901920313>
114. Sfanos, K. S., Markowski, M. C., Peiffer, L. B., Ernst, S. E., White, J. R., Pienta, K. J., Antonarakis, E. S., & Ross, A. E. (2018). Compositional differences in gastrointestinal microbiota in prostate cancer patients treated with androgen axis-targeted therapies. *Prostate Cancer and Prostatic Diseases*, 539–548. <https://doi.org/10.1038/s41391-018-0061-x>
115. Sherman, S. B., Sarsour, N., Salehi, M., Schroering, A., Mell, B., Joe, B., & Hill, J. W. (2018). Prenatal androgen exposure causes hypertension and gut microbiota dysbiosis. *Gut Microbes*, 1–22. <https://doi.org/10.1080/19490976.2018.1441664>
116. Simões, I. C. M., Fontes, A., Pinton, P., Zischka, H., & Wieckowski, M. R. (2018). Mitochondria in non-alcoholic fatty liver disease. *The International Journal of Biochemistry & Cell Biology*, 93–99. <https://doi.org/10.1016/j.biocel.2017.12.019>
117. Singh, K. K., & Modica-Napolitano, J. S. (2017). Special Issue: Mitochondria in Cancer. *Seminars in Cancer Biology*, iv–vi. <https://doi.org/10.1016/j.semcancer.2017.10.013>
118. Singh, S., Moksha, L., Sharma, N., Titiyal, J. S., Biswas, N. R., & Velpandian, T. (2014). Development and evaluation of animal models for sex steroid deficient dry eye. *Journal of Pharmacological and Toxicological Methods*, 29–34. <https://doi.org/10.1016/j.vascn.2014.03.004>
119. Slominski, A., Tobin, D. J., Shibahara, S., & Wortsman, J. (2004). Melanin Pigmentation in Mammalian Skin and Its Hormonal Regulation. *Physiological Reviews*, 1155–1228. <https://doi.org/10.1152/physrev.00044.2003>
120. Smith, J. T., Dungan, H. M., Stoll, E. A., Gottsch, M. L., Braun, R. E., Eacker, S. M., Clifton, D. K., & Steiner, R. A. (2005). Differential Regulation of KiSS-1 mRNA Expression by Sex Steroids in the Brain of the Male Mouse. *Endocrinology*, 2976–2984. <https://doi.org/10.1210/en.2005-0323>
121. Sobel, V., Schwartz, B., Zhu, Y.-S., Cordero, J. J., & Imperato-McGinley, J. (2006). Bone

Mineral Density in the Complete Androgen Insensitivity and 5 α -Reductase-2 Deficiency Syndromes. *The Journal of Clinical Endocrinology & Metabolism*, 3017–3023.

<https://doi.org/10.1210/jc.2005-2809>

122. Soory, M., & Viridi, H. (1998). Effects of the anti-androgen finasteride on 5 α -reductase activity in human gingival fibroblasts in response to minocycline. *Journal of Clinical Periodontology*, 67–73. <https://doi.org/10.1111/j.1600-051x.1998.tb02365.x>
123. Spaanderman, D. C. E., Nixon, M., Buurstede, J. C., Sips, H. H. C. M., Schilperoort, M., Kuipers, E. N., Backer, E. A., Kooijman, S., Rensen, P. C. N., Homer, N. Z. M., Walker, B. R., Meijer, O. C., & Kroon, J. (2019). Androgens modulate glucocorticoid receptor activity in adipose tissue and liver. *Journal of Endocrinology*, 51–63. <https://doi.org/10.1530/joe-18-0503>
124. Srinath, R., & Dobs, A. (2014). Enobosarm (GTx-024, S-22): a potential treatment for cachexia. *Future Oncology*, 187–194. <https://doi.org/10.2217/fon.13.273>
125. Steffens, Joao P., Coimbra, L. S., Ramalho-Lucas, P. D., Rossa, C., Jr., & Spolidorio, L. C. (2012). The Effect of Supra- and Subphysiologic Testosterone Levels on Ligature-Induced Bone Loss in Rats — A Radiographic and Histologic Pilot Study. *Journal of Periodontology*, 1432–1439. <https://doi.org/10.1902/jop.2012.110658>
126. Steffens, Joao Paulo, Coimbra, L. S., Rossa, C., Jr, Kantarci, A., Van Dyke, T. E., & Spolidorio, L. C. (2015). Androgen receptors and experimental bone loss — an in vivo and in vitro study. *Bone*, 683–690. <https://doi.org/10.1016/j.bone.2015.10.001>
127. Steffens, João Paulo, Santana, L. C. L., Pitombo, J. C. P., Ribeiro, D. O., Albaricci, M. C. C., Warnavin, S. von S. C., Kantarci, A., & Spolidorio, L. C. (2018). The role of androgens on periodontal repair in female rats. *Journal of Periodontology*, 486–495. <https://doi.org/10.1002/jper.17-0435>
128. Steffens, João Paulo, Valenga, H. M., Santana, L. C. L., Albaricci, M. C. da C., Kantarci, A., & Spolidorio, L. C. (2019). Role of testosterone and androgen receptor in periodontal disease progression in female rats. *Journal of Periodontology*. <https://doi.org/10.1002/jper.19-0099>
129. Szöllösi, A. G., Oláh, A., Bíró, T., & Tóth, B. I. (2017). Recent advances in the endocrinology of the sebaceous gland. *Dermato-Endocrinology*, e1361576. <https://doi.org/10.1080/19381980.2017.1361576>

130. Taylor, R. J., Taylor, A. D., & Smyth, J. V. (2002). Using an artificial neural network to predict healing times and risk factors for venous leg ulcers. *Journal of Wound Care*, 101–105. <https://doi.org/10.12968/jowc.2002.11.3.26381>
131. Thiboutot, D., Gilliland, K., Cong, Z., Jabara, S., McAllister, J. M., Sivarajah, A., & Clawson, G. (2003). Human Skin is a Steroidogenic Tissue: Steroidogenic Enzymes and Cofactors Are Expressed in Epidermis, Normal Sebocytes, and an Immortalized Sebocyte Cell Line (SEB-1). *Journal of Investigative Dermatology*, 905–914. <https://doi.org/10.1046/j.1523-1747.2003.12244.x>
132. Toraldo, G., Bhasin, S., Bakhit, M., Guo, W., Serra, C., Safer, J. D., Bhawan, J., & Jasuja, R. (2012). Topical androgen antagonism promotes cutaneous wound healing without systemic androgen deprivation by blocking β -catenin nuclear translocation and cross-talk with TGF- β signaling in keratinocytes. *Wound Repair and Regeneration*, 61–73. <https://doi.org/10.1111/j.1524-475x.2011.00757.x>
133. Torres, P. J., Skarra, D. V., Ho, B. S., Sau, L., Anvar, A. R., Kelley, S. T., & Thackray, V. G. (2019). Letrozole treatment of adult female mice results in a similar reproductive phenotype but distinct changes in metabolism and the gut microbiome compared to pubertal mice. *BMC Microbiology*. <https://doi.org/10.1186/s12866-019-1425-7>
134. Traish, A., Bolanos, J., Nair, S., Saad, F., & Morgentaler, A. (2018). Do Androgens Modulate the Pathophysiological Pathways of Inflammation? Appraising the Contemporary Evidence. *Journal of Clinical Medicine*, 549. <https://doi.org/10.3390/jcm7120549>
135. Valanejad, L., Ghareeb, M., Shiffka, S., Nadolny, C., Chen, Y., Guo, L., Verma, R., You, S., Akhlaghi, F., & Deng, R. (2018). Dysregulation of 5α - 3β -oxosteroid 5α -reductase in diabetic patients: Implications and mechanisms. *Molecular and Cellular Endocrinology*, 127–141. <https://doi.org/10.1016/j.mce.2017.10.005>
136. van der Blik, A. M., Sedensky, M. M., & Morgan, P. G. (2017). Cell Biology of the Mitochondrion. *Genetics*, 843–871. <https://doi.org/10.1534/genetics.117.300262>
137. Vanderschueren, D., Laurent, M. R., Claessens, F., Gielen, E., Lagerquist, M. K., Vandenput, L., Börjesson, A. E., & Ohlsson, C. (2014). Sex Steroid Actions in Male Bone. *Endocrine Reviews*, 906–960. <https://doi.org/10.1210/er.2014-1024>

138. Viselli, S. M., Reese, K. R., Fan, J., Kovacs, W. J., & Olsen, N. J. (1997). Androgens Alter B Cell Development in Normal Male Mice. *Cellular Immunology*, 99–104. <https://doi.org/10.1006/cimm.1997.1227>
139. Walecki, M., Eisel, F., Klug, J., Baal, N., Paradowska-Dogan, A., Wahle, E., Hackstein, H., Meinhardt, A., & Fijak, M. (2015). Androgen receptor modulates Foxp3 expression in CD4+CD25+Foxp3+ regulatory T-cells. *Molecular Biology of the Cell*, 2845–2857. <https://doi.org/10.1091/mbc.e14-08-1323>
140. Wang, Q., Kessler, M. J., Kensler, T. B., & Dechow, P. C. (2015). The mandibles of castrated male rhesus macaques (*Macaca mulatta*): The effects of orchidectomy on bone and teeth. *American Journal of Physical Anthropology*, 31–51. <https://doi.org/10.1002/ajpa.22833>
141. Wang, X.-J., Zhuo, J., Luo, G.-H., Zhu, Y.-P., Yu, D.-J., Zhao, R.-Z., Jiang, C.-Y., Shi, Y.-F., Li, H., Chen, L., Hao, K.-Y., Han, X., Zhao, S., Bei, X.-Y., Jing, Y.-F., & Xia, S.-J. (2017). Androgen Deprivation Accelerates the Prostatic Urethra Wound Healing After Thulium Laser Resection of the Prostate by Promoting Re-Epithelialization and Regulating the Macrophage Polarization. *The Prostate*, 708–717. <https://doi.org/10.1002/pros.23301>
142. Wang, Y., & Hekimi, S. (2015). Mitochondrial dysfunction and longevity in animals: Untangling the knot. *Science*, 1204–1207. <https://doi.org/10.1126/science.aac4357>
143. Wang, Yiwei, Simanainen, U., Cheer, K., Suarez, F. G., Gao, Y. R., Li, Z., Handelsman, D., & Maitz, P. (2016). Androgen actions in mouse wound healing: Minimal in vivo effects of local antiandrogen delivery. *Wound Repair and Regeneration*, 478–488. <https://doi.org/10.1111/wrr.12420>
144. Wei, L., Lai, E. C.-C., Kao-Yang, Y.-H., Walker, B. R., MacDonald, T. M., & Andrew, R. (2019). Incidence of type 2 diabetes mellitus in men receiving steroid 5 α -reductase inhibitors: population based cohort study. *BMJ*, 11204. <https://doi.org/10.1136/bmj.11204>
145. Wersinger, S. R., Haisenleder, D. J., Lubahn, D. B., & Rissman, E. F. (1999). Steroid Feedback on Gonadotropin Release and Pituitary Gonadotropin Subunit mRNA in Mice Lacking a Functional Estrogen Receptor ?. *Endocrine*, 137–144. <https://doi.org/10.1385/endo.11:2:137>
146. Wickham, L. A., Gao, J., Toda, I., Rocha, E. M., Ono, M., & Sullivan, D. A. (2000). Identification of androgen, estrogen and progesterone receptor mRNAs in the eye. *Acta Ophthalmologica Scandinavica*, 146–153. <https://doi.org/10.1034/j.1600-0420.2000.078002146.x>

147. Wiren, K. M., Semirale, A. A., Zhang, X.-W., Woo, A., Tommasini, S. M., Price, C., Schaffler, M. B., & Jepsen, K. J. (2008). Targeting of androgen receptor in bone reveals a lack of androgen anabolic action and inhibition of osteogenesis. *Bone*, 440–451. <https://doi.org/10.1016/j.bone.2008.04.026>
148. Wu, J., Henning, P., Sjögren, K., Koskela, A., Tuukkanen, J., Movérare-Skrtic, S., & Ohlsson, C. (2019). The androgen receptor is required for maintenance of bone mass in adult male mice. *Molecular and Cellular Endocrinology*, 159–169. <https://doi.org/10.1016/j.mce.2018.10.008>
149. Xu, J., Gingras, K. M., Bengston, L., Di Marco, A., & Forger, N. G. (2001). Blockade of Endogenous Neurotrophic Factors Prevents the Androgenic Rescue of Rat Spinal Motoneurons. *The Journal of Neuroscience*, 4366–4372. <https://doi.org/10.1523/jneurosci.21-12-04366.2001>
150. Xu, W., Morford, J., & Mauvais-Jarvis, F. (2019). Emerging role of testosterone in pancreatic β cell function and insulin secretion. *Journal of Endocrinology*, R97–R105. <https://doi.org/10.1530/joe-18-0573>
151. Xu, W., Niu, T., Xu, B., Navarro, G., Schipma, M. J., & Mauvais-Jarvis, F. (2017). Androgen receptor-deficient islet β -cells exhibit alteration in genetic markers of insulin secretion and inflammation. A transcriptome analysis in the male mouse. *Journal of Diabetes and Its Complications*, 787–795. <https://doi.org/10.1016/j.jdiacomp.2017.03.002>
152. Yu, I.-C., Lin, H.-Y., Liu, N.-C., Sparks, J. D., Yeh, S., Fang, L.-Y., Chen, L., & Chang, C. (2012). Neuronal Androgen Receptor Regulates Insulin Sensitivity via Suppression of Hypothalamic NF- κ B-Mediated PTP1B Expression. *Diabetes*, 411–423. <https://doi.org/10.2337/db12-0135>
153. Yu, I.-C., Lin, H.-Y., Sparks, J. D., Yeh, S., & Chang, C. (2014). Androgen Receptor Roles in Insulin Resistance and Obesity in Males: The Linkage of Androgen-Deprivation Therapy to Metabolic Syndrome. *Diabetes*, 3180–3188. <https://doi.org/10.2337/db13-1505>
154. Yu, Z. (2006). Androgen-dependent pathology demonstrates myopathic contribution to the Kennedy disease phenotype in a mouse knock-in model. *Journal of Clinical Investigation*, 2663–2672. <https://doi.org/10.1172/jci28773>
155. Yuksel, B., Ozturk, I., Seven, A., Aktas, S., Aktas, H., Kucur, S., Polat, M., & Kilic, S. (2015).

Tear function alterations in patients with polycystic ovary syndrome. *European Review for Medical and Pharmacological Sciences*, 19(19), 3556–3562.

<https://www.ncbi.nlm.nih.gov/pubmed/26502843>

156. Zhang, M. A., Rego, D., Moshkova, M., Kebir, H., Chruscinski, A., Nguyen, H., Akkermann, R., Stanczyk, F. Z., Prat, A., Steinman, L., & Dunn, S. E. (2012). Peroxisome proliferator-activated receptor (PPAR) and - regulate IFN and IL-17A production by human T cells in a sex-specific way. *Proceedings of the National Academy of Sciences*, 9505–9510. <https://doi.org/10.1073/pnas.1118458109>
157. Zhao, Ruizhe, Wang, X., Jiang, C., Shi, F., Zhu, Y., Yang, B., Zhuo, J., Jing, Y., Luo, G., Xia, S., & Han, B. (2017). Finasteride accelerates prostate wound healing after thulium laser resection through DHT and AR signalling. *Cell Proliferation*, e12415. <https://doi.org/10.1111/cpr.12415>
158. Zhao, Ru?Zhou, Jiang, S., Zhang, L., & Yu, Z. (2019). Mitochondrial electron transport chain, ROS generation and uncoupling (Review). *International Journal of Molecular Medicine*. <https://doi.org/10.3892/ijmm.2019.4188>
159. Zhu, L., Zhou, J., Pan, Y., Lv, J., Liu, Y., Yu, S., & Zhang, Y. (2019). Glucagon-like peptide-1 receptor expression and its functions are regulated by androgen. *Biomedicine & Pharmacotherapy*, 109555. <https://doi.org/10.1016/j.biopha.2019.109555>
160. Zitzmann, M., Gromoll, J., von Eckardstein, A., & Nieschlag, E. (2003). The CAG repeat polymorphism in the androgen receptor gene modulates body fat mass and serum concentrations of leptin and insulin in men. *Diabetologia*, 31–39. <https://doi.org/10.1007/s00125-002-0980-9>
-

The role of the AR in areas relevant to the neurological and psychological symptoms of PFS

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/the-role-of-the-ar-in-neurological-and-psychological-symptoms-of-pfs/>

Cognitive dysfunction, Anhedonia and Anxiety

Anxiety, loss of motivation, loss of aggression, lack of feelings of wellbeing, severe anhedonia and visuospatial and cognitive impairment are some the most frequent neuropsychological complaints in PFS. These can manifest with a profound severity and entail a devastating impact on quality of life. This is strongly suggestive of impairment in executive, reward and motivational circuitry in the brain. The important role of metabolite neurosteroids (Diotel et al., 2018), shown to be broadly deregulated in PFS (Melcangi et al., 2017), may have additional relevance to these symptomatic areas. However, this is beyond the scope of this review, which is focused upon a potential pleiotropic pathomechanism with direct relevance to the full clinical picture and underlying the pathology.

Low serum testosterone is strongly associated with an increase in depression in aging men (Ford et al., 2016) and men undergoing ADT (Lee et al., 2014). Androgens have mostly anxiolytic and antidepressant properties in humans and animals (Liang et al., 2018; McHenry et al., 2014; Zarrouf et al., 2009). Androgens regulate gene expression in key areas of the brain that are fundamental to the etiology of depression and anxiety (McHenry et al., 2014). AR-deficient mice rapidly develop depressive-like behaviour with exposure to chronic mild stress (Hung et al., 2019) and significant comparative reductions in AR in the hypothalamic paraventricular nucleus (PVN) has been identified in autopsied depression patients (Wang et al., 2008). Androgen administration has anti-depressive effects in middle-aged men with low testosterone levels (Amanatkar et al., 2014). Owens et al. reported significantly increased AR mRNA in the PFC of patients with bipolar disorder as surprising given the association of depression with low androgen levels but noted the association of excessive androgen signaling with psychological illness (Owens et al., 2019).

Impaired executive functioning and visuospatial abilities are the most frequently reported cognitive consequences of androgen deprivation therapy (Nelson et al., 2008). Additionally, multiple lines of evidence including in anabolic steroid abuse and polycystic ovary syndrome suggests increased androgen action is markedly associated with psychological illnesses including schizophrenia, psychosis, bipolar disorder, tics, anxiety and depression (Cesta et al., 2016; Piacentino et al., 2015; Wood, 2008). In an important review of the role of androgens in the mesolimbic system and of evidence that both high and

low androgen signaling causes cognitive impairment in both human and animals, Tobiansky et al. suggested that optimally required levels of androgen signaling are required within the mesolimbic system (Tobiansky et al., 2018). Mesolimbic areas crucial to executive function including the ventral tegmental area (VTA), nucleus accumbens (NAc), and prefrontal cortex (PFC) express AR (Kritzer, 1997; Low et al., 2017; Tobiansky et al., 2018), areas which functionally align with the effects of androgens on behaviour (Kritzer, 1997). As the AR relevant to function in these areas is often not concentrated in neuronal nuclei, this has been traditionally difficult to quantify and easily overlooked. Executive functioning, which includes behavioural prioritisation of goal attainment, attention, inhibitory control and working memory, critically depends on PFC function (Tobiansky et al., 2018). Importantly, all major prefrontal cortical projections in the VTA are substantially AR enriched. Androgen signaling regulates the essential dopamine innervation of the PFC and regulates glutamate signaling, potentially through these circuits (Aubele & Kritzer, 2011). The NAc is critically involved in reward behaviour and is an integrative and convergent site for reward systems in the brain (Sesack & Grace, 2009). Neurons in the NAc respond to both excitatory and inhibitory afferents from the ventral hippocampus (vHPC) (Scudder et al., 2018). In the rat, AR is colocalised with dopamine neurons in the midbrain that project to the amygdala and nucleus accumbens (Creutz & Kritzer, 2004). In line with human studies suggesting an increase in testosterone raises striatal dopamine (Hermans et al., 2010), studies in the male rat have demonstrated AR-driven modulation of molecular measures of dopamine responsiveness of the nigrostriatal pathway including regulating mRNA, levels of molecules involved in pre-synaptic dopamine synthesis, dopamine reuptake, packaging, breakdown and reception (Purves-Tyson et al., 2014). Dopamine is increased in reward regions of the rat brain in under 30 minutes (de Souza Silva et al., 2009), and the testosterone-induced effect on reward behaviour is abolished by administration of the dopamine receptor antagonist α -flupentixol (Packard et al., 1998). Coincident with a sharp decline in voluntary physical activity, AR knockout mice show a substantial loss of dopamine and dopamine receptor expression in the striatum, with upregulation of mRNA levels of the metabolic enzymes monoamine oxidase A and B (Jardí et al., 2018). Alongside a significant reduction in voluntary activity, mice with knockout of hypothalamus-specific AR exhibited a large decrease in D₂ receptor mRNA and an increase in MOAB mRNA (Clarke et al., 2019). Interestingly, androgen-anabolic steroids significantly decrease D₂ receptor in the NAc (Kindlundh et al., 2001) and testosterone administration impairs D₂ receptor-dependent set-shifting behaviour in rats (Wallin & Wood, 2015). DHT treatment inhibits the open-field induced dopamine increase in the PFC (Handa et al., 1997). This has important implications for cognitive functioning considering the importance of PFC functions. Loss of adequate D₂ receptor function in the PFC of Rhesus macaques causes cognitive deficits close to surgical ablation of the site (Brozoski et al., 1979; Tobiansky et al., 2018).

In late adolescent rats, finasteride remarkably decreases the activity of the dopaminergic system, exploratory and motor behaviours through decreasing DHT production and consequently androgen receptor activation on dopamine neurons in the Substantia nigra and VTA. Interestingly, this effect was not seen in older or younger rats (Li et al., 2017). The reported reduction in brain DHT of late adolescent rats had not been observed in younger rats in a previous study (Giatti et al., 2015), suggesting significant interruption in brain dopaminergic activity occurs when AR activation is inhibited during the time testosterone levels are at their natural peak (Li et al., 2017). This spatiotemporal observation of age-related difference is of potential relevance to the prevalence of PFS in young adult men of fertile age.

Androgens have been demonstrated to modulate the HPA stress response and modulate anxiety behaviours (Mhaouty-Kodja, 2018). While all metabolites of testosterone, including DHT, influence anxiety-like behaviours in animal models, aged male rats are more anxious than female counterparts. This difference is abolished by prepubertal orchidectomy, demonstrating this difference is androgen dependent (Domonkos et al., 2017). Evidence suggests the anxiolytic effect of T is mediated at least in part through the AR. Men treated with flutamide experience increased anxiety (Almeida et al., 2004). Intrahippocampal flutamide increases anxiety behaviour of intact and DHT-replaced male rats, but not when independently administered to gonadectomised rats (Edinger & Frye, 2006). Corticotropin-releasing hormone is an important regulator of the HPA axis and response. AR mediates regulation of corticotropin-releasing hormone mRNA in the PVN, possibly via AR-colocalising projecting neurons in the bed nucleus of the stria terminalis (Heck & Handa, 2019).

Williams et al. demonstrated sex differences in the resilience to stress-induced anhedonia in mice and revealed an androgen-mediated mechanism underlying lower vHPC-NAc excitability and correlated increase in subchronic stress resistance in male mice. Reduced sucrose preference following subchronic variable stress (SVS) was demonstrated to be female specific. Orchidectomy rendered male mice vulnerable to SVS-induced anhedonia. Testosterone to female mice was protective of SVS-induced anhedonia and decreased vHPC-NAc excitability in females. Ovariectomy, by contrast, did not affect female vHPC-NAc neuron excitability, suggesting direct mediation by the AR. It was determined that vHPC-NAc projection neurons, and many surrounding vHPC CA1 pyramidal cells highly express AR, and that bath application of the antiandrogen flutamide increased the excitability of cells (Williams et al., 2020), further suggesting interruption of androgen signalling conferred this susceptibility. The identification of this specific androgen-driven circuitry and its causal link to anhedonia suggests that a tissue-specific deregulation of the AR, as we propose in PFS, would have significant implications for dopaminergic signalling in the NAc and consequently anhedonia symptoms.

Providing a vital addition to the understanding of both the rapid effect of nonclassical androgen signaling on human social behaviour and the AR-dependency of testosterone's influence on aggression, Geniole et al. demonstrated that a single administration of testosterone to men with high-risk personality profiles increased aggression. This effect was negatively correlated with AR CAG repeat length, with shorter CAG repeat subjects exhibiting an enhanced effect. These effects were associated with increase reward feelings associated with aggression as opposed to anger associated with aggression, suggesting a rapid AR-mediated modulation of dopamine pathways in line with existing evidence (Geniole et al., 2019).

Conclusively, significant evidence indicates the curvilinear tissue response of androgen action is relevant to anxiety and mood (Owens et al., 2019) as well as cognitive function (Tobiansky et al., 2018).

Memory and spatial processing

Severe memory impairment is a common problem reported by PFS patients, with many extremely serious implications for the patient's life. The hippocampus is critical to a broad range of learning, memory, visual, spatial, and navigatory functions in mammals (Eichenbaum, 2017; Rolls & Wirth, 2018). In humans, CA1 neurons are crucial to memory formation and retrieval, as well as self-continuity, autoethic consciousness and detailed memory revisitation (Bartsch et al., 2011). The AR is highly expressed in the hippocampus, particularly in CA1 pyramidal neurons. In addition to nuclear and cytoplasmic presence, AR is localised in spines, and synaptic AR rapidly responds to androgen, directly modulating spine density by kinase network activation (Hatanaka et al., 2015; Soma et al., 2018). Pyramidal CA1 neurons require NMDA receptors for spatial and temporal memory (Huerta et al., 2000). Neural AR deletion in mice impaired NMDAR activation and prevented temporal differentiation between objects seen, revealing hippocampal CA1 AR is critical for processing of visual temporal information, possibly through an observed modulation of glutamatergic transmission (Picot et al., 2016).

AR overexpression is demonstrated to strongly alter memory-related genes in the CA1 region (Ramzan et al., 2018). Finasteride has been demonstrated to significantly decrease brain DHT levels and reversibly reduce neurogenesis in the hippocampus of mice, affecting neuronal plasticity on a structural level (Römer et al., 2010). Hippocampal AR in humans is highly expressed in both sexes. Remarkably, this is of the same order of magnitude as AR expression in the prostate of BPH patients (Beyenburg et al., 2000). Multiple studies suggest androgens as important organisational modulators of hippocampal physiology that maintain active hippocampal functions throughout life (Hamson et al., 2016; Kerr et al., 1995). Perceived male sex-related advantages in spatio-visual and navigatory abilities have been attributed to androgens rather than evolutionary adaptation (Clint et al., 2012). Reports on the effects of androgens on spatial ability have provided contradictory results, suggestive of complex regulation (Shahrzad & Nasser, 2015). Men with Alzheimer's disease have lower brain testosterone, and findings suggest that low androgens may predispose to Alzheimer's (Rosario et al., 2011). In Alzheimer's models, testosterone has been demonstrated to exert a protective effect via an AR-mediated increase hippocampal neurons, synaptic plasticity and dendritic spine density (Jia et al., 2019). However, pre-hippocampal testosterone injection causes impairment in spatial learning and memory in male Wistar rats (Gholaminejad et al., 2019). Clearly, crucial sites involved in learning, memory and spatial processing are markedly sensitive to alteration in androgen signaling.

Insomnia and sleep disordered breathing

PFS has driven patients to suicide through the rapid and persistent destruction of their ability to sleep. In

severely affected patients, this can be total. A patient who had resumed finasteride for a very short time with a stated aim of maintaining his hair for upcoming wedding photographs committed suicide after describing the rapid onset of extreme health complaints including debilitating anxiety and insomnia that prevented any sleep for a month. Severely affected patients often describe poor-quality, brief and interrupted sleep many years after brief use of the drug. This is an important and disabling symptom, the severity of which does not appear to be appreciated in literature. Additionally, patients have reported onset or worsening of sleep apnoea. Irwig found that insomnia was a common complaint in the medical records of 6 patients who committed suicide following use of Finasteride and development of persistent symptoms, and this was amongst their most debilitating symptoms (Irwig, 2020). Evidence suggests that, as well as low testosterone being associated with a decrease in sleep quality (Barrett-Connor et al., 2008), increased androgen signaling may be associated with sleep disruption and disordered breathing. Higher testosterone levels are associated with lower sleep intensity and higher ventilatory instability in men (Morselli et al., 2018), and whole genome methylation analysis has shown elevated AR protein is associated with obstructive sleep apnoea (OSA) via ventilatory instability (Chen et al., 2016). High dose exogenous testosterone can cause significant disruption of sleep to the extent of clinically relevant harm, as well as inducing and exacerbating OSA (Kim & Cho, 2019; Liu et al., 2003). Exogenous T has induced sleep apnoea in a female patient (Johnson et al., 1984). In the hyperandrogenic condition PCOS, meta-analysis of research has indicated a significant association of OSA with the syndrome (Helvacı et al., 2017). As previously mentioned, a high occurrence of sleep disorders has been reported in SBMA patients (Romigi et al., 2014). Androgens act locally in the suprachiasmatic nucleus, the hypothalamic structure controlling behavioural and physiological circadian rhythms, to influence plastic structural reorganisation and alter circadian period (Model et al., 2015). Androgen receptors are present in the suprachiasmatic nucleus, are regulated locally by androgens, and thus are an obvious site of action for a direct effect of androgen steroids (Karatsoreos & Silver, 2007). Significant clinical differences in the response of healthy men and women to a single dose of Olanzapine (Giménez et al., 2011) suggest sex differences in the mechanisms regulating sleep (Mong & Cusmano, 2016). The exact influence of sex steroids over sleep remains an important knowledge gap (Mong & Cusmano, 2016).

Head pressure

A central and potentially causative role of androgen signaling was recently demonstrated in idiopathic intracranial hypertension (IIH), which entails an increase of CSF pressure. O'Reilly et al. identified a pattern of androgen excess in female IIH patients. Like human choroid plexus, rat cells expressed AR along with androgen-metabolising enzymes. It was demonstrated that testosterone drove CSF output in rodent choroid plexus cells (O'Reilly et al., 2019). O'Reilly et al. noted that while a determinant role for androgens in IIH may seem biologically implausible considering IIH occurs less frequently in men, androgens are now known to exert sexually dimorphic effects on metabolism. The metabolic phenotype of hypogonadal men resembles that of women with androgen excess, including an increased risk of type 2 diabetes, non-alcoholic fatty liver disease and cardiovascular mortality (Ding et al., 2006; Kautzky-Willer et al., 2016). O'Reilly et al. suggest epigenetic modifications to local androgen action or differences in AR signaling in both sexes as a plausible explanation, with IIH potentially representing a

distinctive manifestation of these sex specific differences (O'Reilly et al., 2019). Interestingly, male IIH patients are more likely to have symptoms typically associated with androgen insufficiency including obstructive sleep apnoea, erectile dysfunction and loss of libido (Fraser et al., 2010). As well, androgen deprivation therapy or hypogonadism can induce IIH symptomatology (Valcamonico et al., 2013). Although in males the metabolic parabola of AR signaling is shifted far to the right compared with females (Ding et al., 2006; Morford et al., 2018), significant increases in AR signaling in men are likely to recapitulate this symptomatology, and we therefore consider it plausible IIH occurs in PFS and contributes to commonly reported symptoms, including feelings of intense pressure in the head. In this context, it is of interest that the pilot study of Melcangi et al. evaluating CSF methylation in PFS patients and controls found only one member of the control group with methylation of SRD5A2, and this patient had normal-pressure hydrocephalus. The majority of PFS patient samples exhibited variable methylation of this gene (Melcangi et al., 2019).

Methylation of SRD5A2

It is of interest that SRD5A2 was reported to be methylated in most CSF samples in a cohort of PFS patients. Interestingly, symptoms and severity per validated scales were found not to correlate to the observed methylation profiles (Melcangi et al., 2019). This is unlikely to represent a key factor in the pathological presentation when considering the symptomatic profile, novel factors of the condition and the lack of significant overlap between PFS and 5 alpha reductase insufficiency (Brinkmann et al., 2007; Imperato-McGinley et al., 1974).

5 alpha reductase type II is localised to many areas abundant in dopamine neurons and sites of projection, and finasteride has been considered for application in conditions associated with increased dopaminergic signaling including Parkinson's disease, Tourette's syndrome and schizophrenia (Castelli et al., 2013). Reduced D2 dopamine receptor binding in the nucleus accumbens has been reported in 5ar2 knockout mice. This was accompanied with behavioural deficits in aggressive, dominance, mating behaviours, along with reduced novelty seeking and risk taking. No anxiety-like, motoric or processing deficits were observed in these mice, and 5ar2 deficiency is not associated with sensorimotor deficit nor abnormalities in anxiety-like or reward-related behaviours (Mosher et al., 2018). Further, sexual desire is usually normal in human patients (Brinkmann et al., 2007). A role in neurosteroidogenesis could have some symptomatic relevance given their behavioural influences (Edinger & Frye, 2005; Ratner et al., 2019). However, hypotheses regarding the pathological alterations in PFS being localised to the nervous system do not plausibly account for the symptoms of patients, nor take appropriate account of reported evidence from investigations of peripheral tissues.

Evidence suggests that the methylation status of SRD5A2 is under regulatory influence of androgen

signaling. Both serum DHT and SRD5A2 mRNA in seminal vesicles have been demonstrated to significantly increase in inducible ARKO mice, demonstrating that SRD5A2 is regulated by the AR through local negative feedback (Wu et al., 2019). 5 α 2 expression in the rat brain has been demonstrated to be under feed-forward regulation of androgens (Torres & Ortega, 2003). In the frog *Silurana tropicalis*, Bisseger and Langlois demonstrated that while SRD5A2 was not altered at the mRNA level, DNA methylation of SRD5A2 significantly increased in the testes and ovaries following treatment with DHT, suggesting androgen modulation of epigenetic mechanisms in both sexes. The methylation statuses of SRD5A1 and SRD5A3 were not changed following androgen exposure (Bisseger & Langlois, 2016).

One possible mechanistic influence of androgen signaling on methylation of SRD5A2 is the role of androgens in inflammatory regulation and a consequential influence on the methyltransferase enzyme DNA methyltransferase 1 (DNMT1). Kang et al. found a majority of BPH samples have methylation of the SRD5A2 promoter, with strong correlation between methylation and low or absent expression of 5 α reductase 2 (Kang et al., 2018). Ge et al. reported that, in human prostate samples, DNMT1 regulates methylation of SRD5A2. The methylation of the promoter was shown to be increased by inflammatory mediators such as tumor necrosis factor (TNF- α), Nuclear factor-kappa B (NF- κ B), and Interleukin-6 (IL-6) which upregulate DNMT1 expression. Inhibition of TNF- α restored the expression of SRD5A2 (Ge et al., 2015). In prostate cancer cells, androgen signaling crosstalk exists with inflammatory signaling (Malinen et al., 2017). As previously discussed, the AR has an upregulatory effect on TNF- α expression and is thus suppressive of cutaneous wound healing (Lai et al., 2009). DHT activates macrophage TNF- α secretion through AR signaling in prostatic urethral tissue (Zhao et al., 2017). In the CNS, epigenetic macrophage activation increases proinflammatory cytokines and chemokines, including TNF- α and IL-6 (Yin et al., 2017).

Page Bibliography

1. Almeida, O. P., Waterreus, A., Spry, N., Flicker, L., & Martins, R. N. (2004). One year follow-up study of the association between chemical castration, sex hormones, beta-amyloid, memory and depression in men. *Psychoneuroendocrinology*, 1071–1081. <https://doi.org/10.1016/j.psyneuen.2003.11.002>
2. Amanatkar, H., Chibnall, J., Seo, B., Manepalli, J., & Grossberg, G. (2014). Impact of exogenous testosterone on mood: a systematic review and meta-analysis of randomized placebo-controlled trials. *Annals of Clinical Psychiatry : Official Journal of the American Academy of Clinical Psychiatrists*, 26(1), 19–32. <https://www.ncbi.nlm.nih.gov/pubmed/24501728>
3. Aubele, T., & Kritzer, M. F. (2011). Androgen Influence on Prefrontal Dopamine Systems in Adult Male Rats: Localization of Cognate Intracellular Receptors in Medial Prefrontal Projections

to the Ventral Tegmental Area and Effects of Gonadectomy and Hormone Replacement on Glutamate-Stimulated Extracellular Dopamine Level. *Cerebral Cortex*, 1799–1812.

<https://doi.org/10.1093/cercor/bhr258>

4. Barrett-Connor, E., Dam, T.-T., Stone, K., Harrison, S. L., Redline, S., & Orwoll, E. (2008). The Association of Testosterone Levels with Overall Sleep Quality, Sleep Architecture, and Sleep-Disordered Breathing. *The Journal of Clinical Endocrinology & Metabolism*, 2602–2609. <https://doi.org/10.1210/jc.2007-2622>
5. Bartsch, T., Dohring, J., Rohr, A., Jansen, O., & Deuschl, G. (2011). CA1 neurons in the human hippocampus are critical for autobiographical memory, mental time travel, and autoethic consciousness. *Proceedings of the National Academy of Sciences*, 17562–17567. <https://doi.org/10.1073/pnas.1110266108>
6. Beyenburg, S., Watzka, M., Clusmann, H., Blümcke, I., Bidlingmaier, F., Elger, C. E., & Stoffel-Wagner, B. (2000). Androgen receptor mRNA expression in the human hippocampus. *Neuroscience Letters*, 25–28. [https://doi.org/10.1016/s0304-3940\(00\)01542-1](https://doi.org/10.1016/s0304-3940(00)01542-1)
7. Bissegger, S., & Langlois, V. S. (2016). Androgens modulate gene expression and specific DNA methylation pattern of steroid 5 α -reductases in the frog *Silurana tropicalis*. *General and Comparative Endocrinology*, 123–132. <https://doi.org/10.1016/j.ygcen.2016.03.021>
8. Brinkmann, L., Schuetzmann, K., & Richter-Appelt, H. (2007). ORIGINAL RESEARCH—INTERSEX AND GENDER IDENTITY DISORDERS: Gender Assignment and Medical History of Individuals with Different Forms of Intersexuality: Evaluation of Medical Records and the Patients' Perspective. *The Journal of Sexual Medicine*, 964–980. <https://doi.org/10.1111/j.1743-6109.2007.00524.x>
9. Brozoski, T., Brown, R., Rosvold, H., & Goldman, P. (1979). Cognitive deficit caused by regional depletion of dopamine in prefrontal cortex of rhesus monkey. *Science*, 929–932. <https://doi.org/10.1126/science.112679>
10. Castelli, M. P., Casti, A., Casu, A., Frau, R., Bortolato, M., Spiga, S., & Ennas, M. G. (2013). Regional distribution of 5 α -reductase type 2 in the adult rat brain: An immunohistochemical analysis. *Psychoneuroendocrinology*, 281–293. <https://doi.org/10.1016/j.psyneuen.2012.06.008>
11. Cesta, C. E., Månsson, M., Palm, C., Lichtenstein, P., Iliadou, A. N., & Landén, M. (2016). Polycystic ovary syndrome and psychiatric disorders: Co-morbidity and heritability in a

nationwide Swedish cohort. *Psychoneuroendocrinology*, 196–203.

<https://doi.org/10.1016/j.psyneuen.2016.08.005>

12. Chen, Y.-C., Chen, T.-W., Su, M.-C., Chen, C.-J., Chen, K.-D., Liou, C.-W., Tang, P., Wang, T.-Y., Chang, J.-C., Wang, C.-C., Lin, H.-C., Chin, C.-H., Huang, K.-T., Lin, M.-C., & Hsiao, C.-C. (2016). Whole Genome DNA Methylation Analysis of Obstructive Sleep Apnea:IL1R2,NPR2,AR,SP140Methylation and Clinical Phenotype. *Sleep*, 743–755.
<https://doi.org/10.5665/sleep.5620>
13. Clarke, M. V., Russell, P. K., Zajac, J. D., & Davey, R. A. (2019). The androgen receptor in the hypothalamus positively regulates hind-limb muscle mass and voluntary physical activity in adult male mice. *The Journal of Steroid Biochemistry and Molecular Biology*, 187–194.
<https://doi.org/10.1016/j.jsbmb.2019.02.018>
14. Clint, E. K., Sober, E., Garland, T., Jr., & Rhodes, J. S. (2012). Male Superiority in Spatial Navigation: Adaptation or Side Effect? *The Quarterly Review of Biology*, 289–313.
<https://doi.org/10.1086/668168>
15. Creutz, L. M., & Kritzer, M. F. (2004). Mesostriatal and mesolimbic projections of midbrain neurons immunoreactive for estrogen receptor beta or androgen receptors in rats. *The Journal of Comparative Neurology*, 348–362. <https://doi.org/10.1002/cne.20229>
16. de Souza Silva, M. A., Mattern, C., Topic, B., Buddenberg, T. E., & Huston, J. P. (2009). Dopaminergic and serotonergic activity in neostriatum and nucleus accumbens enhanced by intranasal administration of testosterone. *European Neuropsychopharmacology*, 53–63.
<https://doi.org/10.1016/j.euroneuro.2008.08.003>
17. Ding, E. L., Song, Y., Malik, V. S., & Liu, S. (2006). Sex Differences of Endogenous Sex Hormones and Risk of Type 2 Diabetes. *JAMA*, 1288. <https://doi.org/10.1001/jama.295.11.1288>
18. Diotel, N., Charlier, T. D., Lefebvre d’Hellencourt, C., Couret, D., Trudeau, V. L., Nicolau, J. C., Meilhac, O., Kah, O., & Pellegrini, E. (2018). Steroid Transport, Local Synthesis, and Signaling within the Brain: Roles in Neurogenesis, Neuroprotection, and Sexual Behaviors. *Frontiers in Neuroscience*. <https://doi.org/10.3389/fnins.2018.00084>
19. Domonkos, E., Borbélyová, V., Csongová, M., Bosy, M., Kašmárová, M., Ostatníková, D., Hodosy, J., & Celec, P. (2017). Sex differences and sex hormones in anxiety-like behavior of aging rats. *Hormones and Behavior*, 159–165. <https://doi.org/10.1016/j.yhbeh.2017.05.019>

20. Edinger, K. L., & Frye, C. A. (2005). Testosterone's anti-anxiety and analgesic effects may be due in part to actions of its 5 α -reduced metabolites in the hippocampus. *Psychoneuroendocrinology*, 418–430. <https://doi.org/10.1016/j.psyneuen.2004.11.001>
21. Edinger, K. L., & Frye, C. A. (2006). Intrahippocampal administration of an androgen receptor antagonist, flutamide, can increase anxiety-like behavior in intact and DHT-replaced male rats. *Hormones and Behavior*, 216–222. <https://doi.org/10.1016/j.yhbeh.2006.03.003>
22. Eichenbaum, H. (2017). The role of the hippocampus in navigation is memory. *Journal of Neurophysiology*, 1785–1796. <https://doi.org/10.1152/jn.00005.2017>
23. Ford, A. H., Yeap, B. B., Flicker, L., Hankey, G. J., Chubb, S. A. P., Handelsman, D. J., Golledge, J., & Almeida, O. P. (2016). Prospective longitudinal study of testosterone and incident depression in older men: The Health In Men Study. *Psychoneuroendocrinology*, 57–65. <https://doi.org/10.1016/j.psyneuen.2015.11.012>
24. Fraser, J. A., Bruce, B. B., Rucker, J., Fraser, L.-A., Atkins, E. J., Newman, N. J., & Biousse, V. (2010). Risk factors for idiopathic intracranial hypertension in men: A case–control study. *Journal of the Neurological Sciences*, 86–89. <https://doi.org/10.1016/j.jns.2009.11.001>
25. Ge, R., Wang, Z., Bechis, S. K., Otsetov, A. G., Hua, S., Wu, S., Wu, C.-L., Tabatabaei, S., & Olumi, A. F. (2015). DNA Methyl Transferase 1 Reduces Expression of SRD5A2 in the Aging Adult Prostate. *The American Journal of Pathology*, 870–882. <https://doi.org/10.1016/j.ajpath.2014.11.020>
26. Geniole, S. N., Procyshyn, T. L., Marley, N., Ortiz, T. L., Bird, B. M., Marcellus, A. L., Welker, K. M., Bonin, P. L., Goldfarb, B., Watson, N. V., & Carré, J. M. (2019). Using a Psychopharmacogenetic Approach To Identify the Pathways Through Which—and the People for Whom—Testosterone Promotes Aggression. *Psychological Science*, 481–494. <https://doi.org/10.1177/0956797619826970>
27. Gholaminejad, A., Gholamipour-Badie, H., Nasehi, M., & Naghdi, N. (2019). Prelimbic of medial prefrontal cortex GABA modulation through testosterone on spatial learning and memory. *Iranian Journal of Pharmaceutical Research*, 18(3), 1429–1444. <https://doi.org/10.22037/IJPR.2019.1100745>

28. Giatti, S., Foglio, B., Romano, S., Pesaresi, M., Panzica, G., Garcia-Segura, L. M., Caruso, D., & Melcangi, R. C. (2015). Effects of Subchronic Finasteride Treatment and Withdrawal on Neuroactive Steroid Levels and Their Receptors in the Male Rat Brain. *Neuroendocrinology*, 746–757. <https://doi.org/10.1159/000442982>
29. Giménez, S., Romero, S., Gich, I., Clos, S., Grasa, E., Rosa-María, A., & Barbanoj, M.-J. (2011). Sex differences in sleep after a single oral morning dose of olanzapine in healthy volunteers. *Human Psychopharmacology: Clinical and Experimental*, 498–507. <https://doi.org/10.1002/hup.1232>
30. Hamson, D. K., Roes, M. M., & Galea, L. A. M. (2016). Sex Hormones and Cognition: Neuroendocrine Influences on Memory and Learning. In *Comprehensive Physiology* (pp. 1295–1337). John Wiley & Sons, Inc. <https://doi.org/10.1002/cphy.c150031>
31. Handa, R. J., Hejna, G. M., & Lorens, S. A. (1997). Androgen inhibits neurotransmitter turnover in the medial prefrontal cortex of the rat following exposure to a novel environment. *Brain Research*, 131–138. [https://doi.org/10.1016/s0006-8993\(96\)01394-7](https://doi.org/10.1016/s0006-8993(96)01394-7)
32. Hatanaka, Y., Hojo, Y., Mukai, H., Murakami, G., Komatsuzaki, Y., Kim, J., Ikeda, M., Hiragushi, A., Kimoto, T., & Kawato, S. (2015). Rapid increase of spines by dihydrotestosterone and testosterone in hippocampal neurons: Dependence on synaptic androgen receptor and kinase networks. *Brain Research*, 121–132. <https://doi.org/10.1016/j.brainres.2014.12.011>
33. Heck, A. L., & Handa, R. J. (2019). Androgens Drive Sex Biases in Hypothalamic Corticotropin-Releasing Hormone Gene Expression After Adrenalectomy of Mice. *Endocrinology*, 1757–1770. <https://doi.org/10.1210/en.2019-00238>
34. Helvacı, N., Karabulut, E., Demir, A. U., & Yildiz, B. O. (2017). Polycystic ovary syndrome and the risk of obstructive sleep apnea: a meta-analysis and review of the literature. *Endocrine Connections*, 437–445. <https://doi.org/10.1530/ec-17-0129>
35. Hermans, E. J., Bos, P. A., Ossewaarde, L., Ramsey, N. F., Fernández, G., & van Honk, J. (2010). Effects of exogenous testosterone on the ventral striatal BOLD response during reward anticipation in healthy women. *NeuroImage*, 277–283. <https://doi.org/10.1016/j.neuroimage.2010.04.019>
36. Huerta, P. T., Sun, L. D., Wilson, M. A., & Tonegawa, S. (2000). Formation of Temporal Memory Requires NMDA Receptors within CA1 Pyramidal Neurons. *Neuron*, 473–480.

[https://doi.org/10.1016/s0896-6273\(00\)80909-5](https://doi.org/10.1016/s0896-6273(00)80909-5)

37. Hung, Huang, Chang, & Kang. (2019). Deficiency in Androgen Receptor Aggravates the Depressive-Like Behaviors in Chronic Mild Stress Model of Depression. *Cells*, 1021. <https://doi.org/10.3390/cells8091021>
38. Imperato-McGinley, J., Guerrero, L., Gautier, T., & Peterson, R. E. (1974). Steroid 5 α -Reductase Deficiency in Man: An Inherited Form of Male Pseudohermaphroditism. *Science*, 1213–1215. <https://doi.org/10.1126/science.186.4170.1213>
39. Irwig, M. S. (2020). Finasteride and Suicide: A Postmarketing Case Series. *Dermatology*, 1–6. <https://doi.org/10.1159/000505151>
40. Jardí, F., Laurent, M. R., Kim, N., Khalil, R., De Bundel, D., Van Eeckhaut, A., Van Helleputte, L., Deboel, L., Dubois, V., Schollaert, D., Decallonne, B., Carmeliet, G., Van den Bosch, L., D'Hooge, R., Claessens, F., & Vanderschueren, D. (2018). Testosterone boosts physical activity in male mice via dopaminergic pathways. *Scientific Reports*. <https://doi.org/10.1038/s41598-017-19104-0>
41. Jia, J.-X., Yan, X.-S., Yang, Z.-J., Song, W., Fang, X., Cai, Z.-P., Huo, D.-S., & Wang, H. (2019). Protective mechanism of testosterone on cognitive impairment in a rat model of Alzheimer's disease. *Neural Regeneration Research*, 649. <https://doi.org/10.4103/1673-5374.245477>
42. Johnson, M., Anch, A., & Remmers, J. (1984). Induction of the obstructive sleep apnea syndrome in a woman by exogenous androgen administration. *The American Review of Respiratory Disease*, 129(6), 1023–1025. <https://doi.org/10.1164/arrd.1984.129.6.1023>
43. Kang, P. M., Kim, Y. J., Seo, W. T., Kang, S. H., Kim, T. S., Chun, B. K., Seo, W. I., Jeong, J.-Y., & Chung, J. I. (2018). Correlation between 5- α reductase type 2 protein expression and methylation of 5- α reductase type 2 promotor gene of benign prostatic hyperplasia. *World Journal of Urology*, 709–718. <https://doi.org/10.1007/s00345-018-2422-4>
44. Karatsoreos, I. N., & Silver, R. (2007). Minireview: The Neuroendocrinology of the Suprachiasmatic Nucleus as a Conductor of Body Time in Mammals. *Endocrinology*, 5640–5647. <https://doi.org/10.1210/en.2007-1083>
45. Kautzky-Willer, A., Harreiter, J., & Pacini, G. (2016). Sex and Gender Differences in Risk,

Pathophysiology and Complications of Type 2 Diabetes Mellitus. *Endocrine Reviews*, 278–316.
<https://doi.org/10.1210/er.2015-1137>

46. Kerr, J. E., Allore, R. J., Beck, S. G., & Handa, R. J. (1995). Distribution and hormonal regulation of androgen receptor (AR) and AR messenger ribonucleic acid in the rat hippocampus. *Endocrinology*, 3213–3221. <https://doi.org/10.1210/endo.136.8.7628354>
47. Kim, S.-D., & Cho, K.-S. (2019). Obstructive Sleep Apnea and Testosterone Deficiency. *The World Journal of Men's Health*, 12. <https://doi.org/10.5534/wjmh.180017>
48. Kindlundh, A. M. S., Lindblom, J., Bergström, L., Wikberg, J. E. S., & Nyberg, F. (2001). The anabolic-androgenic steroid nandrolone decanoate affects the density of dopamine receptors in the male rat brain. *European Journal of Neuroscience*, 291–296.
<https://doi.org/10.1046/j.0953-816x.2000.01402.x>
49. Kritzer, M. F. (1997). Selective colocalization of immunoreactivity for intracellular gonadal hormone receptors and tyrosine hydroxylase in the ventral tegmental area, substantia nigra, and retrorubral fields in the rat. *The Journal of Comparative Neurology*, 247–260.
[https://doi.org/10.1002/\(sici\)1096-9861\(19970310\)379:2<247::aid-cne6>3.0.co;2-3](https://doi.org/10.1002/(sici)1096-9861(19970310)379:2<247::aid-cne6>3.0.co;2-3)
50. Lai, J.-J., Lai, K.-P., Chuang, K.-H., Chang, P., Yu, I.-C., Lin, W.-J., & Chang, C. (2009). Monocyte/macrophage androgen receptor suppresses cutaneous wound healing in mice by enhancing local TNF- α expression. *Journal of Clinical Investigation*, 3739–3751.
<https://doi.org/10.1172/jci39335>
51. Lee, M., Jim, H. S., Fishman, M., Zachariah, B., Heysek, R., Biagioli, M., & Jacobsen, P. B. (2014). Depressive symptomatology in men receiving androgen deprivation therapy for prostate cancer: a controlled comparison. *Psycho-Oncology*, 472–477. <https://doi.org/10.1002/pon.3608>
52. Li, L., Kang, Y.-X., Ji, X.-M., Li, Y.-K., Li, S.-C., Zhang, X.-J., Cui, H.-X., & Shi, G.-M. (2017). Finasteride inhibited brain dopaminergic system and open-field behaviors in adolescent male rats. *CNS Neuroscience & Therapeutics*, 115–125. <https://doi.org/10.1111/cns.12781>
53. Liang, Y., Li, J., Jin, T., Gu, T., Zhu, Q., Hu, Y., Yang, Y., Li, J., Wu, D., Jiang, K., & Xu, X. (2018). Bisphenol-A inhibits improvement of testosterone in anxiety- and depression-like behaviors in gonadectomized male mice. *Hormones and Behavior*, 129–138.
<https://doi.org/10.1016/j.yhbeh.2018.05.012>

54. Liu, P. Y., Yee, B., Wishart, S. M., Jimenez, M., Jung, D. G., Grunstein, R. R., & Handelsman, D. J. (2003). The Short-Term Effects of High-Dose Testosterone on Sleep, Breathing, and Function in Older Men. *The Journal of Clinical Endocrinology & Metabolism*, 3605–3613. <https://doi.org/10.1210/jc.2003-030236>
55. Low, K. L., Ma, C., & Soma, K. K. (2017). Tyramide Signal Amplification Permits Immunohistochemical Analyses of Androgen Receptors in the Rat Prefrontal Cortex. *Journal of Histochemistry & Cytochemistry*, 295–308. <https://doi.org/10.1369/0022155417694870>
56. McHenry, J., Carrier, N., Hull, E., & Kabbaj, M. (2014). Sex differences in anxiety and depression: Role of testosterone. *Frontiers in Neuroendocrinology*, 42–57. <https://doi.org/10.1016/j.yfrne.2013.09.001>
57. Melcangi, R. C., Casarini, L., Marino, M., Santi, D., Sperduti, S., Giatti, S., Diviccaro, S., Grimoldi, M., Caruso, D., Cavaletti, G., & Simoni, M. (2019). Altered methylation pattern of the SRD5A2 gene in the cerebrospinal fluid of post-finasteride patients: a pilot study. *Endocrine Connections*, 1118–1125. <https://doi.org/10.1530/ec-19-0199>
58. Melcangi, R. C., Santi, D., Spezzano, R., Grimoldi, M., Tabacchi, T., Fusco, M. L., Diviccaro, S., Giatti, S., Carrà, G., Caruso, D., Simoni, M., & Cavaletti, G. (2017). Neuroactive steroid levels and psychiatric and andrological features in post-finasteride patients. *The Journal of Steroid Biochemistry and Molecular Biology*, 229–235. <https://doi.org/10.1016/j.jsbmb.2017.04.003>
59. Mhaouty-Kodja, S. (2018). Role of the androgen receptor in the central nervous system. *Molecular and Cellular Endocrinology*, 103–112. <https://doi.org/10.1016/j.mce.2017.08.001>
60. Model, Z., Butler, M. P., LeSauter, J., & Silver, R. (2015). Suprachiasmatic nucleus as the site of androgen action on circadian rhythms. *Hormones and Behavior*, 1–7. <https://doi.org/10.1016/j.yhbeh.2015.05.007>
61. Mong, J. A., & Cusmano, D. M. (2016). Sex differences in sleep: impact of biological sex and sex steroids. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 20150110. <https://doi.org/10.1098/rstb.2015.0110>
62. Morford, J. J., Wu, S., & Mauvais-Jarvis, F. (2018). The impact of androgen actions in neurons on metabolic health and disease. *Molecular and Cellular Endocrinology*, 92–102. <https://doi.org/10.1016/j.mce.2017.09.001>

63. Morselli, L. L., Temple, K. A., Leproult, R., Ehrmann, D. A., Van Cauter, E., & Mokhlesi, B. (2018). Determinants of Slow-Wave Activity in Overweight and Obese Adults: Roles of Sex, Obstructive Sleep Apnea and Testosterone Levels. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2018.00377>
64. Mosher, L. J., Godar, S. C., Morissette, M., McFarlin, K. M., Scheggi, S., Gambarana, C., Fowler, S. C., Di Paolo, T., & Bortolato, M. (2018). Steroid 5 α -reductase 2 deficiency leads to reduced dominance-related and impulse-control behaviors. *Psychoneuroendocrinology*, 95–104. <https://doi.org/10.1016/j.psyneuen.2018.02.007>
65. Nelson, C. J., Lee, J. S., Gamboa, M. C., & Roth, A. J. (2008). Cognitive effects of hormone therapy in men with prostate cancer. *Cancer*, 1097–1106. <https://doi.org/10.1002/cncr.23658>
66. O'Reilly, M. W., Westgate, C. S. J., Hornby, C., Botfield, H., Taylor, A. E., Markey, K., Mitchell, J. L., Scotton, W. J., Mollan, S. P., Yiangou, A., Jenkinson, C., Gilligan, L. C., Sherlock, M., Gibney, J., Tomlinson, J. W., Lavery, G. G., Hodson, D. J., Arlt, W., & Sinclair, A. J. (2019). A unique androgen excess signature in idiopathic intracranial hypertension is linked to cerebrospinal fluid dynamics. *JCI Insight*. <https://doi.org/10.1172/jci.insight.125348>
67. Owens, S. J., Purves-Tyson, T. D., Webster, M. J., & Shannon Weickert, C. (2019). Evidence for enhanced androgen action in the prefrontal cortex of people with bipolar disorder but not schizophrenia or major depressive disorder. *Psychiatry Research*, 112503. <https://doi.org/10.1016/j.psychres.2019.112503>
68. Packard, M. G., Schroeder, J. P., & Alexander, G. M. (1998). Expression of Testosterone Conditioned Place Preference Is Blocked by Peripheral or Intra-accumbens Injection of α -Flupenthixol. *Hormones and Behavior*, 39–47. <https://doi.org/10.1006/hbeh.1998.1461>
69. Piacentino, D., Kotzalidis, G., Casale, A., Aromatario, M., Pomara, C., Girardi, P., & Sani, G. (2015). Anabolic-androgenic Steroid use and Psychopathology in Athletes. A Systematic Review. *Current Neuropharmacology*, 101–121. <https://doi.org/10.2174/1570159x13666141210222725>
70. Picot, M., Billard, J.-M., Dombret, C., Albac, C., Karamah, N., Daumas, S., Hardin-Pouzet, H., & Mhaouty-Kodja, S. (2016). Neural Androgen Receptor Deletion Impairs the Temporal Processing of Objects and Hippocampal CA1-Dependent Mechanisms. *PLOS ONE*, e0148328. <https://doi.org/10.1371/journal.pone.0148328>

71. Purves-Tyson, T. D., Owens, S. J., Double, K. L., Desai, R., Handelsman, D. J., & Weickert, C. S. (2014). Testosterone Induces Molecular Changes in Dopamine Signaling Pathway Molecules in the Adolescent Male Rat Nigrostriatal Pathway. *PLoS ONE*, e91151. <https://doi.org/10.1371/journal.pone.0091151>
72. Ramzan, F., Azam, A. B., Monks, D. A., & Zovkic, I. B. (2018). Androgen receptor is a negative regulator of contextual fear memory in male mice. *Hormones and Behavior*, 10–18. <https://doi.org/10.1016/j.yhbeh.2018.08.012>
73. Ratner, M. H., Kumaresan, V., & Farb, D. H. (2019). Neurosteroid Actions in Memory and Neurologic/Neuropsychiatric Disorders. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2019.00169>
74. Rolls, E. T., & Wirth, S. (2018). Spatial representations in the primate hippocampus, and their functions in memory and navigation. *Progress in Neurobiology*, 90–113. <https://doi.org/10.1016/j.pneurobio.2018.09.004>
75. Römer, B., Pfeiffer, N., Lewicka, S., Ben-Abdallah, N., Vogt, M. A., Deuschle, M., Vollmayr, B., & Gass, P. (2010). Finasteride Treatment Inhibits Adult Hippocampal Neurogenesis in Male Mice. *Pharmacopsychiatry*, 174–178. <https://doi.org/10.1055/s-0030-1249095>
76. Romigi, A., Liguori, C., Placidi, F., Albanese, M., Izzi, F., Uasone, E., Terracciano, C., Marciani, M. G., Mercuri, N. B., Ludovisi, R., & Massa, R. (2014). Sleep disorders in spinal and bulbar muscular atrophy (Kennedy's disease): a controlled polysomnographic and self-reported questionnaires study. *Journal of Neurology*, 889–893. <https://doi.org/10.1007/s00415-014-7293-z>
77. Rosario, E. R., Chang, L., Head, E. H., Stanczyk, F. Z., & Pike, C. J. (2011). Brain levels of sex steroid hormones in men and women during normal aging and in Alzheimer's disease. *Neurobiology of Aging*, 604–613. <https://doi.org/10.1016/j.neurobiolaging.2009.04.008>
78. Scudder, S. L., Baimel, C., Macdonald, E. E., & Carter, A. G. (2018). Hippocampal-Evoked Feedforward Inhibition in the Nucleus Accumbens. *The Journal of Neuroscience*, 9091–9104. <https://doi.org/10.1523/jneurosci.1971-18.2018>
79. Sesack, S. R., & Grace, A. A. (2009). Cortico-Basal Ganglia Reward Network: Microcircuitry. *Neuropsychopharmacology*, 27–47. <https://doi.org/10.1038/npp.2009.93>

80. Shahrzad, P., & Nasser, N. (2015). GABA_B Receptor Antagonist (CGP35348) Improves Testosterone Induced Spatial Acquisition Impairment in Adult Male Rat. *Journal of Behavioral and Brain Science*, 491–502. <https://doi.org/10.4236/jbbs.2015.511047>
81. Soma, M., Kim, J., Kato, A., & Kawato, S. (2018). Src Kinase Dependent Rapid Non-genomic Modulation of Hippocampal Spinogenesis Induced by Androgen and Estrogen. *Frontiers in Neuroscience*. <https://doi.org/10.3389/fnins.2018.00282>
82. Tobiansky, D. J., Wallin-Miller, K. G., Floresco, S. B., Wood, R. I., & Soma, K. K. (2018). Androgen Regulation of the Mesocorticolimbic System and Executive Function. *Frontiers in Endocrinology*. <https://doi.org/10.3389/fendo.2018.00279>
83. Torres, J. M., & Ortega, E. (2003). Differential regulation of steroid 5 α -reductase isozymes expression by androgens in the adult rat brain. *The FASEB Journal*, 1428–1433. <https://doi.org/10.1096/fj.02-1119com>
84. Valcamonico, F., Arcangeli, G., Consoli, F., Nonnis, D., Grisanti, S., Gatti, E., Berruti, A., & Ferrari, V. (2013). Idiopathic intracranial hypertension: A possible complication in the natural history of advanced prostate cancer. *International Journal of Urology*, 335–337. <https://doi.org/10.1111/iju.12273>
85. Wallin, K. G., & Wood, R. I. (2015). Anabolic–androgenic steroids impair set-shifting and reversal learning in male rats. *European Neuropsychopharmacology*, 583–590. <https://doi.org/10.1016/j.euroneuro.2015.01.002>
86. Wang, S.-S., Kamphuis, W., Huitinga, I., Zhou, J.-N., & Swaab, D. F. (2008). Gene expression analysis in the human hypothalamus in depression by laser microdissection and real-time PCR: the presence of multiple receptor imbalances. *Molecular Psychiatry*, 786–799. <https://doi.org/10.1038/mp.2008.38>
87. Williams, E. S., Manning, C. E., Eagle, A. L., Swift-Gallant, A., Duque-Wilckens, N., Chinnusamy, S., Moeser, A., Jordan, C., Leininger, G., & Robison, A. J. (2020). Androgen-Dependent Excitability of Mouse Ventral Hippocampal Afferents to Nucleus Accumbens Underlies Sex-Specific Susceptibility to Stress. *Biological Psychiatry*, 492–501. <https://doi.org/10.1016/j.biopsych.2019.08.006>

88. Wood, R. I. (2008). Anabolic–androgenic steroid dependence? Insights from animals and humans. *Frontiers in Neuroendocrinology*, 490–506. <https://doi.org/10.1016/j.yfrne.2007.12.002>
89. Wu, J., Henning, P., Sjögren, K., Koskela, A., Tuukkanen, J., Movérare-Skrtic, S., & Ohlsson, C. (2019). The androgen receptor is required for maintenance of bone mass in adult male mice. *Molecular and Cellular Endocrinology*, 159–169. <https://doi.org/10.1016/j.mce.2018.10.008>
90. Yin, J., Valin, K. L., Dixon, M. L., & Leavenworth, J. W. (2017). The Role of Microglia and Macrophages in CNS Homeostasis, Autoimmunity, and Cancer. *Journal of Immunology Research*, 1–12. <https://doi.org/10.1155/2017/5150678>
91. Zarrouf, F. A., Artz, S., Griffith, J., Sirbu, C., & Kommor, M. (2009). Testosterone and Depression. *Journal of Psychiatric Practice*, 289–305. <https://doi.org/10.1097/01.pra.0000358315.88931.fc>
92. Zhao, R., Wang, X., Jiang, C., Shi, F., Zhu, Y., Yang, B., Zhuo, J., Jing, Y., Luo, G., Xia, S., & Han, B. (2017). Finasteride accelerates prostate wound healing after thulium laser resection through DHT and AR signalling. *Cell Proliferation*, e12415. <https://doi.org/10.1111/cpr.12415>
-

Androgen mediated pleiotropy?

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/androgen-mediated-pleiotropy/>

The case for diverse pathological effects arising from androgen-mediated pleiotropic mechanisms is increasingly clear beyond conditions already discussed such as PCOS and SBMA. The AR and its role in health is a fast-expanding research area of high priority (Takayama, 2017). The largest genome-wide association study to date in AGA research established a statistically significant positive association between AGA and other age and androgen related traits such as bone mineral density and early puberty, supporting a case for an androgen mediated pleiotropy underlying multiple conditions, as proposed by Yap et al (Yap et al., 2018). Considering the common androgenic pathogenesis of both AGA and BPH, Ramsamy et al. found that as the grade of AGA increased, there was an increase in the size of the prostate, with 66.7% of men evaluated experiencing severe AGA having an enlarged prostate (Subramaniyan et al., 2016). AGA patients are more prone to prostate enlargement and related symptoms (Monib et al., 2018).

Pleiotropic epigenetic factors can mediate a multi-system and clinically significant repression of AR expression. AIS type II is a type of Androgen Insensitivity Syndrome that presents clinically without mutation in the AR gene sequence (N. C. Hornig et al., 2016). Fewer than 40% of patients with diagnosed Partial AIS exhibit AR gene mutation, suggesting epigenetic involvement in androgen-insensitive phenotypes without sequence alterations. Hornig et al. recently provided a molecular diagnosis for the clinical presentation of AIS type II. Identifying significant reduction in AR mRNA levels in the genital fibroblasts of 57% of the cases, they additionally demonstrated methylation levels of two CpG sites in the proximal AR promoter region inversely correlated significantly to the expression of AR mRNA expression levels (Nadine C Hornig et al., 2018).

Noting the incomplete understanding of major chronic disease and the advancing understanding of the effects of androgens on major contributors to global mortality including immune function, cancer, cardiovascular disease and diabetes, Schooling considered the potential for androgens to be considered in a pleiotropic context to explain the higher vulnerability to disease mortality and earlier death observed in males than women. She suggests that "considering androgens as potential contributors to major diseases represents a major paradigm shift that flies in the face of individual level data from observational studies", and that a "rethink of the role of androgens, particularly, in immune function, cancer and cardiovascular disease, as potentially providing an underlying explanatory mechanism that could address the noted sex disparity in life expectancy, help identify new specific targets of intervention, explain unexpected side effects of commonly used drugs and eventually provide targets for precision medicine" (Schooling, 2015). In this context, PFS is likely to provide novel insights and considerable translational benefits to wider biological understanding and of mechanistic factors in better-recognised disease states.

Page Bibliography

1. Hornig, N. C., Ukat, M., Schweikert, H. U., Hiort, O., Werner, R., Drop, S. L. S., Cools, M., Hughes, I. A., Audi, L., Ahmed, S. F., Demiri, J., Rodens, P., Worch, L., Wehner, G., Kulle, A. E., Dunstheimer, D., Müller-Roßberg, E., Reinehr, T., Hadidi, A. T., ... Holterhus, P.-M. (2016). Identification of an AR Mutation-Negative Class of Androgen Insensitivity by Determining Endogenous AR Activity. *The Journal of Clinical Endocrinology & Metabolism*, 4468–4477. <https://doi.org/10.1210/jc.2016-1990>
2. Hornig, Nadine C, Rodens, P., Dörr, H., Hubner, N. C., Kulle, A. E., Schweikert, H.-U., Welzel, M., Bens, S., Hiort, O., Werner, R., Gonzalves, S., Eckstein, A. K., Cools, M., Verrijn-Stuart, A., Stunnenberg, H. G., Siebert, R., Ammerpohl, O., & Holterhus, P.-M. (2018). Epigenetic Repression of Androgen Receptor Transcription in Mutation-Negative Androgen Insensitivity Syndrome (AIS Type II). *The Journal of Clinical Endocrinology & Metabolism*, 4617–4627. <https://doi.org/10.1210/jc.2018-00052>
3. Monib, K. M. E., Hussein, M. S., & Kandeel, W. S. (2018). The relation between androgenetic thin hair diagnosed by trichoscope and benign prostatic hyperplasia. *Journal of Cosmetic Dermatology*, 1502–1506. <https://doi.org/10.1111/jocd.12835>
4. Schooling, C. M. (2015). Could androgens be relevant to partly explain why men have lower life expectancy than women? *Journal of Epidemiology and Community Health*, 324–328. <https://doi.org/10.1136/jech-2015-206336>
5. Subramaniyan, R., Ramsamy, K., & Patra, A. (2016). An observational study of the association between androgenetic alopecia and size of the prostate. *International Journal of Trichology*, 62. <https://doi.org/10.4103/0974-7753.188034>
6. Takayama, K. (2017). The biological and clinical advances of androgen receptor function in age-related diseases and cancer [Review]. *Endocrine Journal*, 933–946. <https://doi.org/10.1507/endocrj.ej17-0328>
7. Yap, C. X., Sidorenko, J., Wu, Y., Kemper, K. E., Yang, J., Wray, N. R., Robinson, M. R., & Visscher, P. M. (2018). Dissection of genetic variation and evidence for pleiotropy in male pattern baldness. *Nature Communications*. <https://doi.org/10.1038/s41467-018-07862-y>

Current situation is dangerous

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/current-situation-is-dangerous/>

An urgent clinical failure

With [humility] comes not only reverence for truth, but also proper estimation of the difficulties encountered in our search for it. ...[T]his grace of humility is a precious gift.

William Osler, Aequanimitas: with other addresses to medical students, nurses and practitioners of medicine, 1849-1919

The stigma associated with sexual and mental dysfunctions, as well as a lack of medical support, are causing PFS to become a hidden epidemic. The scale and human cost of this failure cannot solely be attributed to pharmaceutical manufacturers. Rather, it is the result of a systemic clinical approach to ADRs that is not fit for purpose when considering a disease that manifests or progresses following withdrawal without a known biomarker. The staunch resistance patients continue to face in attempts to establish the very existence of the condition does not stem solely from the significant financial interest in antiandrogenic substances as first-line treatments in dermatology, but its ostensible implausibility given a remarkable reality and broad clinical endpoints. A perfect storm of novelty, rarity, and counter-intuitive clinical presentation compound clinical, pharmaceutical and regulatory failures to entrench a situation in which internet resources such as propeciahelp represent the only support for patients suffering profoundly following exposure to antiandrogenic endocrine disruptors. As symptoms vary between patients from moderate functional impairments to a life-threatening physiological and neuropsychological breakdown, this is an unsustainable situation that cannot continue.

Disturbingly, clinicians appear significantly more likely to report an ADR resulting from 5ari therapy in older men typically prescribed finasteride 5mg, despite the ADRs in this group being fewer and less frequently associated with lasting disability. Considering FAERS adverse event reports in the period April 2011 to October 2014, a significant majority of ADRs resulting from use of 1mg Finasteride by younger men were self-reported to the FDA despite a higher reported incidence of disability. Contrastingly, most side effects in older patients were reported by their doctors (Baas et al., 2018)?. With consideration to the alarming dissatisfaction amongst PFS patients with regards to clinical care reported by Ganzer et al. (Ganzer et al., 2014)?, this could indicate a widespread dismissal at the clinical level due to an erroneous assumption that patients' symptoms are not possible and/or psychosomatic in

nature. This would reflect the ubiquitous dissatisfaction of patients describing their experiences seeking help from primary care physicians and the ostensibly appropriate specialists in fields to which their symptomatology can generally be associated including urology and psychology. This deters patients with already stigmatising problems from professional engagement. This is extremely serious, as pharmacovigilant entities including the European Medicines Agency rely upon doctors to submit adverse event reports when reported by patients. Traish suggested that a misleading narrative that the condition does not exist has arisen from the current dearth of awareness and knowledge in the clinical community (Traish, 2018) despite the body of literature suggesting epigenetic susceptibility in a subset of consumers (Traish, 2020). He notes that patients are frustrated by the perception in the medical community that such condition does not exist and that they are labelled to suffer from psychological disorder, rather than an organic disorder, attributed to the inhibition of a key biochemical pathway in steroid biosynthesis and metabolism. Traish suggests this, along with the lack of attention to improve care for afflicted patients, has "translated into loss of credibility and confidence by patients in their doctors and huge loss of faith in the medical community at large" (Traish, 2018).

As of 2020, the status quo in frontline care is presenting a perilous circumstance to both existing PFS patients and the wider public. Awareness of PFS as a clinical entity is unacceptably poor amongst the medical profession and education is urgently needed (Garreton et al., 2016; Traish, 2020). Failure to acknowledge the novelty and clinical scope of the pathology continues to delay progress towards etiological understanding. The fact that Vice media have demonstrated a deeper understanding of the post-withdrawal "crash" than medical literature is a matter of concern (Morgans, 2018). The consequence of this void in clinical understanding has not only led to a lack of basic science, but the potential for PFS patients to be prescribed therapies that can result in additional and permanent harm, including SSRI medications. It is deeply concerning that, instead of psychological support being offered as adjunctive care alongside appropriate recognition of what is a serious physiological disorder, doctors are frequently issuing rapid and inappropriate psychosomatic diagnosis for what is nearly always a striking and clear description of health problems never before experienced by the patient following taking and ceasing Finasteride. Healy et al. note that this is similarly the experience for patients suffering persistently after SSRI antidepressant use, commenting that even though patients report normal sexual function prior to use and neither depression nor anxiety can account for symptomatic presentations, "physicians appear to default to attributing problems a patient has after treatment to manifestations of an underlying nervous diathesis" (David Healy et al., 2018). This is unacceptable and unjustifiable given how deeply complex the issue is and how much there is yet to know regarding the physiological consequences of endocrine disruption with 5 α is (Traish et al., 2015).

Psychosomatic misdiagnosis has, in extreme cases, caused patients to be deprived of their liberty through admission to psychiatric institutions. Patients have expressed feeling intense fear after being pressured into taking psychiatric drugs that have had a profound negative impact their condition. Routinely, additional stress, confusion and harm is caused to those suffering extreme symptoms by what is tantamount to "gaslighting" (Thomas, 2018) by clinicians and psychologists. The combination of clinical arrogance and ignorance is egregious and difficult to excuse at this stage. Maksym concluded that the lasting consequences of antiandrogen therapy on the organism remain obscure, and can be highly

complex and multilateral, noting the extensive metabolism of steroid hormones in the central nervous system. They state that the presence of severe and persistent effects caused by the treatment of an aesthetic issue raises great concern for the clinician given the widespread use in young and healthy individuals, and that the low estimated prevalence of PFS cannot excuse nonvigilance (Maksym et al., 2019)?.

Those PFS patients who are most severely affected are those who are most vulnerable to these systemic failings. Many patients are very young, and young men left unable to function socially, work or continue their studies due to debilitating physiological and neurological symptoms can be left reliant on support from family and friends who cannot always understand or appreciate the etiology of their behavioural changes. Those around the patient will understandably defer to professional assessment, and simplistic misattribution is frequently the outcome. When physiological processes far beyond the patient's control are responsible, this psychosomatic misattribution by those in positions of medical authority unfamiliar with PFS or literature regarding the condition can often have devastating interpersonal consequences for patients already in an unimaginably desperate situation. The potential etiological overlap between the recognised persistent syndrome occurring rarely with serotonergic treatment and PFS is an emergent consideration in medical literature (David Healy et al., 2018; Giatti et al., 2018)?. Importantly, recent research has identified profound interruption of the androgen steroid pathway by SSRI antidepressants (Griffin & Mellon, 1999; Hansen et al., 2017; Jacobsen et al., 2015; Munkboel et al., 2018)?. In context of anecdotal reports from PFS patients of significant worsening following exposure to serotonergic drugs, an extremely cautious approach should therefore be taken when considering prescription of serotonergic medications to patients reporting enduring health problems not experienced prior to finasteride use.

Recognising a consistent and concerning failure in the clinical care of our patients, we issued Post-Finasteride patients the Short Assessment of Patient Satisfaction, a robust measure of patient satisfaction with their experience in clinical practice (Hawthorne et al., 2014)? as part of a wider survey. Patients were asked to complete the assessment once if they had seen only one professional with regards to PFS. If they had seen more than one clinical professional about PFS, we asked them to complete the questionnaire twice: Once considering their most positive experience with regards to an appointment, and once considering their most negative experience. After 170 submissions, the results were remarkable and alarming. The average score regarding even the most positive experiences PFS patients have had with a clinical appointment is on the verge between dissatisfaction and serious dissatisfaction, denoting that “severe and urgent failings” are the norm for PFS patients seeking healthcare support, and that the very best they can hope for is a dissatisfactory clinical outcome (Propeciahelp Post Drug Syndrome Survey: Data not provided). We will seek to publish this data in the future.

Disappointingly, PFS represents a neglected opportunity to broaden scientific understanding of biological mechanisms critical to human health and will undoubtedly bridge identified knowledge gaps in the understanding of endocrine disruption (Solecki et al., 2016)?. As well as a virtue, professional humility is important to being a good doctor (Chou et al., 2014; DuBois et al., 2013; Mahant et al., 2012; Wear,

2008)?, and the vast anecdotal experience of our patients attests to a widespread shortcoming in this regard. That physicians commonly deem what has happened to those suffering PFS as implausible or impossible is telling as to the biological significance of this disease. In his commentary stressing the importance of humility in medical professionals and scientists to avoid future harms, Ritterman notes that the "problem of mistaken ideas persisting despite scientific evidence to the contrary has been present since the onset of the scientific method...This problem is of particular concern in medical science, where outmoded ideas translate into excess morbidity and mortality" (Ritterman, 2017)?. What differentiates our remarkable situation from examples of historic medical ignorance such as this is that, in 2020, there exists compelling objective evidence which can be contextualised, as we have attempted to the best of our ability, in a broader framework of biological understanding. It is now abundantly clear that the androgen pathway has critical roles across the entire organism, and that understanding of the implications of this on health has expanded rapidly. With so much yet to be elucidated, and such profound effects described by a subpopulation of consumers for years, the arrogance faced by our patients when reporting their drug-induced symptoms is impossible to justify. "If the toxin is professional arrogance," Ritterman wrote, "the antidote is professional humility".

In the absence of the acknowledgement of the true scope of the condition, informed consent to the risk of PFS is never obtained from AGA patients commencing Finasteride therapy. Demand for - and marketing of - antiandrogenic hair loss remedies such as Finasteride is expanding, and an inevitable consequence will be more cases of PFS. As of 2020, emergent subscription services are engaging in social media advertising campaigns with modern production values. Hims present a video of young woman in a lab coat visibly laughing while saying that "anything (sic) can write anything on google". Another woman, also wearing a lab coat, assures consumers that "fewer than 1% of men actually experience side effects, but don't be scared; this happens to very few men, and we're here to help you if it does" (Hims, 2018)?. What that help consists of is difficult to infer, considering we nor professors engaged with the issue in the fields of neuroendocrinology, urology, andrology, steroid biology and psychology seeking an explanation as to this breakdown of expected function in the androgen pathway are aware of any effective and safe treatment. Manual are an internet-based prescription company who at time of writing advertise on social networks including the image sharing service Snapchat. They state on a web page intending to answer frequent questions about Finasteride that "Animal studies did not show negative effects on fertility." (Manual, 2019)?. As we have previously discussed, animal studies have entailed a deficiency in fertility parameters that is transgenerational (Garcia et al., 2012; Kolasa-Wo?osiuk et al., 2019)?. We are already receiving new PFS patients citing having taken finasteride after receiving the marketing of the companies mentioned. Considering increasing primary objective evidence in study of PFS patients, the multitude of deleterious molecular level effects in animal research and the numerous reviews stating that this is a rare and distinct clinical entity, those promoting Finasteride as a safe product for young men without warning of PFS can easily be likened to the tobacco executives of the 1980s. As public appreciation grew of the dangers associated with smoking, advertisement campaigns designed to obfuscate reality, including a smoker depicted to be saying "Please don't tell me my cigarette smoke is harmful to you. There's just no convincing proof that it is" (?United States v. Philip Morris USA Inc.?, 2006).

Certain dermatologists remain opposed to acknowledgement of what is physiologically happening to a

subset of consumers after taking finasteride. In a report of a single AGA patient without a depressive history who presented with sexual dysfunction following Finasteride withdrawal, Trüeb et al. presented a hypotheses that PFS is a "delusional disorder" (Trüeb et al., 2019). Trüeb suggests that the airing of a documentary on Swiss television may have had a psychosomatic influence on this patient and hypothesise the condition to be one of a "mass hysteria". The authors define mass hysteria as many people believing "obviously false and potentially distressing things based purely on hearsay". By their own definition, this does not apply to PFS considering basic science and animal research, as well as the outcomes of several case-controlled studies of PFS patients, which are not addressed. ADR data additionally refutes this suggestion: Ali et al. had previously considered the potential for bias due to stimulated reporting of persistent sexual dysfunction. Analysing FAERS adverse event data, Ali et al. acknowledged an increase in ADRs, but noted that significant signals with a 95% confidence interval lower limit of 2.0 or greater exists before and after 2011, irrespective of the public's knowledge of sexual dysfunction as a safety concern associated with finasteride. Ali et al. considered underreporting likely and the actual incidences of persistent sexual dysfunction to be potentially underestimated (Ali et al., 2015). Based upon their hypothesis and without any reported success in remediating the symptoms of their single patient, Trüeb et al. encourage the prescription of psychiatric medicines for an "underlying psychopathological disorder". First line treatments for depression ordinarily fail and have commonly worsened PFS patients as we have discussed. SSRIs are frequently antiandrogenic (Hansen et al., 2017; Jacobsen et al., 2015; Munkboel et al., 2018) and are associated with a remarkably similar persistent cognitive, physiological and sexual dysfunction with the potential to represent a single syndrome (David Healy et al., 2018). As such, this baseless and irresponsible recommendation is not without the potential to result in harm to profoundly vulnerable patients should it influence clinical practitioners. It is interesting, however, that even in a commentary seeking to cast doubt on the existence of PFS as an organic condition, there is some awareness of a key novelty of the syndrome common to self-reports: The subsequential nature of onset or intensification frequently featured in patient reports (Trüeb et al., 2019). The author declares no conflict of interest despite stating that his private hair clinic continues to prescribe the drug after two decades of doing so.

Patient driven platforms cannot compensate for clinical disregard

While propeciahelp continues to provide as much support to patients as is feasible, this is a serious medical problem and patient-operated support platforms cannot possibly compensate for the entrenched failures in clinical practice that both patients and medical literature continue to highlight. Maksym et al. recognise the variable reporting in different healthcare settings is making the problem hard to evaluate (Maksym et al., 2019). Patients who have suddenly stopped visiting the propeciahelp forum are impossible to account for due to anonymity. Failure to achieve diagnosis of this syndrome and improper clinical inquiry means valuable medical records and investigation are usually non-existent or cannot be pursued in context. This is especially serious in the cases of those experiencing extremely severe and degenerative health problems, who can disappear suddenly and untraceably after expressing suicidality due to the extent of their symptoms. Clinical appreciation of PFS must be improved to provide patients accurate diagnosis and ensure the proper contextual documenting of patients with appropriate follow-up.

As severely affected patients are regularly left unable to work due to resultant disabilities, the lack of professional recognition is hampering their ability to receive much-needed financial support from welfare systems. Post-mortem study will likely be extremely beneficial to a mechanistic understanding of the induced epigenetic changes in PFS, and this is dependent on appropriate diagnosis and clinical profiling.

Clinical disregard and a dire need for hope compounds the potential for the exploitation of a vulnerable and often desperate cohort by individuals or businesses offering simplistic explanations and suggesting treatments. The risk of additional harms resulting from self-medication in attempts to relieve debilitating symptoms is significant, particularly amongst the worst affected. The inability of specialist doctors to provide answers or symptomatic relief drives some patients to embark upon self-experimentation. Patients will commonly express belief that supradietary doses of concentrated "natural" extracts, vitamins or minerals have a preferential safety profile as compared with that of pharmaceutical drugs in the attempted alleviation of PFS symptoms. In patients that can exhibit a novel fragility to any further disruption of the androgen pathway, therapeutic attempts with both clinically prescribed pharmaceuticals and self-sought nutraceuticals have led to permanent worsening and directly preceded completed suicide. It is urgent and imperative that clinicians presented with PFS patients inform the patient of a physiological vulnerability to substances with endocrine disruptive properties. This is particularly important for severely affected cases of PFS who present following a short exposure to Finasteride or other causative antiandrogenic substance.

A desperate need for improvement often results in a significant selection bias on the part of patients when considering other patient reports. This can often involve a rejection of the complex situation in favour of alternative health or pseudoscientific concepts. Strong views and poorly defined etiological conclusions can be rapidly formed. Significant heterogeneity in clinical endpoints results in many patients having a poor appreciation of the situation for other patients, or as indicative of a vast array of etiologically distinct disease states. A well-known parable describes a group of blind men touching an elephant. Grasping the tusk, one believes it to be a spear. Another touching its leg is sure it is a tree. A third man near the trunk asserts it is a snake, while the man touching its ear believes it to be a fan, and the tale concludes with vehement arguments based on a selective perception (Snyder & Ford, 1987). This analogy is appropriate and can be well observed. Mild to moderately affected patients can find reading the experiences and clinical condition of severely affected patients to be psychologically difficult, potentially owing to uncertainty surrounding their own prognoses. This contributes to an incohesive community and increases the difficulty of representing the true scale of the issue.

Propeciahelp's volunteer staff, who are suffering ourselves, are placed in a deeply difficult situation in which we must constantly advise patients to be wary of theoretical proclamations and treatment suggestions online, while being aware that clinicians cannot currently provide practical help and often present an equal risk to PFS patients. It is our opinion that there will be serious questions to be answered in the future as to why such obstacles were faced in the clinical acknowledgement of a disease as deeply serious and biologically significant as Post-Finasteride Syndrome.

Regulatory activity is overdue

Reappraisal of the use cases for these substances is necessitated at the regulatory level, ensuring adequate warning and mandating informed consent as to the potential of developing this condition. PFS has no known predictive factors, unpredictable severity between patients, a complete absence of dose-dependence, and no available therapeutic options for any of the affected symptomatic domains. PFS does not conform to the presentation of known disease state and consumers are not placed to imagine the potential implications on their physiology, minds, and lives; an impact which cannot be overstated and is not currently widely appreciated at the clinical level. PFS patients almost invariably express shock and disbelief at what is happening to them. In this regard we support the conclusion of Motofei et al. insofar as the patient must be informed and consent to the full potential health risk. Motofei notes that this is especially important in aesthetic treatment, as therapy of AGA with dutasteride places treatment of an aesthetic condition on the same level as a life-threatening disease (Motofei et al., 2019). In the rare instance a consumer were so psychologically distraught by hair loss they would countenance the risk of irreparable physical damage and the permanent loss of sexual, neurological and physiological function, there is simply no excuse for the current situation in which consumers are not informed that this disease even exists as novel clinical entity with all it entails *per se*. Measures to address this must begin now. In particular, the dermatology profession should at a minimum address failure in assessing patient's pre-treatment conditions, pursue a fully informed consent, and begin effective reporting of adverse events according to national and supranational guidelines. As we are discussing frontline drugs of the dermatology profession that represent a significant worldwide revenue, at this late stage we pragmatically recognise that regulatory action will have to precede any widespread self-initiated clinical responsibility. A simple truth can be represented by the words of Upton Sinclair: "It is difficult to get a man to understand something, when his salary depends upon his not understanding it" (Sinclair, 1994).

A good therapy should have tissue selectivity to the pathogenesis and not broadly interfere with other important processes in humans (Zheng et al., 2006). Finasteride interferes with fundamental and ubiquitous physiological processes (Traish, 2020), with PFS manifesting in some consumers as a disastrous and permanent result of this. Many scientific insights into the critical role of androgens across the body and brain were not appreciated at the time of its approval. While there is now significant post-marketing evidence and animal research illustrating the systemic influence and thus potential danger of finasteride, the evidentiary basis for its continued presentation as safe product is not robust (Belknap et al., 2015). Adverse reaction warnings in the product leaflet of Finasteride remain direly inadequate and do not include PFS as a distinct entity, do not mention the post-withdrawal development seen in the majority of PFS cases, nor most of the multi-systemic symptoms PFS entails. These are well recognised in publications centring on patient reports, as we have discussed, and it is noted that despite this evidence the leaflet continues to make little mention of the broad symptom profile (Walf et al., 2018). It is therefore unacceptable that consumers are still presented with a wolf in sheep's clothing. PFS is not imaginable by those thankfully able to take what it can strip away for granted: The emotional, physical

and intellectual joys of human experience, and, often, even the ability earn a subsistence income. Patisaul and Belcher suggest that, when considering risk from EDCs, the human brain performs a risk assessment as it would in anything else: "Using an imperfect calculus incorporating intuition, experience, a mix of facts (and more often fiction) combined with something like raw gut instinct", generally favouring short term benefit over the possibility of long term harm (Patisaul & Belcher, 2017)?. This "common sense" risk calculus is not adequate in the absence of accurate information, as it is novel amongst adverse drug reactions and astonishingly counter-intuitive. It is wholly unreasonable to presume consumers are placed to consider such an outcome as even the remotest possibility, particularly in absence of their doctors nor the product labelling making it abundantly clear that men are experiencing horrendous and progressive changes to the physiological structure and function of their bodies and minds as a result of taking as little as one tablet.

It cannot be emphasised strongly enough that we are primarily discussing finasteride prescribed as a cosmetic product. It is our strong contention that members of the public will rightly consider such an indication to be held to a considerably higher bar of safety than drugs for serious medical conditions, yet the de facto reality speaks to the opposite. Review after review now acknowledges evidential support for the existence of PFS and therefore the need for adequate consent to the potential risk to health and quality of life upon prescription of finasteride (Irwig, 2015; Maksym et al., 2019; Motofei et al., 2019; Said & Mehta, 2018; Than et al., 2018; Traish, 2020)?. In the absence of accurate clinical communication of the full risk to the patient, informed consent can never currently be obtained. Multidisciplinary scientific conclusion is not reflected in pharmacovigilant activity. The status quo begs the question: What precisely will it take to achieve the most basic of protections for the public? Discussing regulatory action in regards to endocrine disruptors for the protection of human health in their 2012 guidance to decision makers, the WHO describe that the 1973 United States ban on tetraethyl lead in gasoline followed "decades of inaction" during which children were continually exposed to a serious health risk. They suggest that "perhaps the answer is in making more use of the precautionary principle to ban or restrict chemicals in order to reduce exposure early, even when there are significant but incomplete data and before there is significant and long-lasting harm" (Bergman et al., 2012)?. Significant data exists with which conclude Finasteride is inestimably dangerous in a subpopulation, causes permanent harm, and that this is not positively correlated to duration of use. This justifies its withdrawal from sale as a cosmetic. Rosario and Bourke, discussing underappreciated cardiovascular risk associated with modern antiandrogen treatments in prostate cancer, suggest that in an era of media soundbytes and "wonder drugs" that men will insist on, the scientific community must respond and remain circumspect, with regulatory bodies, trial oversight committees, reviewers and editors having a duty of care "to ensure the correct health warnings go out alongside the positive messages" (Rosario & Bourke, 2020)?. If this is deemed the necessary response in the treatment of life threatening cancer, it is unfathomable that this vigilance should not be all the more appropriate to the prescription of antiandrogens to young healthy men for conditions like AGA and acne - conditions which are very mild in terms of androgen-mediated pathologies (Heemers & Tindall, 2007)?. Wolfgang Becker-Brüser, editor in chief of the German medical journal *Arznei-Telegramm*, recently stated that the "very serious side effects caused by finasteride [are] absolutely unacceptable for a lifestyle drug. Rationally, one cannot advocate for this medicine or justify the fact that it's still on the market. Actually, it should be banned" (Südwestrundfunk, 2019)?.

Some authors continue to call for repeated placebo-controlled trials to determine the existence of persistent effects from finasteride, considering little else to be of sufficient evidential quality (Basaria et al., 2016; Diviccaro et al., 2020; Gray & Semla, 2019)?. Further study of this kind will not be enlightening nor practically useful to the scientific community, the PFS patient or consumer in the medium term, if ever. It is urgently necessary to acknowledge both the novel nature of the condition and the rarity of the syndrome (Traish, 2018)? for a pragmatic approach. Considering the medical history of 6 PFS patients who committed suicide, Irwig noted that a prospective study that may determine causality would likely require at least 10,000 participants in each arm and a duration of at least 5 years, making it practically and financially unfeasible (Irwig, 2020)?. It is very possible that even in such a trial, occurrence of PFS would not reach signal. Rarity, however, cannot and should not be construed to justify dismissal of the gravity of this condition (Maksym et al., 2019)?. This is particularly relevant with consideration to the unpredictability and dose-independence of PFS, and its atypical progression following withdrawal. Dismissal of retrospective studies is often attempted owing to a perceived lack of credibility in normal instances of ADR. This disease is not a normal ADR, to the point that existing drug reaction algorithms are unable to accommodate it (David Healy et al., 2018)?. As well as statistical rarity, objective differences at the molecular level in control study of patients are increasingly established in PFS. A pragmatic approach to any progress must take the reality of this issue into account, not defer to an arbitrary standard of perceived evidential quality appropriate to a more ordinary adverse drug reaction while patients continue to be driven to suicide by profound and unresolvable suffering. Insistence on the application of a formula that is not fit for purpose in this circumstance manifests as a dereliction of duty. This will be at the expense of lives that could be saved by the most basic of warnings. It is astonishing to consider that mechanistic elucidation may now plausibly precede acknowledgement of a syndrome that has been clinically reported by patients for two decades. Patients cannot continue to shoulder this global problem in lieu of clinicians.

Page Bibliography

1. Ali, A. K., Heran, B. S., & Etminan, M. (2015). Persistent Sexual Dysfunction and Suicidal Ideation in Young Men Treated with Low-Dose Finasteride: A Pharmacovigilance Study. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 687–695. <https://doi.org/10.1002/phar.1612>
2. Baas, W. R., Butcher, M. J., Lwin, A., Holland, B., Herberts, M., Clemons, J., Delfino, K., Althof, S., Kohler, T. S., & McVary, K. T. (2018). A Review of the FAERS Data on 5-Alpha Reductase Inhibitors: Implications for Postfinasteride Syndrome. *Urology*, 143–149. <https://doi.org/10.1016/j.urology.2018.06.022>
3. Basaria, S., Jasuja, R., Huang, G., Wharton, W., Pan, H., Pencina, K., Li, Z., Travison, T. G., Bhawan, J., Gonthier, R., Labrie, F., Dury, A. Y., Serra, C., Papazian, A., O’Leary, M., Amr, S., Storer, T. W., Stern, E., & Bhasin, S. (2016). Characteristics of Men Who Report Persistent Sexual Symptoms After Finasteride Use for Hair Loss. *The Journal of Clinical Endocrinology &*

Metabolism, 4669–4680. <https://doi.org/10.1210/jc.2016-2726>

4. Belknap, S. M., Aslam, I., Kiguradze, T., Temps, W. H., Yarnold, P. R., Cashy, J., Brannigan, R. E., Micali, G., Nardone, B., & West, D. P. (2015). Adverse Event Reporting in Clinical Trials of Finasteride for Androgenic Alopecia. *JAMA Dermatology*, 600. <https://doi.org/10.1001/jamadermatol.2015.36>
5. Bergman, Å., Heindel, J., Jobling, S., Kidd, K., & Zoeller, R. T. (2012). State-of-the-science of endocrine disrupting chemicals, 2012. *Toxicology Letters*, S3. <https://doi.org/10.1016/j.toxlet.2012.03.020>
6. Chou, C. M., Kellom, K., & Shea, J. A. (2014). Attitudes and Habits of Highly Humanistic Physicians. *Academic Medicine*, 1252–1258. <https://doi.org/10.1097/acm.0000000000000405>
7. David Healy, Joanna Le Noury, & Derelie Mangin. (2018). Enduring sexual dysfunction after treatment with antidepressants, 5 α -reductase inhibitors and isotretinoin: 300 cases. *International Journal of Risk & Safety in Medicine*, 125–134. <https://doi.org/10.3233/JRS-180744>
8. Diviccaro, S., Melcangi, R. C., & Giatti, S. (2020). Post-finasteride syndrome: An emerging clinical problem. *Neurobiology of Stress*, 100209. <https://doi.org/10.1016/j.ynstr.2019.100209>
9. DuBois, J. M., Kraus, E. M., Mikulec, A. A., Cruz-Flores, S., & Bakanas, E. (2013). A Humble Task. *Academic Medicine*, 924–928. <https://doi.org/10.1097/acm.0b013e318294fd5b>
10. Ganzer, C. A., Jacobs, A. R., & Iqbal, F. (2014). Persistent Sexual, Emotional, and Cognitive Impairment Post-Finasteride. *American Journal of Men's Health*, 222–228. <https://doi.org/10.1177/1557988314538445>
11. Garcia, P. V., Barbieri, M. F., Perobelli, J. E., Consonni, S. R., Mesquita, S. de F. P., Kempinas, W. de G., & Pereira, L. A. V. (2012). Morphometric-stereological and functional epididymal alterations and a decrease in fertility in rats treated with finasteride and after a 30-day post-treatment recovery period. *Fertility and Sterility*, 1444–1451. <https://doi.org/10.1016/j.fertnstert.2012.03.025>
12. Garreton, A. S., Valzacchi, G. R., & Layus, O. (2016). Post-Finasteride Syndrome: About 2 Cases and Review of the Literature. *Andrology-Open Access*. <https://doi.org/10.4172/2472-1212.1000170>

13. Giatti, S., Diviccaro, S., Panzica, G., & Melcangi, R. C. (2018). Post-finasteride syndrome and post-SSRI sexual dysfunction: two sides of the same coin? *Endocrine*, 180–193.
<https://doi.org/10.1007/s12020-018-1593-5>
14. Gray, S. L., & Semla, T. P. (2019). Post-finasteride syndrome. *BMJ*, 15047.
<https://doi.org/10.1136/bmj.15047>
15. Griffin, L. D., & Mellon, S. H. (1999). Selective serotonin reuptake inhibitors directly alter activity of neurosteroidogenic enzymes. *Proceedings of the National Academy of Sciences*, 13512–13517. <https://doi.org/10.1073/pnas.96.23.13512>
16. Hansen, C. H., Larsen, L. W., Sørensen, A. M., Halling-Sørensen, B., & Styrihave, B. (2017). The six most widely used selective serotonin reuptake inhibitors decrease androgens and increase estrogens in the H295R cell line. *Toxicology in Vitro*, 1–11.
<https://doi.org/10.1016/j.tiv.2017.02.001>
17. Hawthorne, G., Sansoni, J., Hayes, L., Marosszeky, N., & Sansoni, E. (2014). Measuring patient satisfaction with health care treatment using the Short Assessment of Patient Satisfaction measure delivered superior and robust satisfaction estimates. *Journal of Clinical Epidemiology*, 527–537.
<https://doi.org/10.1016/j.jclinepi.2013.12.010>
18. Heemers, H. V., & Tindall, D. J. (2007). Androgen Receptor (AR) Coregulators: A Diversity of Functions Converging on and Regulating the AR Transcriptional Complex. *Endocrine Reviews*, 778–808. <https://doi.org/10.1210/er.2007-0019>
19. Hims. (2018, November 7). *Ask hims: Can I have sex while taking finasteride?* [Youtube Video]. YouTube. <https://www.youtube.com/watch?v=KU8ciNr0GfY>
20. Irwig, M. S. (2015). Safety concerns regarding 5 α reductase inhibitors for the treatment of androgenetic alopecia. *Current Opinion in Endocrinology & Diabetes and Obesity*, 248–253.
<https://doi.org/10.1097/med.0000000000000158>
21. Irwig, M. S. (2020). Finasteride and Suicide: A Postmarketing Case Series. *Dermatology*, 1–6.
<https://doi.org/10.1159/000505151>

22. Jacobsen, N. W., Hansen, C. H., Nellemann, C., Styrihave, B., & Halling-Sørensen, B. (2015). Effects of selective serotonin reuptake inhibitors on three sex steroids in two versions of the aromatase enzyme inhibition assay and in the H295R cell assay. *Toxicology in Vitro*, 1729–1735. <https://doi.org/10.1016/j.tiv.2015.07.005>
23. Kolasa-Woźosiuk, A., Tarnowski, M., Baranowska-Bosiacka, I., Chlubek, D., & Wiszniewska, B. (2019). Antioxidant enzyme expression of mRNA and protein in the epididymis of finasteride-treated male rat offspring during postnatal development. *Archives of Medical Science*, 797–810. <https://doi.org/10.5114/aoms.2017.68528>
24. Mahant, S., Jovcevska, V., & Wadhwa, A. (2012). The Nature of Excellent Clinicians at an Academic Health Science Center. *Academic Medicine*, 1715–1721. <https://doi.org/10.1097/acm.0b013e3182716790>
25. Maksym, R., Kajdy, A., & Rabijewski, M. (2019). Post-finasteride syndrome - does it really exist? *The Aging Male : The Official Journal of the International Society for the Study of the Aging Male*, 22(4), 250–259. <https://doi.org/10.1080/13685538.2018.1548589>
26. Manual. (2019). *Finasteride: Questions answered*. Manual.Co. <https://www.manual.co/hair-loss/finasteride>
27. Morgans, J. (2018, April 24). *I Need to Quit Hair Loss Drugs Before They Kill Me*. Vice. https://www.vice.com/en_uk/article/43bm3m/i-need-to-quit-hair-loss-drugs-before-they-kill-me
28. Motofei, I. G., Rowland, D. L., Tampa, M., Sarbu, M.-I., Mitran, M.-I., Mitran, C.-I., Stoian, A. P., Diaconu, C. C., Paunica, S., & Georgescu, S. R. (2019). Finasteride and androgenic alopecia; from therapeutic options to medical implications. *Journal of Dermatological Treatment*, 1–7. <https://doi.org/10.1080/09546634.2019.1595507>
29. Munkboel, C. H., Larsen, L. W., Weisser, J. J., Møbjerg Kristensen, D., & Styrihave, B. (2018). Sertraline Suppresses Testis and Adrenal Steroid Production and Steroidogenic Gene Expression While Increasing LH in Plasma of Male Rats Resulting in Compensatory Hypogonadism. *Toxicological Sciences*, 609–619. <https://doi.org/10.1093/toxsci/kfy059>
30. Patisaul, H. B., & Belcher, S. M. (2017). Endocrine Disruptors, Brain, and Behavior. In *Oxford Scholarship Online*. Oxford University Press. <https://doi.org/10.1093/acprof:oso/9780199935734.001.0001>

31. Ritterman, J. (2017). To Err is Human: Can American Medicine Learn from Past Mistakes? *The Permanente Journal*. <https://doi.org/10.7812/tpp/16-181>
32. Rosario, D. J., & Bourke, L. (2020). Cardiovascular Disease and the Androgen Receptor: Here We Go Again? *European Urology*, 167–169. <https://doi.org/10.1016/j.eururo.2019.08.017>
33. Said, M. A., & Mehta, A. (2018). The Impact of 5 α -Reductase Inhibitor Use for Male Pattern Hair Loss on Men's Health. *Current Urology Reports*. <https://doi.org/10.1007/s11934-018-0814-z>
34. Sinclair, U. (1994). *I, Candidate for Governor: And How I Got Licked*. University of California Press .
35. Snyder, C. R., & Ford, C. E. (Eds.). (1987). *Coping with Negative Life Events*. Springer US. <https://doi.org/10.1007/978-1-4757-9865-4>
36. Solecki, R., Kortenkamp, A., Bergman, Å., Chahoud, I., Degen, G. H., Dietrich, D., Greim, H., Håkansson, H., Hass, U., Husoy, T., Jacobs, M., Jobling, S., Mantovani, A., Marx-Stoelting, P., Piersma, A., Ritz, V., Slama, R., Stahlmann, R., van den Berg, M., ... Boobis, A. R. (2016). Scientific principles for the identification of endocrine-disrupting chemicals: a consensus statement. *Archives of Toxicology*, 1001–1006. <https://doi.org/10.1007/s00204-016-1866-9>
37. Südwestrundfunk. (2019). *Finasterid: Tote Hose statt kahler Kopf?* SWR. <https://www.swrfernsehen.de/marktcheck/Finasterid-Tote-Hose-statt-kahler-Kopf.av-01121495-100.html>
38. Than, J. K., Rodriguez, K., & Khera, M. (2018). Post-finasteride Syndrome: A Review of Current Literature. *Current Sexual Health Reports*, 152–157. <https://doi.org/10.1007/s11930-018-0163-4>
39. Thomas, L. (2018). Gaslight and gaslighting. *The Lancet Psychiatry*, 117–118. [https://doi.org/10.1016/s2215-0366\(18\)30024-5](https://doi.org/10.1016/s2215-0366(18)30024-5)
40. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>

41. Traish, A. M. (2020). Post-finasteride syndrome: a surmountable challenge for clinicians. *Fertility and Sterility*, 21–50. <https://doi.org/10.1016/j.fertnstert.2019.11.030>
 42. Traish, A. M., Melcangi, R. C., Bortolato, M., Garcia-Segura, L. M., & Zitzmann, M. (2015). Adverse effects of 5 α -reductase inhibitors: What do we know, don't know, and need to know? *Reviews in Endocrine and Metabolic Disorders*, 177–198. <https://doi.org/10.1007/s11154-015-9319-y>
 43. Trüeb, R. M., Régnier, A., Dutra Rezende, H., & Gavazzoni Dias, M. F. R. (2019). Post-Finasteride Syndrome: An Induced Delusional Disorder with the Potential of a Mass Psychogenic Illness? *Skin Appendage Disorders*, 320–326. <https://doi.org/10.1159/000497362>
 44. *United States v. Philip Morris USA Inc.* (2006). Public Health Law Center. <https://www.publichealthlawcenter.org/sites/default/files/resources/doj-final-opinion.pdf>
 45. Walf, A. A., Kaurejo, S., & Frye, C. A. (2018). Research Brief: Self-Reports of a Constellation of Persistent Antiandrogenic, Estrogenic, Physical, and Psychological Effects of Finasteride Usage Among Men. *American Journal of Men's Health*, 900–906. <https://doi.org/10.1177/1557988317750989>
 46. Wear, D. (2008). On Outcomes and Humility. *Academic Medicine*, 625–626. <https://doi.org/10.1097/acm.0b013e318178379f>
 47. Zheng, C. J., Han, L. Y., Yap, C. W., Ji, Z. L., Cao, Z. W., & Chen, Y. Z. (2006). Therapeutic Targets: Progress of Their Exploration and Investigation of Their Characteristics. *Pharmacological Reviews*, 259–279. <https://doi.org/10.1124/pr.58.2.4>
-

Research going forward

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/research-going-forward/>

The clinical picture and molecular level understanding of PFS are not where they should be after decades of clear reports of profound suffering and suicides caused by exposure to a cosmetic product. Understanding of a disease, its basic molecular mechanisms, accurate experimental models with predictive value of the disease, and access to technologies for target validation are important for progress towards a therapy (Gashaw et al., 2011) and we urge immediate steps to these ends. Finasteride is a potent endocrine disruptor that targets diverse tissues across the organism. The severity of the symptoms must be considered in parallel with scientific observations on the long-term physiological changes and post-withdrawal effects induced by finasteride and the vast array of physiological processes reliant on the appropriate function of the androgen pathway. Further study of PFS using a precision medicine approach is necessary (Cauci et al., 2017; Coskuner et al., 2019; La Marra, 2010; Traish, 2018). As patients and as patient advocates, we desperately need further molecular level investigation to be undertaken by functional geneticists, epigeneticists, scientists and epidemiologists engaged with both the emerging understanding of androgen signaling and appreciative of the full clinical and pathological picture of PFS. Although there is an increase in reported adverse events (Ali et al., 2015) associated with use of Finasteride 1mg, the numbers of PFS patients are not clearly indicative of the problem when balanced against the millions using this drug. However, considering the multisystemic nature of the persistent health changes and the current void in clinical appreciation and scientific knowledge pertaining to this condition, it is extremely likely the number of young patients experiencing insidious health problems without attribution to a causative antiandrogen to be significant. We strongly advocate for a networked approach with a focus on epigenetic assay as a necessity to move towards mechanistic understanding and ultimately disease modifying treatment. Such an approach has been urged in SBMA and significant steps towards organisation are being achieved (Greensmith et al., 2019; Rinaldi et al., 2015). The advent of adaptive genome and epigenome editing technologies make a treatment feasible following the determination of key mechanistic factors at the molecular level. The suggestion of reversibility of gene dysregulation as a consequence of AR-mediated toxicity in models of other disease states, as discussed, is suggestive of eventual therapeutic possibility.

We recommend far more thorough clinical considerations of PFS patients, particularly severe phenotypes, to be conducted in line with the clinical findings known in androgen-mediated toxicity and the previously reported findings in PFS patients. Primary research must be directed towards the underlying biological differences in the patient cohort. Patients differ greatly in symptomatically affected physiological sites and symptomatic severity, so patient selection based on symptomatic presentation is important in the design of clinical research. We strongly urge that prior 5alpha reductase inhibitor, retinoid or serotonergic drug prescription and use be ascertained in completed incidences of young male suicides in North America and European nations. Currently, completed suicides that the patient themselves or their surviving families explicitly attribute to the physical, sexual and neuropsychological damage induced by

Finasteride are not appropriately attributed to the drug, as suicide is often occurring months or years after cessation when the drug is no longer in their body.

There are many avenues by which to pursue immediate clinical evaluation of PFS patients beyond appropriate basic endocrinological and urological evaluation, and these should account for the specific symptoms of individual patients. Serum creatine-kinase levels may be worthy of assessment during the post-withdrawal crash period or subsequent periods of muscle wasting, as some patients have reported elevated findings. Histological study of affected muscles, including the markedly AR-sensitive perineal muscles, would allow consideration of signs of atrophy and myogenic defects. Area calculation of the bulbocavernosus via ultrasonography has been suggested as a measure of decreased end-organ activity of androgens (Gupta et al., 2017)?, and this could potentially be a low-cost and non-invasive investigation in PFS patients who have experienced atrophic changes. Electromyography to assess abnormalities including signs of perineal muscle denervation may also be worthwhile. MRI protocols including localizer scans, T1-weighted imaging and 2-point Dixon sequences have proven a useful measure of muscle appearance and diffuse involvement in SBMA and could be useful in the phenotype profiling of PFS in patients with broad muscle atrophy. Dual-energy X-ray absorptiometry of bone including lumbar/thoracic spine, femur and sites of complaint, along with serum C-telopeptide testing to assess bone mineral density and trabecular bone health may be worthwhile in patients with bone-related symptomatology and who report structural alteration. Lipid profiling of patient cohorts would additionally provide insight into metabolic dysregulation.

Above all, a far greater focus on molecular level research and basic science is an overdue necessity. Due to low patient numbers, genome wide association study is unlikely to be a practical option, and full genome sequencing of existing PFS patients should be pursued to explore the potential of predisposing factors at the genomic level. Discovery of such genetic differences could eventually be used to screen for risk in young consumers considering use of antiandrogenic products or supplements. Proteomic study may yield insight into the mechanisms of toxicity. Assaying of gene expression data, study of chromatin structure and associated proteins, and methylome analysis of pathologically relevant tissues will advance understanding of deregulated genes as driving factors in this new and novel disease that develops following endocrine disruption. PFS patients are usually in good health prior to use of the associated antiandrogenic substance and can extraordinarily rapidly develop secondary disease states, many of which are associated with advanced age. Advancing the understanding of PFS is therefore likely to yield important mechanistic insights into a diverse array of pathologies. Comparative epigenetic profiling of patients suffering from the disease states following Accutane and SSRI antidepressant use could provide grounds for the wider consideration of the hypothesis regarding a common post-androgen deprivation syndrome and thus a ground-breaking discovery.

Page Bibliography

1. Ali, A. K., Heran, B. S., & Etminan, M. (2015). Persistent Sexual Dysfunction and Suicidal Ideation in Young Men Treated with Low-Dose Finasteride: A Pharmacovigilance Study. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*, 687–695. <https://doi.org/10.1002/phar.1612>
 2. Cauci, S., Chiriaco, G., Cecchin, E., Toffoli, G., Xodo, S., Stinco, G., & Trombetta, C. (2017). Androgen Receptor (AR) Gene (CAG)_n and (GGN)_n Length Polymorphisms and Symptoms in Young Males With Long-Lasting Adverse Effects After Finasteride Use Against Androgenic Alopecia. *Sexual Medicine*, e61–e71. <https://doi.org/10.1016/j.esxm.2016.11.001>
 3. Coskuner, E. R., Ozkan, B., & Culha, M. G. (2019). Sexual Problems of Men With Androgenic Alopecia Treated With 5-Alpha Reductase Inhibitors. *Sexual Medicine Reviews*, 277–282. <https://doi.org/10.1016/j.sxmr.2018.07.003>
 4. Gashaw, I., Ellinghaus, P., Sommer, A., & Asadullah, K. (2011). What makes a good drug target? *Drug Discovery Today*, 1037–1043. <https://doi.org/10.1016/j.drudis.2011.09.007>
 5. Greensmith, L., Pradat, P. F., Sorarù, G., & Pennuto, M. (2019). 241st ENMC international workshop: Towards a European unifying lab for Kennedy's disease. 15–17th February, 2019 Hoofddorp, The Netherlands. *Neuromuscular Disorders*, 716–724. <https://doi.org/10.1016/j.nmd.2019.07.008>
 6. Gupta, N., Carvajal, M., Jurewicz, M., & Gilbert, B. R. (2017). Bulbocavernosus muscle area as a novel marker for hypogonadism. *Asian Journal of Urology*, 3–9. <https://doi.org/10.1016/j.ajur.2016.11.002>
 7. La Marra, F. (2010). *The post-finasteride syndrome in patients with alopecia*. university of Udine.
 8. Rinaldi, C., Malik, B., & Greensmith, L. (2015). Targeted Molecular Therapies for SBMA. *Journal of Molecular Neuroscience*, 335–342. <https://doi.org/10.1007/s12031-015-0676-5>
 9. Traish, A. M. (2018). The Post-finasteride Syndrome: Clinical Manifestation of Drug-Induced Epigenetics Due to Endocrine Disruption. *Current Sexual Health Reports*, 88–103. <https://doi.org/10.1007/s11930-018-0161-6>
-

Conclusion

by phadmin - Monday, March 30, 2020

<https://www.propeciahelp.com/post-androgen-deprivation-syndrome-conclusion/>

Post-Finasteride Syndrome represents a Post-Androgen Deprivation Syndrome following exposure to antiandrogenic endocrine disruptors and without specificity to finasteride. The common pharmacological interruption of steroid signaling and remarkably similar clinical endpoints may imply that a single mechanistic disease state occurs in predisposed consumers following the use of medications including dutasteride, isotretinoin and serotonergic antidepressants. The lasting and profound changes to physiological and neurological health are alarming and the permanence suggests that, in predisposed individuals, epigenetic reorganisation is possible in somatic cells and postmitotic neurons following significant interruption of androgen signaling. The past decade has seen broad appreciation that either excessive or insufficient androgen signaling can prove deleterious to cellular homeostasis and biologic function (Gibson et al., 2018). These mechanistic underpinnings trace back to the influential work of Charles Huggins in the mid-20th century (Huggins, 1965).

We hypothesise a clinically significant AR deregulation is an aberrant manifestation of a conserved mechanism of cellular adaptation to lowered levels of availability or potency of androgenic ligand or interruption of appropriate transactivation of androgen regulated genes. Potential mechanistic factors can be contextualised by the rapidly expanding understanding of the ability of the androgen receptor to affect the basic epigenetic machinery and the structure of chromatin in addition to its essential regulatory functions. Understanding the pathology may provide extremely valuable insight regarding an increasingly apparent androgen-mediated pleiotropy of relevance to a broad spectrum of disease states often associated with the ageing process. Scientific elucidation of predisposing genetic factors will aid in establishing urgent protections for the public. Regulatory level action is necessary and overdue. We ask the medical community to begin efforts towards education regarding this novel condition and to shoulder the clinical responsibility of accurate diagnosis and appropriate follow up of PFS patients.

This document was authored by axolotl and awor, the administrators of propeciahelp.com. We are extremely grateful for your time and consideration. If you are a specialised scientist working in next-generation sequencing, genomics, epigenetics or androgen receptor signaling and are interested in researching this devastating disease, please email us at: contact@propeciahelp.com.

We thank the site volunteer staff for their practical support, scientists who provided us with feedback and support, and the academic institutions that provided access to the resources necessary to complete this work.

axolotl and awor

Page Bibliography

1. Gibson, D. A., Saunders, P. T. K., & McEwan, I. J. (2018). Androgens and androgen receptor: Above and beyond. *Molecular and Cellular Endocrinology*, 1–3.
<https://doi.org/10.1016/j.mce.2018.02.013>
 2. Huggins, C. (1965). Two principles in endocrine therapy of cancers: Hormone deprivation and hormone interference. *Cancer Research*, 25(7), 1163–1167.
-

propeciahelp.com

Post-Finasteride Syndrome info & discussion forum

PDF generated April 11, 2020 at 10:57 AM by Kalin's PDF Creation Station WordPress plugin